Assessment of Hostile to Diarrheal Capability of Moringa Oleifera (Lam.) Clears Out

Chandana E1* and Darion M2

¹Sri Krishnadevaraya University, Anantapur, Andhra Pradesh, India ²American University, USA

Review Article

ABSTRACT

Received: 01/09/2016 Accepted: 21/09/2016 Published: 26/09/2016

*For Correspondence

Chandana E, Sri Krishnadevaraya University, Anantapur, Andhra Pradesh, India

E-mail: chandana.eddula@gmail.com

Keywords: Phytochemicals, Methanol, Intestinal mucosa, Alkaloid, Tannin The target of present study was to assess in vivo hostile to diarrheal capability of *Moringa oleifera* takes off. Test was broke down for quantative estimation of phytochemicals and against diarrheal movement of ethanol concentrate, at 150 and 300 mg/kg body weight (b.w) was examined utilizing castor oil instigated model. Study uncovers that protein was most extreme in leaves (23.35%) trailed by fiber, starch, oil, sugar, alkaloid, tannin, flavanoid and phenolic. At doses of 150 and 300 mg/kg b.w extricate indicated noteworthy (p<0.01) action, when contrasted with control. In course of perception for 4 hour, oral measurement of 300 mg/kg b.w indicated surprising dosage subordinate postponement in onset of extensive loose bowels, diminish in the recurrence of cleansing, weight of wet stools, weight of aggregate stool and rate assurance. Concentrate, along these lines implies the potential clinical impact of concentrate in illness and should be broadened further for movement guided extraction/disconnection of the dynamic substance moiety.

INTRODUCTION

Moringa oleifera Lam. (Moringaceae) is a little to medium-sized tree, bounteously found in all over the fields of India. It is specified as "Shigon" in the "Shushruta Samhita", supporting the proof that development of this tree in India was goes back to a great many a long time. A few sections of the specie were utilized as a part of tribal/customary medication for the infections like injuries, diarrhea, pneumonia, disease, and so forth ^[1-5]. Moringa contains different phytochemicals, some of which are of high premium due to their restorative qualities; specifically this plant is rich in a genuinely one of a kind gathering of glycoside mixes called as glucosinolates and isothiocyanates.

In creating nations, a lion's share of individuals living in country regions solely utilize customary drug in treating a wide range of maladies including loose bowels, which is extremely basic furthermore, repeating sickness in group. Looseness of the bowels is a noteworthy wellbeing issue particularly for youngsters less than 5 years old and up to 17% of tainted kids cease to exist with this infection. Overall dissemination of looseness of the bowels represents more than 5-8 million passings every year in newborn children and youngsters underneath 5 years particularly in creating nations. As per WHO, gauges, around 7.1 million passings were brought on by looseness of the bowels ^[6-15]. It is in this manner critical to recognize and assess accessible normal contrasting options to at present utilized hostile to diarrheal medications, which are not generally free from antagonistic impacts. Writing proposed that a few sections of this species had potential as against diarrheal operator. A study on against diarrheal movement of *Moringa* roots had reported before.

In different examinations against diarrheal action in methanol, fluid and hydroalcoholic concentrate of *M. oleifera* leaves were likewise assessed ^[16-20]. A few studies were additionally done on antibacterial movement of different parts of species, which incorporates examine on coliform microscopic organisms (major causative microorganism for loose bowels), too. In comparable arrangement of activity an endeavor has been made to break down the different phytochemicals present in the leaves and to assess the action potential in ethanol concentrate of *M. oleifera* leaves through castor oil instigated model in test creatures.

Preparation of Extract and Preliminary Phytochemical Screening

Powdered specimen (500 g) was at first macerated for 7 hour with petroleum ether (40-60°C) to evacuate the greasy segment and afterward subjected to liquor (absolute ethanol, 99.99%) as a dissolvable for 7 days (Temp. 27 \pm 2°C) with irregular shaking ^[21-26]. Unrefined concentrate was separated and dried under decreased weight at 40°C. Subjective estimation of sugars, proteins, alkaloids flavanoids, tannins, saponins, steroids, anthraquinones and cyanogenic glycosides were executed according to standard methodology in ethanol concentrate of clears out.

EXPERIMENTAL ANIMALS

Animals (Sprague-Dawley rats) of 125-175 g were chosen and arbitrarily separated into six gatherings (n=6) for screening. Two gatherings for test measurements (150 and 300 mg/kg) of ethanol concentrate, while one each for standard medication and control individually ^[27-34]. Animals were set in pens, bolstered with standard eating routine and water (Temp 27 \pm 2 °C). Before treatment creatures were fasted overnight of sustenance however not water. Conditions were kept up according to creature moral advisory group rules.

CASTOR OIL-INDUCED MODEL

The animals were separated into four gatherings of 6 animals each. The gathering 1 served as the control and got 0.5% CMC suspended in refined water. The following three gatherings got castor oil (Paras Chemicals) in the measurements of 1 ml for every creature p.o. 30 minutes after castor oil organization, bunch 2 and 3 get extricate at measurements of 150 and 300 mg/kg body weight, p.o. also, the gathering 4 get Loperamide (3 mg/kg; p.o.) individually ^[35-41]. Taking after their organization, the animals were set independently in acrylic confines with channel paper, which was changed each hour.

The seriousness of the diarrhoea was surveyed every hour till 4 hour. The aggregate number of defecation (dry and wet stool) and diarrhoeal dung (wet stool) discharged in record time were scored and contrasted and control bunch ^[42-46]. The aggregate score of diarrhoeal dung of control gathering was viewed as that of 100%. The outcomes were communicated in rate of hindrance.

The study supports the nearness of different phytochemicals in *M. oleifera* leaves as appeared in figure 1 and there evaluation uncovers that protein substance was observed to be most noteworthy (23.35%) trailed by fiber, starch, oil, sugar, alkaloid, tannin, flavonoids and phenolic i.e. 23.35, 16.94, 9.86, 9.21, 4.81, 1.8, 1.78, 1.46, 0.65% separately ^[47-60].

This portrays the wholesome significance and financial utilization of leaves separated from consumable organic products, more in provincial and under-fed part of the world. *M. oleifera* leaves when treated with ethanol, yielded 9.75% of concentrate on weight premise ^[61-78]. Subjective estimation of phytochemicals in ethanol concentrate of leaves as compressed in table 1 demonstrates the nearness of sugar, protein, steroid, flavonoids, tannin, alkaloid and glycosides.

The onset of the diarrhea is deferred, after organization of dosages i.e. 52 and 62 min for 150 and 300 mg/kg b.w separately and is noteworthy to that of control, which indicates diarrheal side effect simply after 40 min ^[79-85]. Over the span of perception for 4 hours after castor oil organization, every one of the creatures in tried gatherings which already delivers bountiful looseness of the bowels, indicates diminish in recurrence of cleansing (lessening in no. of wet stool and aggregate no. of stools), weight of wet stools and weight of aggregate stool.

DISCUSSION

The aftereffect of present study would propose that ethanolic concentrate of *M. oleifera* leaves display noteworthy activity against castor oil instigated looseness of the bowels ^[86-90]. Castor oil causes loose bowels because of its dynamic metabolite, ricinoleic corrosive, which fortify peristaltic action in small digestive tract, prompting changes in the electrolytic porousness of the intestinal mucosa and in this manner expands the volume of intestinal substance by keeping the reabsorption of water.

The liberation of ricinoleic corrosive additionally brings about disturbance and aggravation of intestinal mucosa prompting arrival of prostaglandins and autocoids. Loperamide at present is a standout amongst the most adequate and broadly utilized hostile to diarrheal operators and successfully offends the activity of castor oil because of its antimotility and antisecretary property ^[91.95].

Since the alcoholic concentrate effectively restrains the castor oil incited looseness of the bowels, the activity may be by means of against secretary instrument. The huge diminishment in recurrence of crap, number of wet stool, mean stool check, weight of wet stool and weight of aggregate stool connotes the viability of ethanolic concentrate of *Moringa* leaves as a viable against diarrheal operator.

Generally heelers and basic man utilizes alcoholic decoction of different herbs to cure the ailment and this is in reality the motivation to break down the counter diarrheal movement in ethanol remove ^[96-100]. Notwithstanding this it is very much reported that few gathering/classes of phytochemicals have against dysenteric and antidiarrhoeal property i.e. tannins, alkaloids, saponins, flavanoids, sterols/triterpenoids, lessening sugars and anthraquinone glycosides, particularly known for their purgative impact.

Subsequently the nearness of tannin, alkaloid, flavonoids, sugar and anthraquinone might be the basic reason for component. Adjacent to this the potentiating activity may likewise be because of denatured proteins, which structure protein tannates, these buildings of tannin make the intestinal mucosa safer and thusly lessens the emission. Along these lines the general conceivable system of activity might be because of hostile to secretary component.

CONCLUSION

Present examination uncovered that alcoholic concentrate of *M. oleifera* contains pharmacologically dynamic phyto molecule(s) with potential against diarrhoeal properties and can be utilized as non-particular hostile to diarrhoeal specialist. Since the concentrate contains a scope of intensifies, the watched movement might be because of single substance moiety and/or gathering of restoratively dynamic parts like protein, flavonoids, tannin and so forth which may add to fundamental reason for the activity. Consequently, assist broad and explained studies are expected to extricate and detach the bioactive compound (s) for better comprehension of such activities in more exploratory way.

REFERENCES

- 1. Saralaya MG, et al. Antidiarrhoeal Activity of Methanolic Extract of Moringa oleifera Lam Roots in Experimental Animal Models. Int J Pharm Res 2010;2:35-39.
- 2. Goyal BR, et al. Phytopharmacology of Moringa oleifera Lam: An overview. Nat Prod Rad 2007;6:347-353.
- 3. Patel S, et al. Moringa oleifera: A Review of medicinal and Economical importance to the health and nation. Drug Invent Tod 2010;2:339-342.
- 4. Fauci AS, et al. Harrison's Principles of Internal medicine. McGraw Hill Company, New York; 1993;1:236-242.
- 5. Park k. Park's textbook of preventive and social medicine. Banarsidas Bharat Publishes, Jabalpur, India; 2000.
- 6. Hardman JG, Limberd LE. The Pharmacological basis of therapeutics. In Goodman and Gilman's. Mcgraw Hill, New York; 1992, 914-931.
- 7. Lakshminarayana M, et al. Antidiarrhoeal activity of leaf extract of Moringa oleifera in experimentally induced diarrhoea in rats. Int J Phytomed 2011;3:68-74.
- Choudhury S, et al. Antidiarrhoeal Potentiality of Leaf Extracts of Moringa oleifera. British J App Sci Tech 2013;3:1086-1096.
- 9. Rahman MM, et al. Antibacterial activity of leaf juice and extracts of Moringa oleifera Lam. against some human pathogenic bacteria. CMU J Nat Sci 2009;8:219-227.
- 10. Bukar A, et al. Antimicrobial Profile of Moringa Oleifera Lam. Extracts against some Food Borne Microorganism. Bayero J Pure Appl Sci; 2010;3:43-8.
- 11. Talreja T. Screening of Crude Extract of Flavanoids of Moringa oleifera against Bacterial and Fungal Pathogen. J phyto 2010;2:31-33.
- 12. Rahman MM, et al. Control of Coliform Bacteria Detected from Diarrhea associated Patients by extract of Moringa oleifera. Nepal med college J 2010;12:12-19.
- 13. Galvez J, et al. Anti Diarrhoeal Activity of Euphorbia Hirta Extract and Isolation of an Active Flavonoid Constituent. Plant Med 1993;59:333-336.
- 14. Adedapo AA, et al. Safety Evaluation of the Aqueous Extract of the Leaves of Moringa oleifera in Rats. J Med Plants Res 1993;3:586-591.
- 15. Anonymous. The Ayurvedic Pharmacopoeia of India, Government of India, Ministry of Health and Family Welfare, New Delhi; 1989;2:17-8.
- 16. Anonymous. Official Methods of Analysis of Association of official Analytical Chemists, Virginia, US; 1984.

- 17. Bray HC and Thorpe WV. Analysis of Phenolic Compounds of Interest in Metabolism. Meth Biochem Anal 1954;1:27-52.
- 18. Mukherjee PK. Quality Control of Herbal Drugs formulation, an approach to evaluation of herbal drug formulations. Galaxy printers, Business Horizons, New Delhi; 2008.
- 19. Lowry OH, et al. Protein measurement with the Folin phenol reagent. J Biol Chem 1951;193:265.
- 20. Harborne JB. Phytochemical Methods. Chapman and Hall, London; 1973;11-21.
- 21. Kokate CK and Purohit AP. Text book of Pharmacognosy. Nirali Prakashan, India; 2004;1-14.
- 22. Trease G and Evans M. Pharmacopeial and related drugs of biological origin. In: A Textbook of Pharmacognosy. WB Saunders, London; 2001;262-270.
- 23. Vogel HG. Drug discovery and evaluation: Pharmacological Assay, Spinger publication, Germany; 2002.
- 24. Doherty SS. Inhibition of Arachidonic Acid Release Mechanism by which Glucocorticoids Inhibit End toxinInduced Diarrhoea. Brit J Pharm 1981;73:549-554.
- 25. Zaval MA, et al. Antidiarrhoeal activity of Waltheria anorlana, Commelina coelestis and Alternanthera repens. J Ethnopharm 1998;61:41-47.
- 26. Adeyemi OO, et al. Evaluation of Anti Diarrhoeal Effect of Sansevieria liberica Gerome & labroy (Agavaceae) Root Extract. J Ethnopharm 2009;459-463.
- 27. Ammon PJ, et al. Effects of oleic and ricinoleic acid net jejuna water and electrolyte movement. J Clin Invest 1974;53:374-379.
- 28. Watson WC and Gordon R. Studies on digestion absorption and metabolism of castor oil. Biochem. Pharmacol 1962;11:229-236.
- 29. Pierce NF, et al. Effect of Prostaglandin, theophylline and cholera exotoxin upon transmucosal water and electrolyte movement in canine jejunum. Gastroenter 1971;60:22-32.
- 30. Tripathi KD. Essentials of Medical Pharmacology. Jay pee brothers Medical Publishers Ltd, 610-620.
- 31. Irwin RS, et al. Managing Cough as a Defense Mechanism and as a Symptom: A Consensus Panel Report of the American College of Chest Physicians. Chest 1998;114:133S-181S.
- 32. Silva TR, et al. Adhatoda vasica: A Critical Review of Ethanopharmacological and Toxicological Data. Journal of Ethnopharmacology 2000;72:1-20.
- 33. Numazaki K, et al. Effect of Glycyrrhizin in Children with Liver Dysfunction Associated with Cytomegalovirus Infection. The Tohoku Journal of Experimental Medicine 1994;172:147-153.
- 34. Tsukiyama R. Antibacterial Activity of Licochalcone A against Spore-Forming Bacteria. Antimicrobial Agents and Chemotherapy 2002;46:1226-1230.
- 35. CSIR, The Wealth of India: A Dictionary of Indian Raw Materials and Industrial Products. Council of Scientific and Industrial Research, New Delhi; 1989;8:96-99.
- 36. Kasirajan B, et al. A Database for Medicinal Plant Use in Treatment of Asthma. Bioinformation 2007;2:105-106.
- 37. Hernandez T, et al. Antimicrobial Activity of the Essential Oil Extracts of Cordia curassavica. Journal of Ethnopharmacology 2007;111:137-141.
- 38. Sabira B, et al. Chemical constituents of Cordia latifolia and Their Nematicidal Activity. Chem Biodiver 2011;8:850-861.
- 39. Jiang JG, et al. Comparison of the Sedative and Hypnotic Effects of Flavonoids, Saponins, and Polysaccharides Extracted from Semen Zizyphus jujuba. Natural Product Research 2007;21:310-320.
- 40. Mahajan RT and Chopda MZ. Phytopharmacology of Zizyphus jujuba Mill—A Plant Review. Pharmacog Rev 2009;3:320-329.
- 41. Kumar N, et al. Onosma L.: A Review of Phytochemistry and Ethnopharmacology. Pharmacog Rev 2013;7:140-151.
- 42. Srivastava S, et al. HPLC Determination of Vasicine and Vasicinone in Adhatoda vasica with Photo Diode Array Detection. J Liquid Chromat Rel Technol 2001;24:153-159.
- 43. Modi J, et al. A Detail Phyto-Chemical Evaluation of Herbo-Mineral Formulation Used in Respiratory Diseases. J Pharmacog Phytochem 2014;2:36-42.

- 44. Desai L, et al. Prospective Process Validation of Polyherbal Cough Syrup Formulation. J Advan Pharma Tech Res 2012;2:225-231.
- 45. Kirtikar KR and Basu BD. Indian Medicinal Plants. Popular Prakashan, Mumbai; 2002;2:94-97.
- 46. Khare CP. Indian Medicinal Plants, Springer Private Limited, New York; 2007;151-152.
- 47. Nandkarni KM. Indian Materia Medica, Popular Parakashan, Mumbai; 2002,3:333-334.
- 48. Khandelwal KR. Practical Pharmacognosy, Techniques and Experiments, Nirali Prakashan, Pune 2004; 12:30-44.
- 49. Trease GE and Evans WC. Pharmacognosy, Harcourt brace & Co. Asia, Pvt. Ltd., W.B. Saunders Company Ltd., 2002;15:542-543.
- 50. Kokoski CJ, Kokoski RJ, Slama FJ. Fluorescence of the powdered vegetable drugs under UV radiation. J Am Pharm Assoc 1958;715-717.
- 51. Daba M. Miracle Tree: A Review on Multi-purposes of Moringa oleifera and Its Implication for Climate Change Mitigation. J Earth Sci Clim Change 2012;7:366.
- 52. Kotikal YK and Math M. Insect and Non-Insect Pests Associated with Drumstick, Moringa oleifera (Lamk.). Entomol Ornithol Herpetol 2016;5:180.
- 53. Bahr HI and Farouk SM. Comparative Ameliorative Effect of Basil Oil and *Moringa oleifera* on Lornoxicam-Mediated Histological and Biochemical Alterations in Albino Rat Liver. J Veterinar Sci Technol 2016;7:309.
- 54. Sasikala S and Muthuraman G. A Laboratory Study for the Treatment of Turbidity and Total Hardness Bearing Synthetic Wastewater/Ground Water Using *Moringa oleifera*. Ind Chem 2016;2:112.
- 55. Sasikala S and Muthuraman G. Chromium(VI) Removal Using Biosorbents Derived from *Moringa oleifera*. Ind Chem Open Access 2015;1:105.
- 56. Abdulkadir IS, et al. Phytochemical Screening and Antimicrobial Activities of Ethanolic Extracts of Moringa oleifera Lam on Isolate of Some Pathogens. J App Pharm 2015;7:203.
- 57. Ogundele VA and Fadeyi OE. Isolation, Characterization and Derivatization of Some Bioactive Components in Moringa Oleifera Leaves. Nat Prod Chem Res 2015;3:189.
- 58. Taha NR, et al. Effect of Moringa oleifera Leaves on Diclofenac Sodium Induced Hepatic Injury in Albino Rats: Ultrastructural and Immunohistochemical Studies. J Cytol Histol 2015;6:315.
- 59. Zaroual A, et al. Electrochemical Study of the Capacity of Moringa oleifera to Chelate Heavy Metals. Pharm Anal Acta 2014;5:310.
- 60. Ozovehe BN. Growth Performance, Haematological Indices and Some Biochemical Enzymes of Juveniles Clarias gariepinus (Burchell 1822) Fed Varying Levels of Moringa oleifera Leaf Meal Diet. J Aquac Res Development 2013;4:166.
- 61. Isitua CC and Ibeh IN. Toxicological Assessment of Aqueous Extract of Moringa oleifera and Caulis Bambusae Leaves in Rabbits. J Clinic Toxicol 2013;S12:003.
- 62. Pandey A. Moringa Oleifera Lam. (Sahijan)- A Plant with a Plethora of Diverse Therapeutic Benefits: An Updated Retrospection. Med Arom Plan 2012;1:101.
- 63. Nguyen HN, et al. Modeling of Moringa Oleifera Oil Solubility in Supercritical Carbon Dioxide. J Chem Eng Process Technol 2011;2:114.
- 64. Tsabang N, et al. Herbal Medicine and Treatment of Diabetes in Africa: Case Study in Cameroon. Diabetes Case Rep 2016;1:112.
- 65. Nole T, et al. Ethnomedical and Ethnopharmacological Study of Plants Used For Potential Treatments of Diabetes and Arterial Hypertension by Indigenous People in Three Phytogeographic Regions of Cameroon. Diabetes Case Rep 2016;1:110.
- 66. Panda T, et al. A Folk Knowledge on Medicinal Plants Used for the Treatment of Skin Diseases in Bhadrak District of Odisha, India. Med Aromat Plants 2016;5:262.
- 67. Eldahshan OA, et al. Medicinal Plants and Addiction Treatment. Med Aromat Plants 2016;5:260.
- 68. Karima N, et al. Storage Biologically Active Substances by Convection Drying Food and Medicinal Plants. J Food Process Technol 2016;7:599.
- 69. Maiti R, et al. Nutrient Profile of Native Woody Species and Medicinal Plants in Northeastern Mexico: A Synthesis. J Bioprocess Biotech 2016;6:283.

- 70. Jerezano Alberto VD, et al. Some Traditional Medicinal Plants of North Region from Puebla, Mexico: Uses and Potential Pharmacological Activity of Rumex spp. Nat Prod Chem Res 2016;4:223.
- 71. Getasetegn M and Tefera Y. Biological Activities and Valuable Compounds from Five Medicinal Plants. Nat Prod Chem Res 2016;4:220.
- 72. Gupta A, et al. Extraction of Proteases from Medicinal Plants and their Potential as Anti-Viral Targets. J Biotechnol Biomater 2016;6:228.
- 73. Norkulova K, et al. Research on Aerodynamics Separator for Medicinal Plants. J Food Process Technol 2016;7:586.
- 74. Alothyqic N, et al. In Vitro Antibacterial Activity of four Saudi Medicinal Plants. J Microb Biochem Technol 2016;8:083-089.
- 75. Kumar M. Rural Communities on Ethno Medicinal Plants, Uses and their Conservation. Med Aromat Plants 2016;S3:e003.
- 76. Opinde HR, et al. Antimicrobial Evaluation of Crude Methanolic Leaf Extracts from Selected Medicinal Plants Against *Escherichia coli*. J Bacteriol Parasitol 2016;7:272.
- 77. Tiwari SS. Medicinal Plants-Perspectives and Needs. J Pharmacogn Nat Prod 2016;2:e105.
- 78. Sathelly K, et al. Establishment of Efficient Regeneration System from Leaf Discs in Long Pepper an Important Medicinal Plant (*Piper Iongum* L.). Med Aromat Plants 2016;5:248.
- 79. Bouyahya A, et al. Determination of Phenol Content and Antibacterial Activity of Five Medicinal Plants Ethanolic Extracts from North-West of Morocco. J Plant Pathol Microbiol 2016;7:342.
- 80. Petropoulos AS. Wild Edible Medicinal Plants of the Mediterranean Basin. Med Aromat Plants 2016;5:e173.
- 81. Rachuonyo HO, et al. Combined Effect of Crude Leaf Extracts of Selected Medicinal Plants against Selected Enteric Bacterial Pathogens and Candida albicans. J Antimicro 2016;2:110.
- 82. Khojasteh SMB, et al. A Review on Medicinal Plants Used for Improvement of Spermatogenesis. Biol Med (Aligarh) 2016;8:292.
- 83. Rachuonyo HO, et al. Antimicrobial Potency of Methanolic Leaf Extracts from Selected Medicinal Plants against *Staphylococcus aureus*. J Med Microb Diagn 2016;5:219.
- 84. Tsabang N, et al. Comparative Study of Epidemiological and Anthropological Aspects of Diabetes and Hypertension in Cameroon. Forest Res 2016;5:165.
- 85. Ayuka F and Barnett R. Place Effects on Alcohol Consumption: A Literature Review. J Addict Res Ther 2015;6:207.
- 86. Baeshen MN, et al. Therapeutic Potential of the Folkloric Medicinal Plant *Rhazya stricta*. Biol Syst Open Access 2015;5:151.
- 87. Arika WM, et al. Mineral Elements Content of Selected Kenyan Antidiabetic Medicinal Plants. Adv Tech Biol Med 2016;4:160.
- Hamza AA, et al. α-Glucosidase Inhibitory Activity of Common Traditional Medicinal Plants Used for Diabetes Mellitus. J Develop Drugs 2015;4:144.
- 89. Amabye TG and Shalkh TM. Phytochemical Screening and Evaluation of Antibacterial Activity of Ruta graveolens L. A Medicinal Plant Grown around Mekelle, Tigray, Ethiopia. Nat Prod Chem Res 2015;3:195.
- 90. Hammadi D, et al. Ethnomedicinal Survey of Medicinal Plants Used in the Western Region of Algeria. Med Aromat Plants 2015;5:221.
- 91. Hassan W, et al. Metallic Content of One Hundred Medicinal Plants. J Nutr Disorders Ther 2015;5:177.
- 92. Shivakumar Singh P, et al. Documentation of Folkloric Knowledge on Medicinal Plants Used in the Treatment of Mouth Ulcers in Kodangal Mandal, Mahabubnagar District, Telangana, India. J Bioanal Biomed 2015;07:174-179.
- 93. Garba S, et al. Antidiarrhoeal Activities of Some Medicinal Plants. Med chem 2015;S2:001.
- 94. Mittal N, et al. High Levels of Diversity in the Phytochemistry, Ploidy and Genetics of the Medicinal Plant *Acorus calamus L*. Med Aromat Plants 2015;S1:002.
- 95. Dawood M and Efferth T. Medicinal Plants and DNA Methylation of Cancer. Med Aromat Plants 2015;4:e161.

- 96. Pagadala VK, et al. Significance of Traditional Medicinal Plants used for Treatment of Rabies at Ambo Town. Med Aromat Plants 2015;4:207.
- 97. Kala CP. Medicinal Plants in Active Trade at Haridwar City of Uttarakhand State in India. Med Aromat Plants 2015;4:204.
- 98. Vunnava A and Motlakunta HB. Anthelminthic and Antibacterial Activity of Various Indians Medicinal Plants. Med Aromat Plants 2014;3:167.
- 99. Ghosh S, et al. Commentary on Therapeutic Potential of Gnidia glauca: A Novel Medicinal Plant. Med chem 2015;5:351-353.
- 100. Tsabang N, et al. Treatment of Diabetes and/or Hypertension Using Medicinal Plants in Cameroon. Med Aromat Plants 2015;S2:003.