

Peptide Therapy in Sepsis and Inflammation: A Novel Strategy to Suppress Inflammation

Hidechika Okada, Alan Okada

Research Institute for Protein Science, Nakayama-cho, Mizuho-ku, Nagoya, Japan.

Review Article

Received date: 05/07/2015

Accepted date: 29/09/2015

Published date: 01/10/2015

*For Correspondence

Hidechika Okada, Research Institute for Protein Science, Nakayama-cho, Mizuho-ku, Nagoya, Japan.

E-mail: hiokada@med.nagoya-cu.ac.jp

Keywords: Urine concentration, Osmolality, Refractometry, Reagent strip, Cow

REVIEW

Antisense Homology Box (AHB)

In 1984, Blalock proposed the possible role of antisense peptides for molecular interaction among proteins ^[1,2].

We speculated that interactions between sense-and antisense-peptides should play a role in formation of the tertiary structure of proteins. We developed a novel computer program named ANTIS to find antisense peptide sequences between proteins to be compared ^[2-4]. ANTIS revealed the presence of an appreciable number of sense and antisense peptide pairs within any protein molecule and those portions were designated as antisense homology boxes (AHB) ^[5-7]. One of the AHB peptides of endothelin receptor (ETR), named ETR-P1/fl, had the capacity to interfere with the function of ETR.

Complementary peptide

Each peptide should have specific structure determined by its amino acid sequence which may react with its antisense peptide ^[8,9]. To generate candidates of complementary peptide (C-pep) reactive to a target amino acid sequence based upon the sense-antisense amino acid relationship ^[10-14]. We invented an evolutionary computer program that generates C-pep sequences that have a potential to interact with a target peptide ^[15]. Out of 19 C-peps targeted to C5a anaphylatoxin, 7 exhibited an inhibitory effect ^[16,17].

Inhibition of C5a anaphylatoxin

C5a anaphylatoxin is considered to be an effective target for treatment of hyperinflammation since C5a stimulates generation of tumor necrosis factor alpha (TNF α) and other inflammatory cytokines ^[18,19]. Although C5a generated *in vivo* is regulated by carboxypeptidase N and more efficiently by carboxypeptidase R (CPR), excessive generation of C5a appears to exceed the capacity of CPR, since administration of lipopolysaccharide (LPS) at a lethal dose to rats exhausted CPR capacity before death ^[20,23].

On the other hand, antibodies to C5a was effective in treating experimental primate models of sepsis, indicating that C5a inhibitors should be useful for treatment of patients suffering from hyperinflammation such as in sepsis and multiple organ failure ^[24-29].

C5a inhibitory peptides

Amino acids 37 to 53 of C5a (RAARISLGPRCIKAFTE) is an antisense peptide to AHB peptides of the C5a receptor (C5aR), and this has been designated PL37 ^[30]. This region of C5a is presumed to be a potential site for C5aR stimulation. Using the computer program MIMETIC, we generated 19 C-peps to PL37 ^[31,32]. One of the 7 inhibitory C-peps to PL37 which interfered with C5a function was termed PepA (ASGAPAPGPAGPLRPMF). To improve stability, we modified PepA by acetylation of its N-terminal alanine generating acetylated PepA (AcPepA) which was more stable in animal experiments. We performed experiments in Cynomolgus monkeys in lieu of using humans ^[33].

One of the inhibitory C-peps of C5a, termed AcPepA, was effective in Cynomolgus monkeys intravenously infused with a lethal dose of bacterial LPS (4 mg/kg) destined to die ^[34]. The monkeys were rescued by intravenous administration of 2 mg/kg/h of AcPepA. The excellent therapeutic effect of AcPepA is likely to be due to restriction of high mobility group box 1 (HMGB1) surge induced by the effect of C5a on C5L2 ^[35,36], which is the second C5a receptor, since the released HMGB1 has the capacity to stimulate TLR4 as an endogeneous ligand resulting in further activation of inflammatory cells to release inflammatory cytokines forming positive feedback circuit of inflammation ^[37,38].

REFERENCES

1. Blalock J E and Smith EM. Hydropathic anti-complementary of amino acids based on the genetic code. *Biochem Biophys Res Comm.* (1984);121: 203-207.
2. Baranyi L, et al. The antisense homology box: A new motif within proteins that encodes biologically active peptides. *Nature Med.* (1995);1: 894-901.
3. Blalock JE. Complementary of peptides specified by 'sense' and 'antisense' strands of DNA. *Trends Biotech.* (1991);6: 140-144.
4. Trospha A, et al. Making sense from antisense: a review of experimental data and developing ideas on sense-antisense peptide recognition. *J Molec Recognition.* (1992);5: 43-53.
5. Eberle AN and Huber M. Antisense peptides: tools for receptor isolation? Lack of antisense MSH and ACTH to interact with their sense peptides and to induce receptor-specific antibodies. *J Receptor Res.* (1991);11: 13-43.
6. Ulla B. Rasmussen and Rolf-Dieter Hesch. On antisense peptides: The parathyroid hormone as an experimental example and a critical theoretical view. *Biochem Biophys Res Comm.* (1987);149: 930-938.
7. Baranyi L, et al. Antisense Homology Box-Derived Peptides Represent a New Class of Endothelin Receptor Inhibitors. *Peptides.* (1998);19: 211-223.
8. Goldstein A and Brutlag DL. Is there a relationship between DNA sequences encoding peptide ligands and their receptors? *Proc National Acad Sci.* (1989);86: 42-45.
9. Guillemette G, et al. The peptide encoded by angiotensin II complementary RNA does not interfere with angiotensin II action. *Biochem J.* (1989);261: 309.
10. de Gasparo M, et al. Are the antibodies to a peptide complementary to angiotensin II useful to isolate the angiotensin II receptor? *Biochem J.* (1989);261:310-311.
11. Campbell W, et al. A Novel Genetic Algorithm for Designing Mimetic Peptides That Interfere with the Function of a Target Molecule. *Microbiol Immunol.* (2002);46: 211-215.
12. Hosokawa M, et al. Inhibition of HIV-1 infection in cells expressing an artificial complementary peptide. *Biochem Biophys Res Comm.* (2004);324: 236-240.
13. Shimomura Y, et al. Modulation of Procarboxypeptidase R (ProCPR) Activation by Complementary Peptides to Thrombomodulin. *Microbiol Immunol.* (2003);47: 241-245.
14. Fujita E, et al. Inactivation of C5a Anaphylatoxin by a Peptide That Is Complementary to a Region of C5a. *J Immunol.* (2004);172: 6382-6387.
15. Ember JA and Hugli TE. Complement factors and their receptors. *Immunopharmacology.* (1997);38: 3-15.
16. Okada N, et al. Increased inhibitory capacity of an anti-C5a complementary peptide following acetylation of N-terminal alanine. *Microbiol Immunol.* (2007);51: 439-443.
17. Hussein MH, et al. An acetylated anti-C5a complementary peptide reduced cytokines and free radicals and prolongs survival time in a neonatal sepsis model. *Mol Immunol.* (2009);46: 2825.
18. Czermak BJ, et al. Protective effects of C5a blockade in sepsis. *Nature Med.* (1999);5: 788-792.
19. Guo RF and Ward PA. Role of C5a in inflammatory responses. *Annu Rev Immunol.* (2005);23: 821-852.
20. Michael K. Targeting complement in therapy. *Immunol Rev.* (2001);180: 177-189.
21. Campbell W, et al. Inactivation of C3a and C5a Octapeptides by Carboxypeptidase R and Carboxypeptidase N. *Microbiol Immunol.* (2002);46: 131-134.
22. Campbell W, et al. Carboxypeptidase R is an inactivator of complement-derived inflammatory peptides and an inhibitor of fibrinolysis. *Immunol Rev.* (2001);180: 162-167.
23. Kato K, et al. Changes in arginine carboxypeptidase (CPR) activity in stressed rats. *Pathophysiology.* (1994);1: 131-136.
24. Riedemann NC, et al. Increased C5a receptor expression in sepsis. *J Clin Invest.* (2002);110: 101-108.

25. Stevens JH, et al. Effects of anti-C5a antibodies on the adult respiratory distress syndrome in septic primates. *J Clin Invest.* (1986);77: 1812-1816.
26. Riedemann NC, et al. Novel strategies for the treatment of sepsis. *Nature Med.* (2003);9: 517-24.
27. Baranyi L, et al. Antisense homology boxes in C5a receptor and C5a anaphylatoxin: a new method for identification of potentially active peptides. *J Immunol.* (1996);157: 4591-4601.
28. Farkas I, et al. Complement C5a ana-phylatoxin fragment causes apoptosis in TGW neuroblastoma cells. *Neuroscience.* (1998);86: 903-911.
29. Abe M, et al. Contribution of Anaphylatoxins to Allergic Inflammation in Human Lungs. *Microbiol Immunol.* (2005);49: 981-986.
30. MizueY, et al. Quantitation of macrophage migration inhibitory factor (MIF) using the one-step sandwich enzyme immunosorbent assay: elevated serum MIF concentrations in patients with autoimmune diseases and identification of MIF in erythrocytes. *Int J Molecular Medicine.* (2000);5: 397-403.
31. Park JS, et al. High mobility group BOX 1 protein interacts with multiple Toll-like receptors. *Am J Physiol Cell Physiol.* (2006);290: 917-924.
32. Wang H, et al. HMG-1 as a Late Mediator of Endotoxin Lethality in Mice. *Science.* (1999);285: 248-251.
33. Yang H, et al. Reversing established sepsis with antagonists of endogenous high-mobility group box 1. *Proc Natl Acad Sci.* (2004);101: 296-301.
34. Okada N, et al. HMGB1 release by C5a anaphylatoxin is an effective target for sepsis treatment. *Nature Precedings.* (2011).
35. Chen NJ, et al. C5L2 is critical for the biological activities of the anaphylatoxins C5a and C3a. *Nature.* (2007);446: 203-207.
36. Yu M, et al. HMGB1 signals through toll-like receptor (TLR) 4 and TLR2. *Shock.* (2006);26: 174-179.
37. Klune JR, et al. HMGB1: endogenous danger signaling. *Mol Med.* (2008);14: 476-484.
38. Rittirsch D, et al. Functional roles for C5a receptors in sepsis. *Nature Med.* (2008);14: 551-557.