

In Vitro Flowering and Rapid Propagation of Physalis Minima Linn. - A Medicinal Plant

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ABSTRACT: *In vitro* flowering and an efficient micropropagation protocol was developed for the annual medicinal herb *Physalis minima* (Solanaceae) by the *in vitro* culture of nodal segments of mature plant. Murashige and Skoog's (MS) medium, 3% sucrose and 0.8% agar supplemented with 4.0mg/l BAP and 0.25mg/l 2,4 - D and 1.0mg/l KIN and 0.25mg/l NAA were found to develop fully functional flowers. MS medium supplemented with 2.0mg/l BAP and 0.25mg/l IAA induced an average of three shoots per explants. Full strength MS medium with 2.0mg/l IBA exhibited the best in *in vitro* rooting. Seventy percent of the rooted shoots survived when transferred to green house and next to the field condition.

KEYWORDS: *In vitro* flowering, micropropagation, medicinal plant, *Physalis minima*, Solanaceae.

I. INTRODUCTION

Physalis minima Linn. is one of the important medicinal plant species belonging to the family Solanaceae. The plants of *Physalis minima* Linn. are bitter, appetizing, tonic, diuretic, laxative, and useful in inflammations, enlargement of the spleen and abdominal troubles. The fruit is considered as a tonic, diuretic and purgative. Extracts from leaves have antimicrobial activity. The juice of the leaves, mixed with mustard oil and water, has been used as a remedy for earache. The Malays apply the leaves which have been smeared with oil and heated, to ulcers wounds and pustules. The leaves are also used to relieve head-ache. A paste of the leaves and stem is used to treat dizziness.

Though the plant has immense medicinal value it is gradually declining from the nature due to over exploitation and environmental pollution [2]. In recent years, numerous studies on *in vitro* propagation of different plant species have shown that this technique may be a solution for rapid propagation of selected plant species. The present paper reports an efficient micropropagation system for generating a large number of plants and *in vitro* flowering in *Physalis minima*.

II. MATERIALS AND METHODOLOGY

A. Plant Materials

The plants were collected from Kerala (mainly, Palakkad and Thrissur districts). From the field grown plants, the young nodal region were excised out and used for micropropagation and *in vitro* flowering. The collected explants were washed thoroughly in running tap water for 30'. Then the explants were rinsed with 1% savlon solution containing 6-8 drops of tween 20 for 15' and again washed with double distilled water to remove the traces of detergent solution. Again, inside the laminar air flow chamber rinse once with autoclaved water. Then the explants washed with 70% alcohol for 30 seconds. Next, the explants rinsed with 0.05% mercuric chloride solution for 5' and again washed with sterile distilled water for 6-8 times. Then the explants were placed in sterile petri plates for inoculation. The sterilized nodal region was used as explant source for *in vitro* culturing.

B. Culture medium and Conditions

Basal medium used for initial set of experiments for shoot proliferation. The MS medium (Murashige, T and Skoog, F, 1962) consists of macronutrients, micronutrients, iron source and vitamins supplemented with sucrose (3%) as a carbon source and agar (1%) as a solidifying agent. The pH of the medium was adjusted to 5.8 before autoclaving at a

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pressure of 15lbs. The basal medium was supplemented with various cytokinins [BAP(6 – Benzyl amino purine) and KIN (kinetin)](Wang, S; Tan, L and Chen, F, 2001) and auxins [NAA(α – Naphthalene acetic acid),IAA (Indole – 3 – acetic acid) , IBA (Indole – 3 – butyric acid) and 2,4 – D (2,4 – Dichloro phenoxy acetic acid)] at different concentrations and combinations(Nitsch and Nitsch,J,P, 1967; Sahoo, Y and Chand,P,K,1998 ; Thiruvengadam,M and Narayansamipillai, J, 2000). The cultures were incubated at $25 \pm 2^{\circ}\text{C}$ with 16 hours photoperiod.

C.Rooting, Acclimatization and Field Transfer

The induction of roots was carried out on full strength MS solid medium supplemented with auxin (IBA and NAA) at different concentrations (Vadawale *et al*, 2006). Healthy shoots with well developed roots were transferred to small pots containing vermicompost at maintained humidity (~ 90%) through misting device inside a greenhouse.

Data were collected on number of shoots, shoot length, number of roots, root morphology, number of flower buds and developed flower.

III.RESULTS AND DISCUSSIONS

The explants cultured on MS medium without supplement of growth regulator did not grow at all. MS medium supplemented with different concentrations of BAP in combination with IAA resulted in micropropagation (Table - I). Nodal explants induced an average of three shoots on MS basal medium supplemented with BAP and IAA (Table – I; Fig – Ia).T he maximum percentage of shoot induction was in BAP (2.0mg/l) and IAA (0.25mg/l).

Table – I

Efficacy of MS medium fortified with different growth regulators on micropropagation after 4 weeks of culture.

Treatment	% response	No. of shoots (Mean \pm S.D)
Growth regulator free medium	-	-
BAP (1.0mg/l) + IAA(0.25mg/l)	65	2.9 \pm 1.45
BAP (2.0mg/l) + IAA(0.25mg/l)	80	4.4 \pm 1.36
BAP (3.0mg/l) + IAA(0.25mg/l)	40	3.4 \pm 0.92
BAP (4.0mg/l) + IAA(0.25mg/l)	60	3.2 \pm 1.33
BAP (5.0mg/l) + IAA(0.25mg/l)	60	3.1 \pm 1.58

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(a) Multiple shoots on MS + 2.0mg/l BAP + 0.25mg/l IAA after 4 weeks,



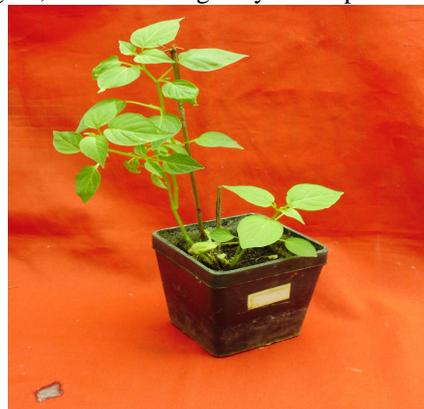
(b) *In vitro* flower bud induction,



(c) Explant cultured on MS + 4.0mg/l BAP + 0.25mg/l 2, 4 - D showing fully developed flower.



(d) Culture showing multiplication, rooting and flower buds



(e) Hardened regenerated plant maintained in greenhouse.

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In vitro flowering

The fully developed shoots from the nodal explants isolated and transferred to fresh MS medium having different concentration of cytokinin (BAP or KIN) in combination with auxin (2,4 – D or NAA) started developing flower bud at the nodal region.

Effect of plant growth regulators on flower induction

The effects of cytokinin (BAP or KIN) and the auxin (2, 4 – D or NAA) on flower induction were examined (Table - II). MS medium containing BAP (4.0mg/l) and 2,4 – D (0.25mg/l); KIN (1.0mg/l) and NAA (0.25mg/l) resulted in uniform flowering with 2 – 3 flowers per plant (Fig. Ib). The flower buds induced on the MS medium supplemented with different concentrations of BAP and 2,4 – D developed into functional flower (Fig.Ic), while those flower buds induced on MS medium supplemented with different concentrations of KIN and NAA developed flowers except 4.0mg/l KIN + 0.25mg/l NAA and 5.0mg/l KIN + 0.25mg/l NAA(Sri Rama Murthy *et al*, 2006).

Table - II

Effect of plant growth regulators on *in vitro* flowering of *Physalis minima* in MS medium after 4 weeks of culture.

Treatment	% of explants producing flower buds	No. of fully developed flower per culture (Mean ± S.D)
MS plain	0	0
BAP(1.0mg/l) + 2,4 – D(0.25mg/l)	10	0.2 ±0.40
BAP(2.0mg/l) + 2,4 – D(0.25mg/l)	10	0.3 ±0.46
BAP(3.0mg/l) + 2,4 – D(0.25mg/l)	20	0.9 ± 0.70
BAP(4.0mg/l) + 2,4 – D(0.25mg/l)	60	1.1 ± 1.04
BAP(5.0mg/l) + 2,4 – D(0.25mg/l)	40	0.8 ± 0.98
KIN (1.0mg/l) + NAA(0.25mg/l)	60	1.0 ± 0.77
KIN (2.0mg/l) + NAA(0.25mg/l)	20	0.8 ± 0.98
KIN (3.0mg/l) + NAA(0.25mg/l)	10	0.8 ± 0.98
KIN (4.0mg/l) + NAA(0.25mg/l)	0	0
KIN (5.0mg/l) + NAA(0.25mg/l)	0	0

Effect of auxin on *in vitro* rooting

IBA was found to be more suitable for root induction compared with NAA (Table – III). When shoots were transferred to the rooting medium supplemented with NAA, stout and shorter roots were formed. The regenerated

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shoots, rooted best on MS medium containing 2.0mg/l IBA (Fig. Id). The roots were thin and with lots of root hairs in the medium with IBA. After three weeks of incubation in the medium with IBA, each shoot developed an average of 5 roots with an average length of 5cm. Slow degradation of IBA facilitates its localization near the site of application and thus, it functions better in inducing roots.

Rooted shoots were transferred directly to small pots filled with vermicompost and moss (1:1). Of the plantlets transferred to soil, 70% survived and exhibited morphological characters similar to those of the field grown plants (Fig. Ie).

Table - III

Effect of auxin on root induction from *in vitro* raised shoots of *Physalis minima* after 4 weeks of culture.

Treatment	No. of roots/shoot (Mean \pm S.D)	Root length(cm) (Mean \pm S.D)	Root morphology
IBA(1.0mg/l)	5.1 \pm 2.02	4.1 \pm 2.21	Thin, Fragile
IBA(2.0mg/l)	6.7 \pm 2.83	6.8 \pm 2.04	Long, Sturdy
IBA(3.0mg/l)	4.7 \pm 2.37	3.6 \pm 1.63	Long, Sturdy
IBA(4.0mg/l)	4.9 \pm 1.92	4.3 \pm 1.68	Long, Sturdy
IBA(5.0mg/l)	3.8 \pm 1.66	4.3 \pm 1.85	Long, Sturdy
NAA(1.0mg/l)	2.9 \pm 1.37	2.9 \pm 1.30	Short, Thick
NAA(2.0mg/l)	2.4 \pm 1.02	2.1 \pm 0.70	Short, Thick
NAA(3.0mg/l)	2.9 \pm 1.58	2.3 \pm 0.90	Short, Thick
NAA(4.0mg/l)	2.9 \pm 1.45	2.8 \pm 0.98	Short, Thick
NAA(5.0mg/l)	3.3 \pm 2.00	1.9 \pm 0.70	Short, Thick

IV.CONCLUSION

A simple and efficient method has been developed for inducing the *in vitro* flowering of *Physalis minima*. In our study, increased *in vitro* flowering were concentration of cytokinin (BAP or KIN) with less concentration of auxin (2, 4 – D or NAA).The protocol also described for the rapid propagation of this medicinal plant.

REFERENCES

- [1]Abri,A .L and Van Staden ,J. Micropropagation of the endangered *Aloe polyphylla*. Plant Growth Regul., 33: 19 – 23,2001.
[2]Farhana Afros, Sayeed Hassan,A.K.M, Laila Shamroze Bari, Rebeka Sultana, Nadira Begam, Miskat Ara Akter Jahan and Rahima Khatun. *In vitro* shoot proliferation and plant regeneration of *Physalis minima* L. – A perrinial medicinal herb. Bangladesh J. Sci. Ind.Res, 44(4):453 -456,2009.
[3]Murashige, T and Skoog, F. A revised medium for rapid growth and bioassay with tobacco tissue cultures. *Physiol. plant*, 15:473-97,1962.

International Journal of Innovative Research in Science, Engineering and Technology

(An ISO 3297: 2007 Certified Organization)

Vol. 4, Issue 1, January 2015

- [4]Nitsch and Nitsch,J.P. The induction flowering *in vitro* in stem segments of *Plumbago indica* II. The production of reproductive buds. *Planta*, 72: 371 – 84,1967.
- [5]Sahoo, Y and Chand,P,K.Micropropagation of *Vitex negundo L.*, a woody aromatic medicinal shrub,through high frequency axillary shoot proliferation. *Plant Cell Rep.*, 18: 301 – 07,1998.
- [6]Sreekumar, S; Seeni, S and Pushpangadan, P.Micropropagation of *Hemidesmus indicus* for cultivation and production of 2 – hydroxyl – 4 – methoxy benzaldehyde. *Plant Cell Tissue Organ Cult.*,62: 211 – 18,2000.
- [7]Sri Rama Murthy,K; Raghu Ramulu,D; Chandrasekhara Rao,J; Emmanuel,S; and Pullaiah, T. *In vitro* flowering of *Spathoglottis plicata* BL(Orchidaceae).*Phytomorphology*, 56(3&4):117-20,2006.
- [8]Thiruvengadam,M and Narayansampillai, J. Mass propagation of *Vitex negundo L. in vitro*. *J. Plant Biotechnol.*, 2: 151 – 55,2000.
- [9]Vadawale, A,V; Meht- bhatt,P and Dave, A,M, . Rapid *in vitro* propagation of Ashwagandha (*Withania somnifera*) through axillary bud multiplication and indirect organogenesis. *Phytomorphology*, 54: 59 – 64,2004.
- [10]Vadawale, A.V; Barve, D.M; and Dave, A.M, . *In vitro* flowering and rapid propagation of *Vitex negundo L.* - A medicinal plant. *Indian J. Biotechnol.*, 5:112-16,2006.
- [11]Wang, S; Tan, L and Chen, F. *In vitro* flowering of bitter melon. *Plant Cell Rep.*, 20: 393 – 97,2001.