

Research & Reviews: Journal of Medicinal & Organic Chemistry

A Broad review on Cancer in the present life cycle

Satish Kumar Voleti*

Department of Biotechnology, Acharya Nagarjuna University, Andhra Pradesh, India.

Review Article

Received: 29/07/2016
Accepted: 09/08/2016
Published: 16/08/2016

*For Correspondence

Satish Kumar Voleti,
Department of Biotechnology,
Acharya Nagarjuna University,
Andhra Pradesh, India.
Tel: 7842371756

E-Mail:
satishy8bt456@gmail.com

INTRODUCTION

Cancer is the irregular development of the cell which influences the digestion system of a life form. The spread of disease in high power is a noteworthy reason for concern internationally. It is creating a disturbing rate independent of age, sex, racial/ethnic gathering, geographic area and tissue attacked. It is set apart by uncontrolled division of cells with the capacity to attack different tissues, either by direct development into the neighboring tissue through intrusion, or by the relocation into the removed locales by metastasis [1]. As per the most recent disease insights, there were 14.1 million new malignancy related passings which is relied upon to ascend by 70% throughout the following two decades with almost 22 million cases. The rate of disease related rate is very nearly 25% higher in men than in ladies [2-5].

Cancer is the outcome of expression of multiple complex interplay of various non-genetic and genetic factors which may act in conjunction, or in succession to initiate or promote carcinogenesis. The non-genetic factors are carcinogens, tobacco, chemicals, radiations and infectious organisms whereas the genetic factors are inherited mutations, hormones, immune conditions and mutations that occur from metabolism; are responsible for cancer development [5-12]. Cancer starts with the activation of oncogenes in the cells which is associated with the inactivation of tumor suppressor genes. However, the exact event of this genetic expression is a matter of debate even after decades of cancer research [12-20].

The growth of the cancer can spend on the cell cycle of the cell in organism. The cell cycle could be a class cells proliferation regulation method and has 4 useful parts: S phase (DNA replication); G2 phase (cells indurate mitosis); M phase (DNA and cellular phase division into two female offspring cells) and G1 phase (cells commit and indurate another spherical of replication). S and M phases are the main and customary processed to all cell cycles for replication of cells. It needs expression of genes in response to growth factors, that induce cell growth from quiescence or maintain ability for cell cycle progression in periods of active proliferation[21-30].

Emphasis upon early detection of malignant cellular growths instead of imaging might permit earlier intervention. Photon emissions from malignant cells even once they represent a small proportion of the traditional organ has been shown to require a technical understanding of the spectral power density profiles which will be expected by Cosic's Molecular Resonance Recognition equation [31-35]. Here we have a tendency to demonstrate by experimentation a more robust detection technique involving specific filters of Photon emissions from cells in culture. Photons from human duct gland malignant cancer cells displayed prominently suppressed spikes of photons inside a slender band (500 nm) however not at 370 nm, 420 nm, 620 nm, 790 nm, or 950 nm increments compared to non-malignant human embryonic urinary organ cells. Given the recent demonstration that malignant cellswill "store" photons inside a selected wavelength once periodic at constant pattern as a yoked flux and re-emit

the photons during this wavelength tens of minutes later, diminishment of power inside specific ten nm increments of visible wavelength spectra might function associate early detection of close at hand malignancy [35-45].

Malignancy undeveloped cells may be separated through a mixture of strategies, including stream cytometry taking into account the statement of particular cell-surface markers, for example, CD133, CD44 and ALDH. The sorting of side populaces of disease cells through Hoechst 33342 color avoidance is a substitute approach [45-60]. Also, late studies have demonstrated that the circle development examine is a similarly effective technique for isolating disease undifferentiated cells from numerous essential tumors or growth cell lines. We and different labs have demonstrated that these self-reestablishing growth undifferentiated cells can be advanced under circle framing conditions[60-70]. At the point when the subsequent thyrospheres are infused orthotopically into the thyroid organs of immunodeficient mice, they produce tumors that nearly take after human thyroid tumors. In past studies we utilized a bioluminescent human thyrosphere model to inspect two patient-determined ATC cell lines: THJ-11T and THJ-16T [71-80]. We found that as few as 100 thyrosphere-inferred single cells were adequate to frame a tumor when orthotopically infused into immunodeficient NOD/SCIDIL2rg-/- mice, and that tumors could be identified with live imaging as ahead of schedule as seven-days after implantation. Conversely, no less than 5×10^5 parental monolayer cells (a 5000-fold increment) were obliged to create a tumor in the same model. This vigorous bioluminescent human thyrosphere model builds up the tumorigenic part of human thyrospheres in advancing ATC [80-90]. Besides, it accepts the malignancy undifferentiated cell model of ATC that as few as 100 thyrosphere cells are adequate to create tumors in mice. Disease cell lines are the model most regularly utilized as a part of growth exploration and their utilization has without a doubt improved our comprehension of malignancy science [90-100].

REFERENCES

1. Bisen PS. Nutritional Therapy as a Potent Alternate to Chemotherapy against Cancer. *J Cancer Sci Ther.* 2016;8: 168-169.
2. Bisen PS, et al . *Biology of oral cancer: Key apoptotic regulators.* CRC Press, Boca Raton, London, New York; 2013.
3. Bisen PS. Cancer Therapy: An Overview. *J Cancer Sci Ther.* 2013; 6: e130
4. Sonnenschein C, et al. *The society of cells: cancer and control of cell proliferation.* Springer Verlag, New York. 1999.
5. Murugan NJ et al. Differentiation of Malignant Compared to Non-Malignant Cells by Their Bio-Photon Emissions May Only Require a Specific Filter around 500 nm. *J Cancer Sci Ther.* 2016;8: 170-171.
6. Bisen PS. Nutritional Therapy as a Potent Alternate to Chemotherapy against Cancer. *J Cancer Sci Ther* 2016;8: 168-169.
7. Khalid A et al. Matrix Metalloproteinases: New Targets in Cancer Therapy. *J Cancer Sci Ther.* 2016;8: 143-153.
8. Nagase H et al. Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc Res.* 2006; 69: 562-573.
9. Visse R et al. Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. *Circ Res.* 2003;92: 827-839.
10. Geleta B et al. N-myc Downstream Regulated Gene (NDRG): Role in Cancer Metastasis Suppression and as Drug Target in Cancer Therapeutics. *J Cancer Sci Ther.* 2016; 8: 154-159
11. Geleta B et al. Cyclic Dependent Kinase (CDK): Role in Cancer Pathogenesis and as Drug Target in Cancer Therapeutics. *J Cancer Sci Ther.* 2016;8: 160-167.

12. Murugan NJ et al. Differentiation of Malignant Compared to Non-Malignant Cells by Their Bio-Photon Emissions May Only Require a Specific Filter around 500 nm. *J Cancer Sci Ther.* 2016;8: 170-171.
13. Brafford P et al. ¹²⁵I is Human Melanoma Depending on the Source. *J Cancer Sci Ther.* 2016;8: 113.
14. Juhasz I et al. Growth and invasion of human melanomas in human skin grafted to immunodeficient mice. *Am J Pathol.* 1993;143: 528-537.
15. Njagi SM et al. In Vitro Antiproliferative Activity of Aqueous Root Bark Extract of *Cassia abbreviata* (Holmes) Brenan. *J Cancer Sci Ther.* 2016;8: 114-121.
16. Ahmad A et al. Kras, Braf, PIK3CA and EGFR Gene Mutations are Associated with Lymph Node Metastasis and Right Sided Colon Carcinoma. *J Cancer Sci Ther.* 2016;8: 122-129
17. Nyamai DW et al. Herbal Management of Benign Prostatic Hyperplasia. *J Cancer Sci Ther.* 2016;8: 130-134.
18. Ahmad F, et al. (2016) CD24 Induces the Activation of β -Catenin in Intestinal Tumorigenesis. *J Cancer Sci Ther;*8: 135-142
19. Barik S. Combination Therapy for Chronic Lymphoid Leukemia. *J Cancer Sci Ther.* 2016;8: 078-079.
20. Maździarz A et al. Benign Metastasizing Leiomyomas of the Lungs: A Case Report. *J Cancer Sci Ther.* 2016;8: 080-083.
21. Stramare R et al. Imaging Features, Differential Diagnosis and Management of Leiomyosarcomas: Case Series and Review of the Literature. *J Cancer Sci Ther.* 2016;8: 084-091.
22. Álvarez-Bañuelos MT et al. Prognostic Factors Associated with Survival in Women with Breast Cancer from Veracruz, Mexico. *J Cancer Sci Ther.* 2016;8: 092-098.
23. Gayatri Devi V et al. Therapeutic Potentials of CD151 shRNA in Targeting Metastasis of Triple Negative Breast Cancer Cell Line MDA-MB-231. *J Cancer Sci Ther.* 2016;8: 104-112.
24. Tot T et al. Radiologically Unifocal Invasive Breast Carcinomas: Large-Section Histopathology Correlate and Impact on Surgical Management. *J Cancer Sci Ther.* 2016; 8: 050-054.
25. Reyad D et al. Hyponatremia and SIADH Frequency in Clinically Euvolemic Patients Receiving Chemotherapy: Prospective Study in Unselected Patients' Cohort. *J Cancer Sci Ther.* 2016;8: 055-058.
26. Ofor O et al. CTCF May Not Directly Regulate ER α mRNA Expression in the ER+ MCF7 Breast Cancer Cell Line. *J Cancer Sci Ther.* 2016;8: 059-065.
27. Garcia-Novoa A et al. Controversies in Axillary Treatment of Breast Cancer Patients and Metastatic Sentinel Lymph Node. *J Cancer Sci Ther.* 2016;8: 066-068.
28. Paul I et al. Chaperones and Glioma Immunotherapy. *J Cancer Sci Ther.* 2016;8: 069-070.

29. Kamphuis GM et al. Storz Professional Image Enhancement System: A New Technique to Improve Endoscopic Bladder Imaging. *J Cancer Sci Ther.* 2016;8: 071-077.
30. Akhenblit PJ et al. Recent Advances in Targeting Tumor Energy Metabolism with Tumor Acidosis as a Biomarker of Drug Efficacy. *J Cancer Sci Ther.* 2016; 8: 020-029.
31. Corrie PG. Cytotoxic chemotherapy: clinical aspects. *Medicine.* 2008; 36: 24–8.
32. Ross JS, et al. Targeted therapies for cancer. *Am J Clin Pathol.* 2004;122: 598–609.
33. Druker BJ. Imatinib alone and in combination for chronic myeloid leukemia *Semin Hematol.* 2003;40: 50-58.
34. Vander Heiden MG. Targeting cancer metabolism: a therapeutic window opens. *Nat Rev Drug Discov.* 2011;10: 671-684.
35. Warburg O. Über den Stoffwechsel der Carcinomzelle. *Naturwissenschaften.* 1924;12: 1131-1137.
36. DeBerardinis RJ et al. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab.* 2008;7: 11-20.
37. Ito A. Cancer Neoantigens: A Promising Source of Immunogens for Cancer Immunotherapy. *J Clin Cell Immunol.* 2015;6:322.
38. Navas-Carrillo D. Novel Findings in Familial Nonmedullary Thyroid Cancer Genetics. *Thyroid Disorders Ther.* 2015;4: 119.
39. Walker AM et al. Evaluation of Arsenic Trioxide Potential for Lung Cancer Treatment: Assessment of Apoptotic Mechanisms and Oxidative Damage. *J Cancer Sci Ther.* 2016;8: 001-009.
40. Chandel SS et al. Evaluation of Role of Concurrent Chemotherapy and Brachytherapy in Locally Advanced Cervical Cancer Patients. *J Cancer Sci Ther.* 2016;8: 010-014.
41. Van den Bergh JMJ et al. Interleukin-15 and Interleukin-15 Receptor α mRNA Engineered Dendritic Cells as Promising Candidates for Dendritic Cell-based Vaccination in Cancer Immunotherapy. *J Cancer Sci Ther.* 2016;8: 015-019
42. Akhenblit PJ et al. Recent Advances in Targeting Tumor Energy Metabolism with Tumor Acidosis as a Biomarker of Drug Efficacy. *J Cancer Sci Ther.* 2016;8: 020-029.
43. Tumor Z et al. Rosmarinic Acid Inhibits Cell Growth and Migration in Head and Neck Squamous Cell Carcinoma Cell Lines by Attenuating Epidermal Growth Factor Receptor Signaling. *J Cancer Sci Ther.* 2015;7: 367-374.
44. Omran AA et al. CD44 and CD44 Variant 6 in Children with Acute Lymphoblastic Leukemia. *J Cancer Sci Ther.* 2015;7: 375-378.
45. Fazioli F et al. Silicate Granules Preconditioned with Human Bone Marrow Mononuclear Cells Improve Osteogenesis in Bone Sarcoma Patients. *J Cancer Sci Ther.* 2015;7: 321-327
46. Wu RL et al. Identification of Differentially Expressed miRNAs in Appendiceal Mucinous Cystadenocarcinoma from Mucinous Cystadenoma. *J Cancer Sci Ther.* 2015;7: 328-335.

47. Ali S et al. Differential Expression of MicroRNAs in Tissues and Plasma Co-exists as a Biomarker for Pancreatic Cancer. *J Cancer Sci Ther.* 2015;7: 336-346.
48. Singh V et al. Development of Novel Anti-Cd20 Monoclonal Antibodies and Modulation in Cd20 Levels on Cell Surface: Looking to Improve Immunotherapy Response. *J Cancer Sci Ther.* 2015;7: 347-358.
49. Hamzawy MA et al. An Additional risk of Lung Cancer from Recurrent Exposure to Ethyl Carbamate (EC) in BALB/C Mice. *J Cancer Sci Ther.* 2015;7: 359-362.
50. Sur D et al. Present Status of Cervical Neoplasia Control and Human Papilloma Virus Epidemiology in India: The Wind is Blowing; Unfolding the Truth. *J Cancer Sci Ther.* 2015;7: 363-366.
51. Jamil K. Biomarkers in Oncological Research. *J Cancer Sci Ther.* 2015; 7: e134.
52. Demir M. Effects of Laughter Therapy on Anxiety, Stress, Depression and Quality of Life in Cancer Patients. *J Cancer Sci Ther.* 2015;7: 272-273.
53. Alvarez-Banuelos MT, et al. Free and DNA Adducted Aflatoxins in Chronic Liver Diseases that Predispose Patients to Hepatocellular Carcinoma in Mexico. *J Cancer Sci Ther.* 2015;7: 274-282.
54. Rehmani N et al. DNA Reactive Activities of Some Endogenous Metabolites and their Putative Role in the Induction of Cancer. *J Cancer Sci Ther.* 2015;7: 283-291.
55. Volovat SR et al. Early and Late Complications after Hepatic Arterial "Port-a-Cath" Implantation in the Treatment of Hepatic Metastasis from Colorectal Cancer. *Journal of Surgery [Jurnalul de chirurgie].* 2015; 10: 277-281.
56. Todosi A et al. Assessment of Tumor Parameters as Factors of Aggressiveness in Colon Cancer. *Journal of Surgery [Jurnalul de chirurgie].* 2015;10: 271-275.
57. Palaghia M et al. Metastatic Colorectal Cancer: Review of Diagnosis and Treatment Options. *Journal of Surgery Jurnalul de chirurgie.* 2015;10:249-256.
58. Olmos J, et al. Apoptosis Comparison Effects Between Synthetic and Natural B-Carotene from Dunaliella salina on MDA-MB-231 Breast Cancer Cells. *J Microb Biochem Technol.* 2015;7:051-056.
59. Slavin S et al. Towards Possible Cure of Cancer by Immunotherapy of Minimal Residual Disease. *J Blood Lymph.* 2015;5:137.
60. Andrea CG et al. Complete Response in Patient with Metastatic Breast Cancer Treated with Metronomic Chemotherapy. *J Blood Lymph.* 2015;5:136.
61. Weiming Xu et al. Targeting Membrane-Bound GRP78 Protein (Arrow) on the GFP-labelled Breast Cancer Cell Surface (Green) by the Quantum Dot-Conjugated Anti-GRP78 ScFv Antibody (Red). *Single Cell Biol.* 2015;4:i101.
62. Zhao X et al. Anti-Cancer Drug Screening Based on a Adipose-Derived Stem Cell/Hepatocyte 3D Printing Technique. *J Stem Cell Res Ther.* 2015;5:273.

63. Chhabra A et al. Cancer Immunotherapy: Targeting Checkpoint Blockade. *Adv Genet Eng.* 2015;3:118.
64. Gajbhiye KR et al. Targeted Brain Delivery of Bioactive Molecules Using Nanocarriers. *J Bioequiv Availab.* 2015;7:112-122.
65. Ruella M et al. The Addition of the BTK inhibitor Ibrutinib to Anti-CD19 Chimeric Antigen Receptor T Cells (CART19) Improves Responses against Mantle Cell Lymphoma. *Clin Cancer Res.* 2016.
66. Fraietta JA et al. Ibrutinib enhances chimeric antigen receptor T-cell engraftment and efficacy in leukemia. *Blood.* 2016;127: 1117-1127.
67. Phillips DC et al. Loss in MCL-1 function sensitizes non-Hodgkin's lymphoma cell lines to the BCL-2-selective inhibitor venetoclax (ABT-199). *Blood Cancer J.* 2015;5: e368.
68. Trent JC et al. Molecular approaches to resolve diagnostic dilemmas: the case of gastrointestinal stromal tumor and leiomyosarcoma. *Future Oncol.* 2007; 3: 629-637.
69. Park MY et al. Preliminary experience using dynamic MRI at 3.0 Tesla for evaluation of soft tissue tumors. *Korean J Radiol.* 2013;14: 102-109.
70. Gemici K et al. Management of patients with retroperitoneal tumors and a review of the literature. *World J Surg Oncol.* 2015; 13: 143.
71. Webb EM et al. Can CT features differentiate between inferior vena cava leiomyosarcomas and primary retroperitoneal masses? *AJR Am J Roentgenol.* 2013;200: 205-209.
72. Huang J et al. Primary intraluminal leiomyosarcoma of the inferior vena cava: value of MRI with contrast-enhanced MR venography in diagnosis and treatment. *Abdom Imaging.* 2011;36: 337-341.
73. Mingoli A et al. International registry of inferior vena cava leiomyosarcoma: analysis of a world series on 218 patients. *Anticancer Res.* 1996;16: 3201-3205.
74. Narata M et al. Primary leiomyosarcoma of the inferior vena cava: case report. *Abdom Imaging.* 2010;35: 481-484.
75. Hemant D et al. Primary leiomyosarcoma of inferior vena cava, a rare entity: Imaging features. *Australas Radiol.* 2001; 45: 448-451.
76. Sostman HD et al. MR imaging and spectroscopy for prognostic evaluation in soft-tissue sarcomas. *Radiology.* 1994;190: 269-275.
77. Illuminati G et al. Outcome of inferior vena cava and noncaval venous leiomyosarcomas. *Surgery.* 2016;159: 613-620.
78. Widmann G et al. State-of-the-art HR-US imaging findings of the most frequent musculoskeletal soft-tissue tumors. *Skeletal Radiol.* 2009; 38: 637-649
79. Beaman FD et al. Superficial soft-tissue masses: analysis, diagnosis, and differential considerations. *Radiographics.* 2007; 27: 509-523.
80. Stramare R et al. Contrast-enhanced ultrasound findings in soft-tissue lesions: preliminary results. *J Ultrasound.* 2013;16: 21-27.

81. Morel M et al. Imaging of the most frequent superficial soft-tissue sarcomas. *Skeletal Radiol.* 2011;40: 271-284.
82. Murase E et al. Uterine leiomyomas: histopathologic features, MR imaging findings, differential diagnosis and treatment. *Radiographics.* 1999;19: 1179-1197.
83. Illuminati G et al. Prosthetic replacement of the infrahepatic inferior vena cava for leiomyosarcoma. *Arch Surg.* 2006; 141: 919-924.
84. Gronchi A et al. Preoperative chemo-radiation therapy for localised retroperitoneal sarcoma: A phase I-II study from the Italian Sarcoma Group. *Eur J Cancer.* 2013;50: 784-792.
85. Bonvalot S et al. Technical considerations in surgery for retroperitoneal sarcomas: position paper from E-Surge, a master class in sarcoma surgery, and EORTC-STBSG. *Ann Surg Oncol.* 2012;19: 2981-2991.
86. Li Z et al. Targeting Six1 by lentivirus-mediated RNA interference inhibits colorectal cancer cell growth and invasion. *Int J Clin Exp Pathol.* 2014; 7: 631-639.
87. McManus MT et al. Gene silencing in mammals by small interfering RNAs. *Nat Rev Genet.* 2002;3: 737-747.
88. Kim DH et al. Strategies for silencing human disease using RNA interference. *Nat Rev Genet.* 2007;8: 173-184.
89. Brummelkamp TR et al. A system for stable expression of short interfering RNAs in mammalian cells. *Science.* 2002;296: 550-553.
90. Sui G et al. A DNA vector-based RNAi technology to suppress gene expression in mammalian cells. *Proc Natl Acad Sci U S A.* 2002;99: 5515-5520.
91. Elbashir SM et al. Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. *Nature.* 2001;411: 494-498.
92. Liu T et al. Targeting CD151 by lentivirus-mediated RNA interference inhibits luminal and basal-like breast cancer cell growth and invasion. *Mol Cell Biochem.* 2015;407: 111-121.
93. Gialeli C, et al. Roles of matrix metalloproteinases in cancer progression and their pharmacological targeting. *FEBS J.* 2011;278: 16-27.
94. Hasegawa M, et al. CD151 dynamics in carcinoma stroma interaction: integrin expression, adhesion strength and proteolytic activity. *Lab Invest.* 2007;87: 882-892.
95. Shiomi T et al. Pericellular activation of proMMP-7 (promatrilysin-1) through interaction with CD151. *Lab Invest.* 2005; 85: 1489-1506.
96. Veeravalli KK et al. MMP-9, uPAR and cathepsin B silencing downregulate integrins in human glioma xenograft cells in vitro and in vivo in nude mice. *PLoS ONE.* 2010;5: e11583.
97. Gustafson-Wagner E et al. The CD9/CD81 tetraspanin complex and tetraspanin CD151 regulate $\alpha\beta 1$ integrin-dependent tumor cell behaviors by overlapping but distinct mechanisms. *PLoS One.* 2013;8: e61834.

98. Shigeta M et al. CD151 regulates epithelial cell-cell adhesion through PKC- and Cdc42-dependent actin cytoskeletal reorganization. *J Cell Biol.* 2003;163: 165-176.
99. Li P, et al. Effects of tetraspanin CD151 inhibition on A549 human lung adenocarcinoma cells. *Mol Med Rep.* 2015;11: 1258- 1265.
100. Mosig RA et al. Application of RNASeq transcriptome analysis: CD151 is an Invasion/Migration target in all stages of epithelial ovarian cancer. *J Ovarian Res.* 2012;5: 4.