

# Isolation of Bacteriocin from Lactobacillus Species and its Application as a Preservative in Dairy Products

Shivani Kashyap\*

Department of Biotechnology, Meerut Institute of Engineering & Technology, Meerut, Uttar Pradesh

## Review Article

Received: 29/08/2016

Revised: 06/09/2016

Accepted: 12/09/2016

### \*For Correspondence

Shivani Kashyap, Department of Biotechnology, Meerut Institute of Engineering & Technology, Meerut, Uttar Pradesh, India, Tel: 8004774540

**Keywords:** Bacteriocin; Dairy products; Chromatography; Microorganism; Antimicrobial metabolites

### E-mail:

shivani Kashyap5661149@yahoo.com

## ABSTRACT

The supernatant fluid tested inhibitory assignment no longer most powerful in opposition to some lactic acid microorganism but additionally, in opposition to some pathogenic and meals-spoilage species, at the side of Clostridium, Listeria and Enterococcus. It used to be as soon as purified to homogeneity by the use of a single four-step process: a crude supernatant fluid purchased from early stationary-part subculture in MRS medium was once as soon as subjected to ammonium sulphate fractionation, CM-Sephadex cation-exchange chromatography, Phenyl-Sepharose hydrophobic chromatography and reverse-phase HPLC chromatography. The bacteriocin used to be produced constitutively at some factor of exponential development. It used to be as soon as bactericidal to sensitive cells and the bactericidal effect used to be now not produced with the help of cell lysis. The amino acid composition of the bacteriocin was once determined and no modified amino acid was once as soon as located among the residues recognized.

## INTRODUCTION

Lactic acid microorganism (LAB) are Gram-optimistic, non-spore forming, catalase-horrible microorganism which are devoid of cytochromes and are of non-aerobic nevertheless are aero-tolerant, fastidious, acid tolerant and strictly fermentative; lactic acid is the important end-manufactured from sugar fermentation. Lactic acid microorganism (LAB) includes a range of bacterial customary within the Phylum fomicutes. The general Enterococcus, Lactobacillus, Lactococcus, Leuconostoc, Milissococcus, Oenococcus, Pediococcus, Streptococcus, Tetragenococcus, Vagococcus and Weissella are famous as lactic acid micro-organism. They may be mostly normally essentially the most generally used bacteria as starter cultures for the trade processing of fermented dairy, meat, vegetable and cereal merchandise. Reduction of pH and conversion of sugars to healthful acids is the principal maintaining moves that these microorganism furnish to fermented meals [1-10].

These ordinary isolates of lactic acid microorganism from spontaneous fermentations can be used as special starter cultures or as adjunct traces, after phenotypic and genotypic characterization, they most commonly signify a potential supply of in general new antimicrobial metabolites. Additionally, the applying of lactic acid microorganism and their antimicrobial metabolites within the prevention of food spoilage and the extension of the shelf lifetime of meals that's equipped to eat, modern-day-tasting, nutrient and diet rich, minimally processed and bio preserved are the foremost challenges for the present meals organization [10-20]. The utilization of bacteriocin-producing lactic acid bacteria as defending traces or bacteriocins in variety of purified or centered compounds as bio-preservatives to manage undesirable microorganism remains a predominant core of recognition of researches involving ingredients defend and satisfactory [21-30].

The antimicrobial undertaking of starter cultures and probiotic microorganism has been attributed to the production of metabolites and same to natural and organic acids (lactic and acetic acid), hydrogen peroxide,

ethanol, diacetyl, acetaldehyde, other low molecular mass compounds with antimicrobial recreation and bacteriocins [31-40].

### REVIEW OF LITERATURE

First of all, lots of the great development in bacteriocin be trained stemmed from investigations of the colicins, the prototype bacteriocins produced via slightly countless members of the adored ones Enterobacteriaceae, and this resulted in large expertise of the genetic groundwork, self-discipline constitution, mode of formation, and killing action of those molecules. Nonetheless, there has now been an potent quantity of be trained assignment based upon the bacteriocin-like ambitions of Gram-constructive microorganism, notably lactic acid microorganism (LAB) [41-50].

The important clear documentation of the personality of an antibiotic agent produced by way of E.Coli was once provided via utilizing making use of Gratia, who validated in 1925 that pressure V (virulent in experimental infections), produced in liquid media, a dialyzable and warmness-steady substance (later referred to as colicin V) that inhibited the development of E. Coli even with a immoderate dilution. Later a series of individual colicins had been discovered over an interval of time [51-53].

The extra natural time period "bacteriocin" used to be coined by way of Jacob et al. Bacteriocins had been principally outlined as protein inhibitors or 'antibiotics' of the colicin variety, i.e., molecules characterized by means of lethality after biosynthesis, predominant intraspecies killing venture, and adsorption to distinctive receptors on the dermis of bacteriocin sensitive cells [54].

An analysis of the bacteriocins of Gram-positive microorganism opened with the comment that a lot of the definitive investigations inside the strength of mind of bacteriocins had headquartered on these of Gram-terrible microorganism nonetheless expected in be trained emphasis on bacteriocins of Gram-optimistic lactic acid microorganism [55-60]. Apparently a lot of the renewed curiosity in these add-ons is an instantaneous response to the perceived expertise beneficial application of these shops, every for the renovation of meals or the prevention and therapy of bacterial infections.

#### ***Components protection***

Despite the fact that nisin is the one bacteriocin in america licensed as a compatible away meals additive, there could also be an quality deal of curiosity in exclusive bacteriocins which have equal residences and show off colossal spectrum inhibitory activity. Bacteriocins produced via fermentation can be purified and brought to foods as pure chemical components to inhibit meals pathogens and spoilage organisms simplest after obtaining approval as an immediate constituents additive with the support of the FDA [61-63]. Many bacteriocins can resist excessive temperature utilized in meals processing and might keep clever over a large pH type. Bacteriocins can also be digested through many enzymes inside the human gastrointestinal tract equal to exceptional proteins inside the healthful eating regimen and not emerge as an trouble for worthy gut microflora. Bacteriocins are non-poisonous, odorless, colorless, and tasteless.

#### ***Bactericidal action***

The low-molecular weight bacteriocins of Gram-confident microorganism traditionally appear to be membrane lively. The lantibiotic subgroup of bacteriocins tends to fluctuate from the reverse companies within the voltage dependence of their membrane insertion. Poration complexes had been proposed to be fashioned between one, two, or virtually obviously much more species of amphipathic peptides, major to ion leakage, lack of proton intent force, and eventually cellphone death [64-66].

#### ***Antimicrobial spectrum***

The low molecular weight bacteriocins of Gram-confident microorganism exhibit bactericidal pastime which is directed basically towards exclusive precise Gram-positive microorganism. For illustration, the lantibiotic nisin has been established to be amazing in opposition to many traces of Gram-constructive microorganism, along with staphylococci, streptococci, bacilli, clostridia, and mycobacteria [50].

### **Detection of bacteriocins produced through lactic acid microorganism (LAB) typical approaches Agar diffusion system**

A original approach for screening bacteriocin mission entails utilizing agar media contained in petri plates. Piddock has reviewed a wide variety of those approaches for detection and dimension of bacteriocin mission. There are really quite a lot of models, most of them derivatives of the “spot-on-outside” procedure, to be equipped to contain an agar overlay [67-70]. On this approach, the bacteriocin from the producer subculture is noticed on the indicator cultures and these plates are incubated even as earlier than examination for zones of inhibition during the growth of producing cultures. Kekessy and Pigué described a system in which the manufacturing and indicator traces had been each and every grown on unique gold common media. The manufacturing tradition used to be spot-inoculated onto a variety plate, and after growth, the agar mass used to be once aseptically dislodged with a spatula from the petri dish bottom and transferred to the lid of the dish [71]. A gentle agar overlay seeded with the indicator was once once then poured over the inverted agar. Following re-incubation, bacteriocin-positive cultures displayed a halo of clearing within the backyard across the average button of progress. This assay minimizes the penalties of acids and bacteriophages, considering that the bacteriocin-producing and indicator strains are bodily separated through an agar layer [72,73].

### **Mass spectrometry**

Mass spectrometry has been tailored for the quick detection of pediocin, nisin, brochocins A and B, and enterocins A and B from subculture supernatants by the use of Rose et al. The system is referred to as matrix-assisted laser desorption/ionization time-of-flight mass spectroscopy (MALDI-TOF MS) and was as quickly as on the devised for the examination of colossal molecules, similar to biopolymers [74-76].

### **Waft cytometry**

In a float cellphone, Mugochi et al. Developed a fast and sensitive system for detection of bacteriocins in fermentation broth. Low concentrations of potassium ions had been measured, so that launched potassium ions from a bacteriocin-touchy indicator stress instantly correlated to concentrations of crude bacteriocin present in fermentation broth injected into the cellular. This system in evaluation excellent to an usual agar exceptional diffusion assay [77,78].

### **PCR detection systems**

The PCR method has been used to rapidly check lactobacilli that may produce just right-characterised bacteriocins, on the other hand of counting on the utilization of problematic biochemical methods in order to even be more almost always than no longer required for the identification and characterization of such bacteriocins. PCR programs had been used to observe genes in fee for bacteriocin development and regulation in bacterial cultures. Rodriguez et al. demonstrated the amplification of a seventy 5 bp gene fragment of the lactocin S structural gene in seven bacteriocinogenic traces of lactobacilli remoted from fermented sausages [79]. In however one different work, Garde et al. detected the genes important for the synthesis of lactocin 481 and nisin utilizing PCR methods with detailed probes on an isolate of *L. Lactis* sub-sp. *Lactis*. To be able to decrease the time priceless for detection, procedures to simplify or even cast off the necessity for put up-amplification gel assays were furnished. A method for PCR quantification has been devised and is by and large known as “genuine-time” PCR when you consider that use of a fluorescent label makes it possible for the character to close to view the expand within the large style of motive DNA as it can be amplified [80-84]. The precise-time PCR process is based on detection and quantification using probably the most a quantity of kinds of fluorescent reporter molecules. This signal raises in direct percentage to the quantity of PCR product inside the response. A thermocycler organized with a fluorescence detection system measures the fluorescence in peculiarly designed tubes that include the response add-ons. The amplification final result in an attribute sigmoid formed curve which represents three phases of PCR: the lag part (little product accumulation), the exponential part (fast product accumulation) and the plateau section (no additional product is amplified) [85-89]. With the help of recording the amount of fluorescence emission in each and every cycle, it can be viable to detect the PCR for the duration of the exponential segment the area the primary large develop inside the fluorescence instantly correlates with the preliminary number of goal template and the PCR cycle at which this fluorescence is measured is most commonly called a threshold cycle (Ct) [90].

### **Components purposes**

Three procedures are mostly used in the utility of bacteriocins for biopreservation of foods:

- (1) Inoculation of food with LAB that produce bacteriocin within the merchandise. The competencies of the LAB to support and produce bacteriocin within the merchandise is essential for its robust use.
- (2) Addition of purified or semi-purified bacteriocins as food preservatives.
- (three) Use of a product beforehand fermented with a bacteriocin-producing stress as an ingredient in ingredients processing.

### **Biopreservation of dairy merchandise**

*L. Monocytogenes* has been the documented cause of an enormous form of outbreaks involving dairy merchandise, corresponding to pasteurized milk and cheese. Nisin has been demonstrated mighty inside the direction of *L. Monocytogenes* in dairy merchandise [91-94]. A important trouble in cheese creation is the *Clostridium*-associated butyric acid fermentation. Nisin is traditionally dropped at pasteurized, processed cheese spreads to prevent the outgrowth of clostridial spores, similar to *Clostridium tyrobutyricum* [95,96]. Lacticin 3147, a giant-spectrum, two-part bacteriocin produced by way of *Lactis* subsp. *Lactis* DPC 3147, is used to control cheddar cheese satisfactory by way of lowering nonstarter LAB populations in the course of ripening. The lacticin 3147 producing transconjugant has moreover been used as a protecting culture to inhibit *Listeria* on the epidermis of a mildew-ripened cheese [97-100]. Aside from using nisin, a bacteriocin (pediocin) produced via *Pediococcus acidilactici* %. Zero has tested to inhibit Gram-confident spoilage organisms like lactobacilli in salad dressings.

### **CONCLUSION**

Bacteriocins are protein-like antibiotics. The macromolecular bacteriocins exert their action by way of adsorption to specific receptors located on the outside surface of the sensitive pressure, followed by way of foremost organic and morphological alterations of both the bacterial telephone and the bacteriocin particle. This results within the killing of the sensitive strain without creation of extra bacteriocin particles, one of the vital foremost aspects which make them evidently exclusive to bacteriophages.

Essentially the most intensively studied companies of bacteriocin are the colicins that are produced via *E.Coli* and carefully related enterobacteria. They are most commonly plasmid-encoded proteins. Although many bacteriocins had been remoted and characterized, only some have confirmed industrial talents in meals application. Nisin is the only purified bacteriocin authorized for meals use in the US. It has been used as a meal preservative in more than 50 international locations, as a rule in cheese, canned vegetables, more than a few pasteurized dairy, liquid egg merchandise, and salad dressings.

Bacteriocins that have been used as meals preservatives have slightly slender endeavor spectra and are customarily now not active in opposition to Gram-bad bacteria. The simultaneous software of bacteriocins and non-thermal processing applied sciences, such as HP and PEF, to reinforce shelf life of foods is attractive since foods produced utilizing these non-thermal technologies most of the time have higher sensory and dietary qualities in comparison with merchandise produced making use of traditional thermal processing system.

### **REFERENCES**

1. Zhao T. Approaches for Reduction of Shiga Toxin-Producing *Escherichia coli* and *Salmonella* on Hide of Cattle. *J Food Microbiol Saf Hyg.* 2016;1:101.
2. Diop MB, et al. Efficiency of Neutralized Antibacterial Culture Supernatant from Bacteriocinogenic Lactic Acid Bacteria Supplemented With Salt in Control of Microorganisms Present in Senegalese Artisanally Handled Fish by Immersion Preservative Technology During Guedj Seafood Processing at 10°C and 30°C. *J Food Microbiol Saf Hyg.* 2016;1:102.
3. Althaus D, et al. Performance of the Assurance GDS® Assay for the Detection of *L. monocytogenes* in Pure Cultures and Spiked Food Samples. *J Food Microbiol Saf Hyg.* 2016;1:103.
4. Gámez MN, et al. Pathogen Persistence in Restaurant Menus: Comparison between Materials. *J Food Microbiol Saf Hyg.* 2016;1:104.
5. Mengual Lombar M, et al. Accessories of Food Handlers and Restaurant Staff as a Source for Food Contamination. *J Food Microbiol Saf Hyg.* 2016;1:105.
6. Pandey KR and Vakil BV. Functional Characterization of Bacteriophage Resistant Mutants of Probiotic *B. coagulans*. *J Food Microbiol Saf Hyg.* 2016;1:106.

7. Amadi EN and Kiin-Kabari DB. Nutritional Composition and Microbiology of Some Edible Insects Commonly Eaten in Africa, Hurdles and Future Prospects: A Critical Review. *J Food Microbiol Saf Hyg*. 2016;1:107.
8. Mishra B and Dinesh SN. Universal Diet and Beverage Code: 'The Rules of Halves in Human Nutrition. *J Nutr Food Sci*. 2016;6:e125.
9. Roberto F. Olive Oil Phenolic Compounds: May Prevent Cancer in Human?. *J Nutr Food Sci*. 2016;6:e126.
10. Tanweer S, et al. Radical Scavenging Linked Antioxidant Comparison and Quantification of Conventional and Supercritical Fluid Ginger Extracts. *J Nutr Food Sci*. 2016;6:511.
11. Birketvedt GS, et al. A Dietary Supplement in Combination with an Education Plan and a Long-Term Follow-up Significantly Decrease Blood Pressure, Body Weight and Body Fat. *J Nutr Food Sci*. 2016;6:512.
12. Khalifa I, et al. Influencing of Guava Processing Residues Incorporation on Cupcake Characterization. *J Nutr Food Sci*. 2016;6:513.
13. Mensah EO, et al. Thermal Stability of  $\beta$ -Amylase Activity and Sugar Profile of Sweet-Potato Varieties during Processing. *J Nutr Food Sci*. 2016;6:515.
14. Nwadioha SI, et al. Microbiologic Review of Seminal Fluids in a Nigerian Tertiary Health Centre. *Arch Clin Microbiol*. 2016;7:4.
15. Kaur J, et al. Comparative Evaluation of CFX96™ Real Time PCR with Conventional PCR for Rapid Diagnosis of Mycobacterium tuberculosis Complex in Clinical Isolates. *Arch Clin Microbiol*. 2016;7:4.
16. Allen C and Parks OW. Photodegradation of Riboflavin in Milks Exposed to Fluorescent Light. *Journal of Dairy Science*. 1979;62:1377-1379.
17. Dimick PS. Photochemical Effects on Flavor and Nutrients of Fluid Milk. *Canadian Institute of Food Science and Technology Journal*. 1982;15:247-256.
18. Allen C and Parks OW. Evidence for Methional in Skim Milk Exposed to Sunlight. *Journal of Dairy Science*. 1975; 58:1609-1611.
19. Finley JW and Shipe WF. Isolation of a Flavor Producing Fraction from Light Exposed Milk. *Journal of Dairy Science*. 1969;54:15-20.
20. Beuchat LR, et al. Standardization of a method to determine the efficacy of sanitizers in inactivating human pathogenic microorganisms on raw fruits and vegetables. *Journal of Food Protection*, 2001;64:1079-1084.
21. Parish ME, et al. Methods to Reduce Eliminate pathogens from fresh and fresh-cut produce. *Comprehensive Reviews in Food Science and Food Safety*, 2003;2:161-173.
22. Iguban EB, et al. The In Vitro Anti-Microbial Activity of Multipurpose Contact Lens Solutions against Standard Strains of Common Ocular Pathogens: The Effect of Duration from First Use. *J Clin Exp Ophthalmol*. 2016, 7:560.
23. Chatterjee M, et al. Effect of Fruit Pectin on Growth of Lactic Acid Bacteria. *J Prob Health*. 2016;4:147.
24. Liu YH, et al. Inhibitory Effect of Lactic Acid Bacteria on Uropathogenic Escherichia coli-Induced Urinary Tract Infections. *J Prob Health*. 2016;4:144.
25. Baruah R, et al. Heteropolysaccharides from Lactic Acid Bacteria: Current Trends and Applications. *J Prob Health*. 2016;4:141.
26. Sanalibaba P and Çakmak GA (2016) Exopolysaccharides Production by Lactic Acid Bacteria. *Appli Micro Open Access* 2:115.
27. Patel S, et al. (2012) Potentials of exopolysaccharides from lactic Acid bacteria. *Indian J Microbiol* 52: 3-12.
28. Ismail B and Nampoothiri KM (2010) Production, purification and structural characterization of an exopolysaccharide produced by a probiotic *Lactobacillus plantarum* MTCC 9510. *Arch Microbiol* 192: 1049-1057.
29. Mozzi F, et al. UDP-galactose 4-epimerase: a key enzyme in exopolysaccharide formation by *Lactobacillus casei* CRL 87 in controlled pH batch cultures. *J Appl Microbiol*. 2003;94: 175-183.
30. Patel A and Prajapati JB. Food and Health Applications of Exopolysaccharides produced by Lactic acid Bacteria. *Adv Dairy Res*. 2013;1:107.
31. Malang SK, et al. Characterization of exopolysaccharide and ropy capsular polysaccharide formation by *Weissella*. *Food Microbiol*. 2015;46: 418-427.
32. Welman AD and Maddox IS. Fermentation performance of an exopolysaccharide-producing strain of *Lactobacillus delbrueckii* subsp. *bulgaricus*. *J Ind Microbiol Biotechnol*. 2003;30: 661-668.
33. Kristo E, et al. The role of exopolysaccharide produced by *Lactococcus lactis* subsp. *cremoris* in structure formation and recovery of acid milk gels. *Int Dairy J*. 2011;21: 656-662.

34. Sheng ZY, et al. The Study of Analytical Identification on Main Monomer Compounds of Spoiled Grass Carp by High Performance Liquid Chromatography of Quadrupole Time of Flight Mass Spectrometry. *J Food Process Technol.* 2016;7:600.
35. Zhou JL, et al. Herbal medicine analysis by liquid chromatography/time-of-flight mass spectrometry. *J Chromatogr A.* 2009;1216: 7582-7594.
36. Qu CL, et al. Studies on fragmentation pathways of amino acids and their interactions with ginsenoside Rb<sub>3</sub> by spectroscopy ionization mass spectrometry. *Chem J Chinese U.* 2008;9:1721-1726.
37. Huang YF and Hu J. Simultaneous analysis of twenty free amino acids in tobacco using liquid chromatography-electrospray ionization/iontraps tandem mass spectrometry. *Chin J Chromatogr.* 2010;6:615-622.
38. Wang Y, et al. Fragmentation characteristics and utility of ammonium ions for peptide identification by MALDI TOF/TOF spectrometry. *Chinese J Anal Chem.* 2014;7:1010-1016.
39. Daniel D, et al. Determination of biogenic amines in beer and wine by capillary electrophoresis-tandem mass spectrometry. *J Chromatogr A.* 2015;1416: 121-128.
40. Wu YL, et al. Simultaneous determination of sixteen amide fungicides in vegetables and fruits by dispersive solid phase extraction and liquid chromatography-tandem mass spectrometry. *J Chromatogr B.* 2015;989:11-20.
41. Zoran K, et al. Liquid chromatography tandem mass spectrometry method for characterization of monoaromatic nitro-compounds in atmospheric particulate matter. *J Chromatogr A.* 2012;1268:35-43.
42. Sun Y, et al. Qualitative and quantitative analysis of phenolics in *Tetrastigma hemsleyanum* and their antioxidant and anti-proliferative activities. *J Agric Food Chem.* 2013;61: 10507-10515.
43. Fu Y, et al. Characterization and identification of baccharane glycosides in *Impatiens Semen* by rapid-resolution liquid chromatography with electrospray ionization quadrupole time-of-flight tandem mass spectrometry. *J Pharm Biomed Anal.* 2012; 65:64-71.
44. Chen XF, et al. Liquid chromatography coupled with time-of-flight and ion trap mass spectrometry for qualitative analysis of herbal medicines. *J Pharmaceut Ana.* 2011; 4:235-245.
45. Lee SJ, et al. Composition of organic acids and physiological functionality of commercial makgeolli. *Korean J Food Sci Technol.* 2011; 43: 206-212.
46. Zhang Q, et al. The Strategies for Increasing Cordycepin Production of *Cordyceps Militaris* by Liquid Fermentation. *Fungal Genom Biol.* 2016; 6:134.
47. Saithi S, et al. Mathematical Modeling of Biomass and Enzyme Production Kinetics by *Aspergillus niger* in Solid-State Fermentation at Various Temperatures and Moisture Contents. *J Microb Biochem Technol.* 2016; 8:123-130.
48. Karine Rebouças H, et al. Evaluating Physicochemical and Rheological Characteristics and Microbial Community Dynamics during the Natural Fermentation of Cassava Starch. *J Food Process Technol.* 2016; 7:578.
49. Feye KM, et al. Abrogation of *Salmonella* and *E. coli* O157:H7 in Feedlot Cattle Fed a Proprietary *Saccharomyces cerevisiae* Fermentation Prototype. *J Vet Sci Technol.* 2016; 7:350.
50. Abdullah JJ, et al. Optimizing Cellulase Production from Municipal Solid Waste (MSW) using Solid State Fermentation (SSF). *J Fundam Renewable Energy Appl.* 2016; 6:206.
51. Raja J, et al. Effect of Dry Salt and Brine on the Fermentation and Colour of Blanched Garlic. *J Nutr Food Sci.* 2016; 6:484.
52. Ranganna S. Handbook of analysis and quality control for fruit and vegetable products. Tata McGraw Hill Publishing Company Ltd, New Delhi. 1986; pp: 1112.
53. de Castro A, et al. Lactic acid fermentation and storage of blanched garlic. *Int J Food Microbiol.* 1998; 39: 205-211.
54. Hugenholtz J. Citrate metabolism in lactic acid bacteria. *FEMS Microbiol.* 1993; 12: 165-178.
55. Joslyn MA and Sano T. The formation and decomposition of green pigment in crushed garlic tissue. *J Food Sci.* 1956; 21:170-183.
56. Lukes TM. Factors governing the greening of garlic puree. *J Food Sci.* 1986; 51:1577-1582.
57. McFeeters RF, et al. Storage stability of vegetable fermented with pH control. *J Food Sci.* 1983; 48: 975-981.
58. Fuselli SR, et al. Microbiological study of dehydrated garlic (*Allium sativum* L.) and onion (*Allium cepa* L.). *Rev Argent Microbiol.* 2004; 36: 139-144.

59. McDonald LC, et al. Acid Tolerance of *Leuconostocmesenteroides* and *Lactobacillus plantarum*. *Appl Environ Microbiol.* 1990; 56: 2120-2124.
60. Frazier WC and Dennise CW. *Food Microbiology* (4<sup>th</sup> edn) Tata McGraw-Hill Publishing Company Limited, New Delhi, India. 1995.
61. Sano T and Joslyn MA. The formation and decomposition of green pigment in crushed garlic tissue. *Food Res.* 2010; 21: 170-183.
62. *American Heritage Dictionary of the English Language Fifth Edition.* Houghton Mifflin Harcourt Publishing Company. 2011.
63. Arroqui C, et al. Effect of different soluble solids in water on the ascorbic acid losses during water blanching of potato tissue. *Journal of Food Eng.* 2001; 47:123-126.
64. Pilnik W and Voragen AGJ. The significance of endogeneous and exogenous pectic enzymes in fruits and vegetable processing. In: *Food enzymology*, PF Fox (edn), Essex, England. 1991; pp: 318.
65. USDA Agricultural Research Service. USDA national nutrient database for standard reference. 2003.
66. Yu TH, et al. Volatile compounds of blanched, fried blanched and baked blanched garlic slices. *J Agric Food Chem.* 1994; 42: 1342-1347.
67. USDA National Genetic Research Programs. Germplasm Resource Information Network Beltsville, Maryland. 2006.
68. Vergara SE and Tchobanoglous G. Municipal solid waste and the environment: a global perspective. *Annual Review of Environment and Resources.* 2012; 37: 277-309.
69. Keeling C. *Canterbury Region Waste Data Report 2009/2010.* Environment Canterbury. 2011.
70. Ray AK, et al. Optimization of fermentation conditions for cellulase production by *Bacillus subtilis* CY5 and *Bacillus circulans* TP3 isolated from fish gut. *Acta Ichthyologicaet Piscatoria.* 2007; 37: 47-53.
71. Rajoka MI. Influence of various fermentation variables on exo-glucanase production in *Cellulomonasflavigena*. *Electr J Biotechnol.* 2010; 7: 07-08.
72. Guruchandran V and Sasikumar C. Cellulase production by *Aspergillus niger* fermented in saw dust and Bagasse. *J Cell Tissue Res.* 2010; 10: 2115.
73. Zhao SH, et al. High-yield cellulase production in solid-state fermentation by *Trichodermareesei* SEMCC-3.217 using water hyacinth (*Eichhorniacrassipes*). *Afr J Biotechnol.* 2013; 10: 10178-10187.
74. Singhanian RR, et al. Solid-state fermentation of lignocellulosic substrates for cellulase production by *Trichodermareesei* NRRL 11460. *Indian J Biotechnol.* 2006; 5: 332-336.
75. Gautam SP, et al. Optimization for the production of cellulase enzyme from municipal solid waste residue by two novel cellulolytic fungi. *Biotechnol Res Int.* 2011; 8.
76. Sluiter A, et al. Determination of sugars, byproducts, and degradation products in liquid fraction process samples. *National Renewable Energy Laboratory.* 2006.
77. Campbell CR. Determination of total nitrogen in plant tissue by combustion. *Plant Anal Ref Proc for S US Southern Coop Ser Bull.* 1992; 368: 20-22.
78. NasirIqbal HM, et al. Media optimization for hyper-production of carboxymethylcellulase using proximally analyzed agroindustrial residue with *Trichodermaharzianum* under SSF. *IJAVMS.* 2010; 4: 47-55.
79. Hokura A, et al. Multielement determination of major-to-ultratrace elements in plant reference materials by ICP-AES/ICP-MS and evaluation of their enrichment factors. *Analytical Sciences.* 2000; 16: 1161-1168.
80. Barlaz MA, et al. Biodegradative analysis of municipal solid waste in laboratory-scale landfills. *Environmental Protection Agency.* 1997.
81. Jones KL, et al. Methane generation and microbial activity in a domestic refuse landfill site. *Eur J Appl Microbiol Biotechnol.* 1983; 18: 242-245.
82. Ham RK, et al. Chemical characterization of fresh kills landfill refuse and extracts. *J Environ Engineer.* 1993; 119: 1176-1195.
83. Gallert C and Winter J. *Bacterial metabolism in wastewater treatment systems.* 2005.
84. Schober TJ (2009) *Manufacture of gluten-free speciality breads and confectionery products.* In E. Gallagher (ed.), *Gluten-free food science and technology* (pp. 130-180). Oxford: Wiley-Blackwell.
85. Adams RP (1995) *Identification of essential oil components by gas chromatography mass spectroscopy.* Illinois: Allured Publishing Corporation.
86. Anderson RA, et al. Gelatinization of corn grits by roll- and extrusion-cooking. *Cereal Sci Today.* 1969;14: 47.

87. Faubion JM and Hosney RC. High-temperature short time extrusion cooking of wheat starch and flour. I. Effect of moisture and flour type on extrudate properties. *Cereal Chem.* 1982;59: 529-533.
88. Oyewole OB. Characteristics and significance of yeasts' involvement in cassava fermentation for 'fufu' production. *Int J Food Microbiol.* 2001;65: 213-218.
89. Ascheri DPR, et al. Correlation between grain nutritional content and pasting properties of pre-gelatinized red rice flour. *Revista Ceres.* 2012;59: 16-24.
90. Adegunwa MO, et al. Effects of fermentation length and varieties on the pasting properties of sour cassava starch. *Afr J Biotechnol.* 2011;10: 8428-8433.
91. Aquino ACMS, et al. Standardization of the sour cassava starch reduces the processing time by fermentation water monitoring. *Int J Food Sci Tech.* 2013;48: 1892-1898.
92. Calleja A, Falqué E (2005) Volatile composition of Mencia wines. *Food Chem* 90: 357-363.
93. Marcon MJA, et al. Effect of the improved fermentation on physicochemical properties and sensorial acceptability of sour cassava starch. *Braz Arch BiolTechnol.* 2007;50: 1073-1081.
94. Calleja A and Falqué E. Volatile composition of Mencia wines. *Food Chem.* 2005;90: 357-363.
95. Marcon MJA, et al. Effect of the improved fermentation on physicochemical properties and sensorial acceptability of sour cassava starch. *Braz Arch BiolTechnol.* 2007;50: 1073-1081.
96. Shih CY, et al. A Study of Rotational Ultrafiltration System for Fructose Recovery from Glucose Fermentation Process. *J Food Process Technol.* 2015;6:494.
97. Danfeng S. Recent Application of Probiotics in Food and Agricultural Science. 2012.
98. Mousavi ZE, et al. Fermentation of pomegranate juice by probiotic lactic acid Bacteria *WJMB.* 2011;27: 123-128.
99. Karaaslan M, et al. Antiproliferative and antioxidant activities of Turkish pomegranate (*Punicagranatum L.*) accessions *IJFS.* 2014;82-904.
100. Heldman DR and Hartel RW. Principles of food processing. *Food Science and Nutrition, Springer Science & Business Media.* 1997.