

RP HPLC Validation of PREGABLIN in Bulk and Dosage Form

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Research Article

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ABSTRACT

In this article we are producing the results obtained checking the validation parameters System suitability, Specificity, Precision in this Method precision, Accuracy (recovery), Linearity of test method, Ruggedness, Robustness, Solution stability and showed that the results obtained all within the limits by using the method developed in trial and error method.

INTRODUCTION

All HPLC ways used for the event of prescription drugs and for the determination of their quality have to be compelled to be valid [1-6]. In cases whereby ways from the Pharmacopoeia's area unit used, it's not necessary to gauge their quality, given that the analyses area unit conducted strictly per the methods' meant use [6-8].

The parameters tested throughout the tactic validation as outlined by the ICH, USP and government agency and different health organizations [9-13] area unit the following: Specificity or property, exactness (repeatability, intermediate exactness, reliableness or ruggedness), accuracy or exactitude or bias, dimensionality vary, limit of detection, limit of quantitation and strength. The terms property and specificity area unit usually used interchangeably [13-19]. The USP treatise defines property of associate analytical methodology as its ability to live accurately an analyte within the presence of interference, like artificial precursors, excipients, enantiomers and celebrated (or likely) degradation merchandise that may be gift within the sample matrix [20-25].

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- Recovery
- Response function
- Sensitivity
- Precision
- Accuracy
- Limit of detection
- Limit of quantization
- Ruggedness
- Robustness
- Stability
- System suitability.

METHOD VALIDATION

System suitability

Suitability delineates for the tactic of study is established by injecting 5 times with normal and double with standard value system [30-33]. Suitability parameters as per the take a look at procedure [34-39].

Purpose

To establish the system quality as per take a look at technique [40-43] (Table 1).

Sequence

Table 1. System suitability sequences.

S. No.	Type of sample	No. of injections
1	Blank	01
2	Standard 1 solution	05
3	Standard 2 solution	02

Evaluate the subsequent system quality parameters [44-47]

%RSD, Theoretical plates & spatiality for traditional

Similarity issue between 2 standards

Acceptance criteria

- The sharp RSD for the retention times of principal peak from ten replicate injections of every normal answer ought to be less than 2.0% [47-50].
- The quantity of theoretical plates (N) for pregablin peaks is NLT 3000 [19].
- The Tailing issue (T) for pregablin peaks is NMT 2.0%
- Similarity issue between 2 standards ought to be zero.985 to 1.015 [39,45].

Observation

The similarities between 2 standards ought to be at intervals limits [51,52].

Specificity

Placebo interference

A study to ascertain the interference of placebo was conducted. Samples were ready in triplicate by taking the placebo such as concerning the burden in portion of take a look at preparation as per the take a look at technique [53,54]. Recording of placebo failed to show any additional peaks. This means that the excipients employed in the formulation don't interfere within the assay of pregablin tablets [55].

Acceptance criteria

No interference at the retention times of pregablin and analyte peak purity ought to be NLTO.99.

Observation

From Placebo chromatograms, it absolutely was finished that there was no interference with placebo as no peaks were determined at the retention times of pregablin peak [56-58].

Interference from degraded products

A study was conducted to demonstrate the effective separation of degradants from pregablin in capsules (150 mg). Separate parts of Drug product and Placebo were exposed to following stress conditions to induce degradation [59-61].

- Water degradation
- Acid degradation
- Base degradation
- Peroxide degradation
- Thermal degradation
- UV degradation
- Humidity degradation

Stressed samples were injected into the HPLC system with photograph diode array detector by following take a look at technique conditions [62,63]. All degrading peaks of pregablin within the chromatograms of all samples and placebo failed to show any substantial peaks beneath the higher than conditions [64-67]. The chromatograms of stressed samples were evaluated for peak purity of pregablin victimisation water's Empower software system [68]. For all forced degradation samples the degradants mustn't interference in quantitating the pregablin [69-71].

Precision

System precision

Standard resolution ready as per check methodology and injected 5 times [39].

Purpose

The purpose of this study is to determine the exactitude of the HPLC system being employed for the analysis [65] (Table 2).

Sequence

Table 2. System precision sequences.

S. No.	Type of sample	No. of injections
1	Blank	01
2	Placebo solution	01
3	Standard solution	05

Acceptance criteria

The % relative variance of pregablin from the six units ought to be less than 2.0% [72].

Method precision

Prepared six sample preparations severally victimisation single batch of pregablin.

Purpose

To check the repeatability of check results obtained by this methodology [73-75] (Table 3).

Sequence

Table 3. Method precision sequences.

S. No.	Sample	No. of injections
1	Diluent	1
2	placebo	1
3	Standard solution	5
4	Precision set 1	2

5	Precision set 2	2
6	Precision set 3	2
7	Precision set 4	2
8	Precision set 5	2
9	Precision set 6	2
10	Bracketing standard	1

Acceptance criteria

The sharp relative variance of pregablin from the six units ought to be less than 2.0% [41]. The assay of pregablin capsules 150 mg ought to be not but 90% and less than 110% [19].

Observation

Test results of pregablin in capsules (150 mg) area unit showing that the check methodology is precise. Refer **Table 3** for system exactitude and for methodology exactitude.

Accuracy (recovery)

Purpose

To establish methodology accuracy.

Study design

Demonstrate the accuracy of the check methodology by getting ready recovery samples (i.e., spiking placebo with is aware of quantities of standard) at the extent of fifty, 100% and one hundred and fiftieth of target concentration [76-79]. Prepare the recovery samples in triplicate and injecting duplicate at every level. The accuracy of the strategy shall is set by recovery experiments [80-83].

Procedure

The accuracy of the strategy shall is set by recovery experiments. The recovery is performed by adding pregablin commonplace to the placebo (**Tables 4 and 5**) (excipients mixture) within the vary of 50%-150% of check concentration [84,85].

Preparations

Table 4. Accuracy sequences.

Spike level (%)	Standard spiked (mg)	Weight of the placebo (mg)	Make up to the volume (ml)	Final concentration in mcg/ml
50%	15	340.0	20	750
100%	30	340.0	20	1500
150%	45	340.0	20	2250

Chromatograph the below samples and calculate the proportion recovery for the quantity value-added. Appraise the exactitude of the recovery at every level by computing the relative variance of triplicate recovery results [86].

Sequence

Table 5. Preparation of solutions to check accuracy.

S. No.	Sample	No. of injections
1	diluent	01
2	placebo	01
3	Standard solution	05
4	Recovery (50%) – Set 1	02
5	Recovery (50%) – Set 2	02
6	Recovery (50%) – Set 3	02
7	Recovery (100%) – Set 1	02
8	Recovery (100%) – Set 2	02
9	Recovery (100%) – Set 3	02
10	Recovery (150%) – Set 1	02
11	Recovery (150%) – Set 2	02
12	Recovery (150%) – Set 3	02
13	Bracketing standard	01

Acceptance criteria

The relative variance of assay mustn't be over 2.0% [39,87]. The typical recovery for every level must not be but 90% and not be over 110% [19].

Observation

The relative variance of assay mustn't be over a 2.0% [88]. The typical recovery for every level mustn't be but 90% and not be over 110% [22].

Linearity of test method

Purpose

To establish the dimensionality of analyte inside the required vary [89].

Study design

Demonstrate the dimensionality of analyte commonplace over the vary of 50%-150% of target concentration as mentioned below [90-92]. Preparation of dimensionality stock solution: Weigh accurately concerning 300 mg pregablin commonplace into 20 ml meter flask (**Tables 6 and 7**), add 15 ml of thinner and sonicate to dissolve, further compose the amount with thinner (1500 ppm) [93-95].

Preparation of linearity solution

Table 6. Preparation of linearity solution.

Linearity level	Stock solution to be taken in ml	Make up to the volume (ml)	Final concentration in mcg/ml
50%	0.5	10	750

60%	0.6	10	900
80%	0.8	10	1200
100%	1.0	10	1500
120%	1.2	10	1800
140%	1.4	10	2100
150%	1.5	10	2250

Sequence

Table 7. Linearity sequences.

S. No.	Sample	No. of injections
1	diluent	1
2	Standard solution	5
3	Linearity level - 1 (50%)	3
4	Linearity level - 1 (60%)	3
5	Linearity level - 1 (80%)	3
6	Linearity level - 1 (100%)	3
7	Linearity level - 1 (120%)	3
8	Linearity level - 1 (140%)	3
9	Linearity level - 1 (150%)	3
10	Bracketing standard	1

Inject these solutions through HPLC and record the height space of dimensionality solutions [96]. Plot a graph of concentrations (in x-axis) vs. peak space (in y-axis) [35]. Evaluate the coefficient of correlation between concentration and peak space [7].

Acceptance criteria

Coefficient of correlation ought to be not but 0.9990 [97]. % RSD for level one and Level half-dozen ought to be less than 2.0% [65].

Observation

The coefficient of correlation was found to be 0.99958. From the higher than study it absolutely was established that the dimensionality of check methodology is from 50% to 150% of the target concentration.

Ruggedness

Purpose

To demonstrate the duplicability of check results obtained by this check methodology for the variability particularly System to system/Analyst to Analyst/column to Column variability study was conducted on totally different HPLC systems, columns and different analysts below similar conditions at different times [98,99].

Study design

Carry out exactitude study in six preparations of assay on one batch sample by 2 totally different analysts, on 2 totally different columns (**Table 8**) and on 2 totally different instruments [100].

Sequence

Table 8. Ruggedness sequences.

S. No.	Sample	No. of injections
1	Diluent	1
2	placebo	1
3	Standard solution	5
4	Precision set 1	2
5	Precision set 2	2
6	Precision set 3	2
7	Precision set 4	2
8	Precision set 5	2
9	Precision set 6	2
10	Bracketing standard	1

Comparison of each the results obtained on 2 totally different HPLC systems, column and different analysts shows that the assay check methodology is rugged for System to system/Analyst to Analyst/column to Column variability [25].

Acceptance criteria

The % relative variance of pregablin from the six sample preparations ought to be less than 2.0% [12]. All individual assays of pregablin capsules 150 mg should be between 90.0%-110.0% [19].

Observation

The % RSD was found with within the limits.

Robustness

Effect of variation of flow rate and column temperature

Purpose

To establish the lustiness of check methodology and to demonstrate its reliableness for minor changes in activity conditions [101] (Table 9).

Sequence

Table 9. Robustness sequences.

S. No.	Sample	No. of injections
1	Diluent	1
2	Placebo	1
3	Standard solution	5
4	Test solution	2
5	Bracketing standard	1

Demonstrate the lustiness of check methodology by perform system quality and assay below traditional condition (i.e., methodology precision) and every of the altered conditions mentioned below [54,79].

Conditions

- Change in column temperature to $25 \pm 5^\circ\text{C}$ [7].
- Amendment in rate of flow to $1.0 \pm 0.2\text{ ml}$ [68].

Acceptance criteria

- System quality ought to be yielding [25].
- Merit RSD mustn't take issue over a pair of 0 from traditional condition study [19].

Observation

The tailing issue for pregablin was found to be within the limits.

Filter variation

Purpose

To demonstrate the filter variation of assay methodology allotted on 2 totally different filters. Perform assay on pregablin capsules 150 mg as per the check methodology, draw sample through zero.45 μm Nylon filter and zero.45 μm PVDF filter. Calculate the distinction accountable for of assay between filtered parts.

Study design

Determine the assay of those samples with totally different filters and appraise distinction pending assay between filter parts (**Table 10**).

Sequence

Table 10. Filter variation sequences.

S. No.	Sample	No. of injections
1	Diluent	1
2	Placebo	1
3	Standard solution	5
4	Sample set-1 (Nylon)	2
5	Sample set-1 (PVDF)	2
6	Bracketing standard	1

Acceptance criteria

The distinction between filtered sample solutions of various kinds of filter mustn't be over 2.0% [36].

Solution stability

Purpose

To demonstrate the steadiness of analytical solutions (i.e., commonplace and sample solution) at temperature (i.e., concerning 25°C) [58].

Study design

Prepare commonplace and sample solutions as per the check methodology and inject these solutions into HPLC system at regular intervals for minimum of 48 h, monitor the realm of each commonplace and sample solutions.

Acceptance criteria

Acceptable preference between initial and stability samples against recent commonplace isn't over 2.0%.

VALIDATION DATA

The validation data is shown from **Tables 11-18**.

Systems suitability

Table 11. System suitability of assay.

Injection	RT	Peak Area	USP Plate count	USP Tailing
1	2.71	1500112	7605	1.24
2	2.74	1507818	5820	1.27
3	2.73	1507655	5651	1.08
4	2.71	1500123	7661	1.12
5	2.72	1501285	6125	1.06
6	2.73	1506996	5658	1.22
7	2.72	1503854	6412	1.09
Mean	2.72	1503977	6418	1.15
SD	0.010135	3261.219	---	---
% RSD			---	---

Specificity

Interference from degraded products

Table 12. Interference from degraded products.

Degradation mechanism/condition	Observation
Protected sample	No interference at RT of analyte peak
Water/Reflux – 30.0 min	No interference at RT of analyte peak
Acid degradation 0.1 N HCl Reflux – 30.0 min	No interference at RT of analyte peak
Base degradation 0.01 N NaOH Reflux 30.0 min	No interference at RT of analyte peak
Peroxide degradation 3.0% H ₂ O ₂ Reflux – 30.0 min	No interference at RT of analyte peak
Thermal degradation At 105 °C - 48 h	No interference at RT of analyte peak
Photolytic degradation At 254 nm - 24 h	No interference at RT of analyte peak
Accelerated degradation At 40 °C/75% RH - 168 h	No interference at RT of analyte peak

Precision

System precision

Table 13. System Precision.

Concentration 100%	Injection	Peak Areas of pregablin
	1	1506996
	2	1508059
	3	1511449
	4	1510532
	5	1514515
Statistical Analysis	Mean	1510310
	SD	2961
	% RSD	0.20

Method precision

Table 14. Method precision.

Capsule ID	% Assay of pregablin	Statistical Analysis of pregablin	
1	100.0	Mean	99.6
2	100.0		
3	99.6	SD	0.53
4	99.4		
5	98.6	% RSD	0.532
6	100.0		

Accuracy (recovery)

Table 15. Accuracy of pregablin.

Concentration % of spiked level	Amount added (mcg/ml)	Amount found (mcg/ml)	% Recovery	Statistical Analysis of % Recovery	
50% Sample 1	748.72	745.97	99.65	MEAN	99.3
50% Sample 2	753.70	746.38	99.05	SD	0.28

50% Sample 3	751.21	745.17	99.2	%RSD	0.29
100 % Sample 1	1501.92	1495.755	99.6	MEAN	100.2
100% Sample 2	1496.94	1506.205	100.65	SD	0.48
100% Sample 3	1500.43	1504.65	100.3	%RSD	0.48
150% Sample 1	2250.64	2236.46	99.35	MEAN	99.3
150% Sample 2	2243.17	2244.93	100.10	SD	0.71
150% Sample 3	2247.15	2215.535	98.6	%RSD	0.72

Linearity

Table 16. Linearity for pregablin.

Linearity Level	Concentration ppm	Average area	% of RSD	Statistical Analysis of pregablin	
L1-50%	750.00	774302	0.19	Correlation Coefficient	0.999996
L2-60%	900.00	919946	0.27		
L3-80%	1200.00	1231232	0.10		
L4-100%	1500.00	1538305	0.18		
L5-120%	1800.00	1822882	0.21		
L6-150%	250.00	2264370	0.08		
				r ²	0.999992

Ruggedness

Table 17. Ruggedness for pregablin.

Capsule ID	% Assay of pregablin	Statistical analysis of pregablin	
1	99.1	Mean	99.1
2	99.1		
3	98.8	SD	0.2
4	99.2		
5	99.1	%RSD	0.21
6	99.4		

Robustness

Table 18. Robustness.

Parameters	Optimum range	Conditions in procedure	Remarks
Filter variation	Nylon PVDF	Ambient temp & 1 ml/min flow	

Flow rate ml/min	0.8-1.2	1.0	At lower flow rates the asymmetry factor was increased and at higher flow rates the relative retentions was decreased
Temperature	25-30°C	Ambient	Beyond the optimum range peak shape and symmetry was lost

Solution stability

Table 19. Solution stability for pregablin.

Capsule sample ID	% Assay of pregablin		Statistical Analysis of pregablin		
	For 24 h	For 48 h		For 24 h	For 48 h
1	97.4	97.5	Mean	97.5	97.3
2	97.6	97.1	SD	8522	7977
			%RSD	0.50	0.48

CONCLUSION

The test method is validated for Specificity, Linearity, Precision, Accuracy, Range, Stability of solution, Ruggedness and Robustness and found to be meeting the predetermined acceptance criteria. The validated method is Specific, Linear, Precise, Accurate, Robust and Rugged for the assay of pregablin capsules 150 mg. Hence from the above data it is concluded that the method is stability indicating.

REFERENCES

1. Tyagi A, et al. HPTLC-densitometric and RP-HPLC method development and validation for determination of salbutamol sulphate, bromhexine hydrochloride and etofylline in tablet dosage forms. Pharm Anal Acta. 2015;6:350.
2. Sonawane LV, et al. Bioanalytical method validation and its pharmaceutical application - A review. Pharm Anal Acta. 2014;5:288.
3. Mittal NK, et al. Development of harmonized bioanalytical method validation guidelines. J Bioequiv Availab. 2013;5:e39.
4. Behera S, et al. UV-Visible spectrophotometric method development and validation of assay of paracetamol tablet formulation. J Anal Bioanal Techniques. 2012;3:151.
5. Pavan KC and Gurupadayya BM. Analytical method development and validation of dimethoate pesticide using HPLC method. Biochem Anal Biochem. 2013;2:127.
6. Atanu KJ. HPLC: Highly accessible instrument in pharmaceutical industry for effective method development. Pharm Anal Acta.2012;3:147.
7. Ankit Tyagi, et.al. HPTLC-densitometric and RP-HPLC method development and validation for determination of salbutamol sulphate, bromhexine hydrochloride and etofylline in tablet dosage forms. Pharmaceut Reg Affairs 2015;6:350.
8. Xinxin Z, et al. Comparative studies on performance of CCC and preparative RP-HPLC in separation and purification of steroid saponins from *Dioscorea zingiberensis* c.h. wright. J Steroids Hormon Sci. 2015;6:150.
9. Chauhan A, et al. Analytical method development and validation: A concise review. J Anal Bioanal Tech. 2012;6:233.

10. Ranjit S. HPLC method development and validation - an overview. *J Pharm Educ Res.* 2012;4: 26-33.
11. Lu Y, et al. Development and optimization of a RP-HPLC method to quantify midazolam in rat plasma after transdermal administration: validation and application in pharmacokinetic study. *Pharm Anal Acta.* 2015;6:329.
12. Mohammad FH, et al. UV-Metric, pH-Metric and RP-HPLC methods to evaluate the multiple pka values of a polyprotic basic novel antimalarial drug lead, cyclen bisquinoline. *Mod Chem Appl* 2014, 2: 145.
13. Sultana N, et.al. Development and validation for the simultaneous quantification of prazosin, amlodipine, diltiazem and verapamil in api, dosage formulation and human serum by rp-HPLC: Application to *in vitro* interaction studies. *Med chem* 2014;4:770.
14. Naveed S. An overview of analytical determination of captopril in active pharmaceutical ingredients (API) formulation and biological fluids. *J Bioequiv Availab.* 2013;5:264-266.
15. Jenkinson C, et al. LC-MS/MS-based assay for free and deconjugated testosterone and epitestosterone in rat urine and serum. *J Anal Bioanal Tech.* 2014;S5:006.
16. Albert K, et al. Improving the understanding of the properties and retention behavior of chemically bonded stationary phases employing suspended-state HR/mas NMR spectroscopy. *J Anal Bioanal Tech.* 2013;S12:001.
17. Shah I, et al. A novel method for determination of fenofibric acid in human plasma using HPLC-UV: Application to a pharmacokinetic study of new formulations. *J Anal Bioanal Tech.* 2014; S12:009.
18. Suresh BVV, et al. Validated HPLC method for determining related substances in compatibility studies and novel extended release formulation for ranolazine. *J Chromatograph Separat Techniq.* 2014;5:209.
19. Gengaihi SEI, et al. Antioxidant activity of phenolic compounds from different grape wastes. *J Food Process Technol.* 2014;5:296.
20. Shanmugam R, et al. Bioanalytical method development and validation for herbal quercetin in nano formulation by RP-UFLC in rabbit plasma. *J Bioequiv Availab.* 2013;5:191-196.
21. Bais S, et al. Method development and validation for desogestrel and ethinylestradiol in combined pharmaceutical dosage form by RP-HPLC. *Pharm Anal Acta.* 2013;4:262.
22. Tengli AR and Gurupadaya BM. Method development and validation of tablet dosage form containing losartan, atenolol and hydrochlorthiazide using internal standard by RP-HPLC. *J Chromat Separation Techniq.* 2013;4:180.
23. Tengli AR, et al. Method development and validation of metformine, pioglitazone and glibenclamide in tablet dosage form by using RP-HPLC. *Biochem Anal Biochem.* 2013;2:130.
24. De Figueiredo NB, et al. Determination of 3,4-methylenedioxyamphetamine (MDMA) in confiscated tablets by high-performance liquid chromatography (HPLC) with diode array detector. *J Forensic Res.* 2010;1:106.
25. Pedro AQ, et al. An Improved HPLC method for quantification of metanephrine with coulometric detection. *J Chromatograph Separat Techniq.* 2014;5:217.
26. Abdallah MA. Validated stability-indicating HPLC and thin layer densitometric methods for the determination of pazufloxacin: Application to pharmaceutical formulation and degradation kinetics. *J Chromatograph Separat Techniq.* 2014;5:218.
27. Abdallah NA. HPLC and densitometric TLC methods for simultaneous determination of gemifloxacin with some co-administered drugs in human plasma. *J Chromatograph Separat Techniq.* 2014;5:220.
28. Vijaya BV, et al. Liquid chromatography/tandem mass spectrometry method for quantitative estimation of cremophor el and its applications. *J Anal Bioanal Tech.* 2013;4:163.
29. Burhenne J. Bioanalytical method validation. *J Anal Bioanal Tech.* 2012;3:e111.
30. Ramani AV, et al. Study of pharmacokinetics and tissue distribution of bits-17 in rat plasma and tissue homogenate using a validated LC method. *J Bioanal Biomed.* 2013;4:079-084.
31. Naveed S. Analytical Determination of lisinopril using UV spectrophotometer and HPLC: An overview. *Mod Chem appl.* 2014;2:137.
32. Murthy TGK and Geethanjali J. Development of a validated RP-HPLC method for simultaneous estimation of metformin hydrochloride and rosuvastatin calcium in bulk and in-house formulation. *J Chromatogr Sep Tech.* 2014;5:252.
33. Pushpa K, et al. Stability indicating RP-HPLC method development and validation of salicylic acid in choline magnesium trisalicylate (trilisate) tablets. *J Pharma Care Health Sys.* 2014;1:4.

34. Whitmire M, et al. LC-MS/MS bioanalysis method development, validation, and sample analysis: Points to consider when conducting non-clinical and clinical studies in accordance with current regulatory guidance. *J Anal Bioanal Techniques*. 2011;S4:001.
35. Nouruddin WA, et al. Development and validation of different chromatographic methods for determination of two hypouricemic drugs in their combined dosage form. *J Anal Bioanal Tech*. 2014;5:211.
36. Ravi PR, et al. Validation of a simple, rapid and sensitive LC method for quantification of riluzole in rat plasma and its pharmacokinetic application. *J Bioanal Biomed*. 2012;S6:007.
37. Lories IB, et al. High performance liquid chromatography, TLC densitometry, first-derivative and first-derivative ratio spectrophotometry for de-termination of rivaroxaban and its alkaline degradates in bulk powder and its tablets. *J Chromatograph Separat Techniq*. 2013;4:202.
38. Mohd Q, et al. DOE-based stability indicating RP-HPLC method for determination of lacidipine in niosomal gel in rat: Pharmacokinetic determination. *Pharm Anal Acta*. 2014;5:314.
39. Mathrusri AM, et al. Simultaneous determination of rosuvastatin and ezetimibe in pharmaceutical formulations by stability indicating liquid chromatographic method. *J Bioequiv Availab*. 2015;6:174-180.
40. Mathrusri AM, et al. A validated stability-indicating liquid chromatographic method for determination of cabazitaxel-a novel microtubule inhibitor. *J Bioequiv Availab*. 2014;6:134-138.
41. Apoorva VR, et al. *In vitro* metabolic stability study of new cyclen based antimalarial drug leads using RP-HPLC and LC-MS/MS. *Mod Chem Appl*. 2014;2:129.
42. Saeed AM, et al. Monitoring of pregabalin in pharmaceutical formulations and human serum using UV and RP-HPLC techniques: Application to dissolution test method. *Pharm Anal Acta*. 2014;5:287.
43. Recep K, et.al. Dyeing properties and analysis by rp-HPLC-dad of silk fabrics dyed with madder (*Rubia tinctorum* L.). *J Textile Sci Eng*. 2014;4:154 .
44. Gurupadaya BM and Disha NS. Stability indicating HPLC method for the simultaneous determination of ceftriaxone and vancomycin in pharmaceutical formulation. *J Chromat Separation Techniq*. 2013;4:207.
45. Paranthaman R and Kumaravel SA. Reversed-phase high-performance liquid chromatography (RP-HPLC) determination of pesticide residues in tender coconut water (elaneer/nariyal pani). *J Chromat Separation Techniq*. 2013;4:208.
46. Lories IB, et al. High performance liquid chromatography, TLC densitometry, first derivative and first-derivative ratio spectrophotometry for de-termination of rivaroxaban and its alkaline degradates in bulk powder and its tablets. *J Chromat Separation Techniq*. 2013;4:202.
47. Nada S, et al. Determination of thiomersal, lidocaine and phenylepherine in their ternary mixture. *J Chromat Separation Techniq*. 2013;4:199.
48. Najmul H, et al. Simultaneous determination of ns aid and antimicrobial preservatives using validated RP-HPLC method: An application in pharmaceutical and clinical laboratories. *Pharm Anal Acta*. 2013;4:263.
49. Muhammad SA and Shabana NS. Liquid chromatographic analysis of prazosin in API, dosage form and serum: Application to drug-metal interaction studies. *J Chromat Separation Techniq*. 2013;4:197.
50. Anil C, et al. Method development and validation for desogestrel and ethinylestradiol in combined pharmaceutical dosage form by RP-HPLC. *Pharmaceut Anal Acta*. 2013;4:262.
51. Aparna G, et al. ACE-inhibitory activity of cheddar cheeses made with adjunct cultures at different stages of ripening. *J Adv Dairy Res*. 2013;1:102.
52. Najma S, et al. RP-HPLC method for the simultaneous determination of captopril and H₂-receptor antagonist: Application to interaction studies. *Med chem*. 2013;3:183.
53. Anandkumar R, et al. Method development and validation of metformine, pioglitazone and glibenclamide in tablet dosage form by using RP-HPLC. *Biochem Anal Biochem*. 2013;2:130.
54. Bhatt KK, et al. Development of a validated stability-indicating RP-HPLC method for dronedarone hydrochloride in pharmaceutical formulation. *J Anal Bioanal Tech*. 2012;4:161.
55. Bhatt KK, et al. Simultaneous estimation of pregabalin and methylcobalamine in pharmaceutical formulation by RP-HPLC method. *J Anal Bioanal Tech*. 2012;4:159.
56. QingYi Lu, et al. Determination of rottlerin, a natural protein kinases c inhibitor, in pancreatic cancer cells and mouse xenografts by RP-HPLC method. *J Chromat Separation Techniq*. 2012;4:162.
57. Keyur BA, et al. Simultaneous estimation of tramadol HCl, paracetamol and domperidone in pharmaceutical formulation by RP-HPLC method. *J Chromat Separation Techniq*. 2012;3:152.
58. Aimen AEA, et al. Development and validation of analytical method by RP-HPLC for quantification of alpha-mangostin encapsulated in PLGA microspheres. *J Anal Bioanal Techniques*. 2012;3:155.
59. Gnana RM, et al. Simultaneous, stability indicating method development and validation for related compounds of ibuprofen and paracetamol tablets by RP-HPLC method. *J Chromat Separation Techniq*. 2012;3:155.

60. Mehul MP and Disha DP. Simultaneous estimation of metoprolol succinate and olmesartan medoxomil in pharmaceutical dosage form by rp-HPLC. *J Chromat Separation Techniq.* 2012;3:151.
61. Paranthaman R, et.al. GC-MS analysis of phytochemicals and simultaneous determination of flavonoids in *Amaranthus caudatus* (sirukeerai) by RP-HPLC. *J Anal Bioanal Techniques.* 2012;3:147.
62. Nouruddin WA, et al. Validated chromatographic methods for simultaneous determination of amlodipine besylate and perindopril arginine in binary mixtures and in pharmaceutical dosage form. *J Chromat Separation Techniq.* 2012;3:134.
63. Jaya PK and Syama SB. Development and validation of an HPLC method for quantifying dapiprazole in bulk preparations. *J Anal Bioanal Techniques.* 2012;3:143.
64. Kumar P, et al. Stability indicating method development for simultaneous estimation of ezetimibe and atorvastatin in pharmaceutical formulations by RP-HPLC. *Pharm Anal Acta.* 2012;3:164.
65. Seemi S, et al. RP-HPLC Method for estimation and stress degradation study of paclitaxel as per ICH guidelines. *J Chromat Separation Techniq.* 2012;3:135.
66. Jagadeeswaran M, et.al. Quantitative estimation of lopinavir and ritonavir in tablets by RP-HPLC method. *Pharm Anal Acta.* 2012;3:160.
67. Smita TK, et al. Simultaneous estimation of fluoxetine and norfluoxetine in plasma by RP-HPLC employing pre-column derivatization for UV-sensitivity enhancement. 2012;S008.
68. Chandrashekhar N, et.al. Simultaneous estimation of aceclofenac, paracetamol and tizanidine in their combined dosage forms by spectrophotometric and RP-HPLC method. *J Anal Bioanal Techniques.* 2011;2:123.
69. Najma S, et al. RP-HPLC method for simultaneous determination of captopril and diuretics: Application in pharmaceutical dosage forms and human serum. *J Chromatograph Separat Techniq.* 2011;2:109.
70. Subramanian N, et al. Improved RP-HPLC method for the simultaneous estimation of tranexamic acid and mefenamic acid in tablet dosage form. *Pharm Anal Acta.* 2011;2:115.
71. Vishnu P, et al. Simultaneous estimation of atorvastatin, ezetimibe and fenofibrate in pharmaceutical formulation by RP-LC-PDA. *Pharm anal acta.* 2010;1:111.
72. Rama SP, et al. Method Development and Validation for estimation of moxifloxacin HCl in tablet dosage form by RP-HPLC method. *Pharm Anal Acta.* 2010;1:109.
73. Lalit VS and Sanjaykumar B. Bari development and validation of RP-HPLC method for the simultaneous estimation of amoxicillin trihydrate and bromhexine hydrochloride from oily suspension. *Pharm Anal Acta.* 2010;1:107.
74. Laxma S, et al. Quantitative HPLC analysis of ascorbic acid and gallic acid in *Phyllanthus emblica*. *J Anal Bioanal Techniques.* 2010;1:111.
75. Sohan SC, et al. Development and validation of spectrophotometric and HPLC method for the simultaneous estimation of salbutamol sulphate and prednisolone in tablet dosage form. *J Anal Bioanal Techniques.* 2011;2:117.
76. Mukesh M and Ranjit S. Development and validation of a stability-indicating HPLC method for the simultaneous determination of salbutamol sulphate and theophylline in pharmaceutical dosage forms. *J Anal Bioanal Techniques.* 2011;2:116.
77. Nevado JJB, et al. Reliable and sensitive SPE-HPLC-dad screening of endocrine disruptor's atrazine, simazine and their major multiresidues in natural surface waters: Analytical validation and robustness study performance. *J Chromatograph Separat Techniq.* 2014;5:215.
78. Shintani H and Hayashi F. Analytical validation of ameziniummetilsulfate by HPLC in human blood plasma from uremia patient treated by dialysis. *Pharm Anal Acta.* 2011;S11:004.
79. Nia Y, et al. Determination of Ti from TiO₂ nanoparticles in biological materials by different ICP-MS instruments: Method validation and applications. *J Nanomed Nanotechnol.* 2015;6:269.
80. Dare M, et al. Method validation for stability indicating method of related substance in active pharmaceutical ingredients dabigatran etexilate mesylate by reverse phase chromatography. *J Chromatogr Sep Tech.* 2015;6:263.
81. Ampos S, et al. Simultaneous quantification of propofol and its non-conjugated metabolites in several biological matrices using gas chromatography/ion trap mass spectrometry method. *J Anal Bioanal Tech.* 2014;5:195.
82. Asha I. Identification, estimation & determination of residual solvents of Olanzapine in bulk & formulation by HPLC, GC & IR. *Pharmaceut Anal Acta.* 2013;4:2.
83. Hafez HM. Quantitative determination of amlodipine besylate, losartan potassium, valsartan and atorvastatin calcium by HPLC in their pharmaceutical formulations. *Pharm Anal Acta.* 2014;5:300.

84. Antil P. UPLC method for simultaneous determination of valsartan & hydrochlorothiazide in drug products. *J Chromat Separation Techniq.* 2013;4:182.
85. Schindera C, et al. Early development of arterial hypertension in an infant with valsartan fetopathy. *J Neonatal Bio.* 2012;1:103.
86. Sunkara G, et al. Assessment of ethnic differences in the pharmacokinetics and pharmacodynamics of valsartan. *J Bioequiv Availab.* 2010;2:120-124.
87. Nevado JJB, et al. Reliable and sensitive SPE-HPLC-dad screening of endocrine disruptor's atrazine, simazine and their major multiresidues in natural surface waters: Analytical validation and robustness study performance. *J Chromatograph Separat Techniq.* 2014;5:215.
88. Anbumathi P, et al. Quantitative analysis of a dynamic cell cycle regulatory model of *Schizosaccharomyces pombe*. *Curr Synthetic Sys Biol.* 2013;1:105.
89. Schrum AG and Gil D. Robustness and specificity in signal transduction via physiologic protein interaction networks. *Clin Exp Pharmacol.* 2013;S3:001.
90. Passe U. The next challenges ahead: Design integration and robustness. *J Archit Eng Tech.* 2012;1:e105.
91. Li H and Wang I. consistent estimation in generalized linear mixed models with measurement error. *J Biomet Biostat.* 2012;S7:007.
92. Fayyad MK, et al. Effect of temperature, wavelength, pH, ion pair reagents and organic modifiers' concentration on the elution of *cystatin c*. stability of mobile phase. *J Anal Bioanal Tech.* 2010;1:103.
93. Androniki T. Evaluation of pigmented skin lesions with optical computed tomography. *J Clin Exp Dermatol Res.* 2014;5:2.
94. Anju A, et al. Novel mass spectrometry method for the quantification of immuno-suppressant drug in human whole blood. *J Anal Bioanal Techniques.* 2012;3:7.
95. Sravani U, et al. Derivative spectrophotometric methods for the determination of Zolpidem Tartrate. *J Bioequiv Availab.* 2012;4:3.
96. Saketha CN, et al. Spectrophotometric methods for the determination of zolpidem tartrate in acetate buffer. *Bioequiv Availab.* 2012;4:3.
97. Hima BG, et al. Analytical method development for the determination of olopatadine. *J Bioequiv Availab.* 2012;4:3.
98. Divya, et al. Derivative spectrophotometric method for the determination of olopatadine. *J Bioequiv Availab.* 2012;4:3.
99. Amrutha SV and Mathrusri AM. Derivative spectrophotometric method for the determination of moxifloxacin in pharmaceutical formulations. *J Bioequiv Availab.* 2012;4:3.
100. Igarashi H, et al. Characteristic features of an analytical column with a pentafluorophenylpropyl stationary phase applied to a determination of a fluorinated phenyl alanyl derivative compound, gw823093, in human urine using an LC-ESI-MS/MS method. *J Anal Bioanal Techniques.* 2011;S5:001.
101. An LTT, et al. Statistical analysis of protein microarray data: a case study in type 1 diabetes research. *J proteomics Bioinform.* 2014;S12:003.