

# Research and Reviews: Journal of Botanical Science

## Bio Fertilizers Equipment's and its Types

Shabana Sultana\*

Department of Biotechnology, Andhra University, Andhra Pradesh, India

### Review Article

#### ABSTRACT

Received: 25/09/2016

Revised: 01/10/2016

Accepted: 07/10/2016

#### \*For Correspondence

Shabana Sultana, Department of Biotechnology,  
Andhra University, Andhra Pradesh, India, Tel:  
9848033228.

E-mail: [Shabanasultana2@gmail.com](mailto:Shabanasultana2@gmail.com).

**Keywords:** Fertilizers, Soil, Equipment, Microbes,  
Plants

Biofertilizers are characterized as arrangements containing living cells or inert cells of productive strains of microorganisms that harvest plants uptake of supplements by their communications in the rhizosphere when connected through seed or soil. They quicken certain microbial procedures in the dirt which expand the degree of accessibility of supplements in a structure effortlessly absorbed by plants.

Regularly microorganisms are not as effective in normal surroundings as one would anticipate that they will be and hence falsely increased societies of proficient chose microorganisms assume an indispensable part in quickening the microbial procedures in soil.

#### INTRODUCTION

##### Types of Bio Fertilizers

Rhizobium, Azospirillum, Cyanobacteria, Azolla, liquid biofertilizers, Azotobacter, Acetobacter.

##### Rhizobium

Rhizobium is a dirt living space bacterium, which can ready to colonize the vegetable roots and fixes the environmental nitrogen advantageously. The morphology and physiology of Rhizobium will change from free-living condition to the bacteroid of knobs. They are the most productive biofertilizer according to the amount of nitrogen settled concerned [1-10]. They have seven genera and profoundly particular to shape knob in vegetables, alluded as cross immunization bunch.

##### Azospirillum

*Azospirillum lipoferum* and *A. brasilense* are essential tenants of soil, the rhizosphere and intercellular spaces of root cortex of graminaceous plants. They play out the cooperative harmonious connection with the graminaceous plants. The microbes of genus *Azospirillum* are N<sub>2</sub> altering creatures disconnected from the root or more ground parts of an assortment of yield plants. They are Gram negative, Vibrio or Spirillum having bounteous amassing of polybetahydroxybutyrate in cytoplasm. Five types of *Azospirillum* have been portrayed to date *A. brasilense*, *A. lipoferum*, *A. amazonense*, *A. halopraeferens* and *A. irakense*. The life form multiplies under both anaerobic and oxygen consuming conditions yet it is especially small scale aerophilic in the nearness or nonappearance of joined nitrogen in the medium [11-20].

### Cyanobacteria

Both free-living and additionally cooperative cyanobacteria have been saddled in rice development in India. A composite society of BGA having heterocystous Nostoc, Anabaena, Aulosira and so forth is given as essential inoculum in plate, polythene lined pots and later mass duplicated in the field for application as soil based chips to the rice developing field at the rate of 10 kg/ha. The last item is not free from unessential contaminants and not regularly observed for checking the nearness of desired algal vegetation.

When such a great amount of announced as a biofertilizer for the rice crop, it has not in the blink of an eye pulled in the consideration of rice producers all over India aside from pockets in the Southern States, quite Tamil Nadu. The advantages because of algalization could be to the degree of 20 kg N/ha to 30 kg N/ha under perfect conditions yet the work arranged procedure for the planning of BGA biofertilizer is in itself a restriction.

### Azolla

Azolla is a free-drifting water greenery that buoys in water and fixes barometrical nitrogen in relationship with nitrogen altering blue green alga *Anabaena azollae*. Azolla fronds comprise of sporophyte with a coasting rhizome and little covering bi-lobed leaves and roots. Rice developing ranges in South East Asia and other underdeveloped nations have as of late been displaying expanded enthusiasm for the utilization of the advantageous N<sub>2</sub> settling water plant Azolla either as another nitrogen sources or as a supplement to business nitrogen manures. Azolla is utilized as biofertilizer for wetland rice and it is known not 40 kg N/ha to 60 kg N/ha per rice crop.

### Liquid Biofertilizers

Biofertilizers are such as Rhizobium, *Azospirillum* and Phosphobacteria provide nitrogen and phosphorous nutrients to crop plants through nitrogen fixation and phosphorous solubilization processes. These Biofertilizers could be effectively utilized for rice, pulses, millets, cotton, sugarcane, vegetable and other horticulture crops [21-30].

Biofertilizers is one of the prime input in organic farming not only enhances the crop growth and yield but also improves the soil health and sustain soil fertility.

### Azotobacter

It is the imperative and surely understood free living nitrogen settling high-impact bacterium. It is utilized as a bio-Fertilizer for all non-leguminous plants particularly rice, cotton, vegetables and so on. Azotobacter cells are not present on the rhizosphere but rather are copious in the rhizosphere district [31-40]. The absence of natural matter in the dirt is a constraining element for the multiplication of Azotobacter in the dirt.

Field analyses were directed in 1992, 1993 and 1994 amid the pre-kharif wet seasons to discover the impact on rice grain yield by the consolidated utilization of N-settling living beings and inorganic nitrogen manure which recorded increment in was yield.

### Acetobacter

This is a sacharophilic microbes and partner with sugarcane, sweet potato and sweet sorghum plants and fixes 30 kgs/N/ha year. Basically this bacterium is marketed for sugarcane crop. It is known not yield by 10 t/section to 20 t/section of land and sugar content by around 10% to 15%.

### **Bio Fertilizer Equipments**

In biofertilizer creation industry, types of gear are the significant foundation, which includes 70% of capital speculation. Any bargain on the use of the accompanying said types of gear may at long last decrease in the nature of biofertilizer. After examining the standard behind the use of all instruments, a portion of the instruments can be supplanted with a society room fitted with a U.V. lamp. Autoclaves, hot air oven, incubators and fixing machines are indigenously made with appropriate specialized particulars [41-50]. The right utilization of types of gear will give continuous presentation with quality inoculum. Some of the biofertilizer equipments are as follows.

#### **Autoclave**

It is a contraption in which materials are cleaned via air free immersed steam at a temperature above 100°C. On the off chance that the steam weight inside the autoclave is expanded to 15 psi, the temperature will ascend to 121°C. this is adequate to decimate every single vegetative cell. Regularly all development medium is sanitized in the autoclave [51-60].

#### **Laminar Air Flow Chamber**

Laminar wind stream chamber gives a uniform stream of sifted air. This nonstop stream of air will counteract settling of particles in the work area. Air borne tainting is evaded in this chamber. Society exchanges and vaccination should be possible here.

#### **BOD Incubators**

Hatcheries giving controlled conditions required for the development and improvement of microorganisms. Duplication of starter society should be possible in this instrument [61-70].

#### **Rotary Shaker**

It is utilized for disturbing society jars by roundabout movement under variable pace control. Shaking gives air circulation to development of societies. Shakers holding upto 20 to 50 cups are by and large utilized. The limit of the shaker might be expanded in the event that it is a twofold decker sort [71-80].

#### **Refrigerator**

This gear is utilized protecting all mother societies utilized for biofertilizer creation. The mother society is intermittently sub-refined and put away in the cooler for long haul use [81-90].

#### **Fermentor**

A fermentor is the gear, which gives the best possible environment to the development of a craved life form. It is for the most part an extensive vessel in which, the life form might be kept at the required temperature, pH, broke up oxygen fixation and substrate focus. Distinctive models of fermentors are accessible relying on the need [91-100]. A straightforward adaptation model contains steam generator, disinfection process gadgets and instigator.

## **REFERENCES**

1. Farfour SA and Al-Saman MA. Root-rot and stem-canker control in faba bean plants by using some biofertilizers agents. *J Plant Pathol Microb.* 2014;5:218.
2. Vinale F. Biopesticides and biofertilizers based on fungal secondary metabolites. *J Biofertil Biopestici.* 2014;5:e119.
3. Raja N. Biopesticides and biofertilizers: Ecofriendly sources for sustainable agriculture. *J Biofertil Biopestici.* 2013;4:e112.
4. Paul N, et al. Evaluation of biofertilizers in cultured rice. *J Biofertil Biopestici.* 2013;4:133.
5. Brar SK, et al. Shelf-life of biofertilizers: An accord between formulations and genetics. *J Biofertil Biopestici.* 2012;3:e109.
6. Balachandar D. Biofertilizers-what next? *J Biofertil Biopestici.* 2012;3:e108.
7. Pandit NP, et al. Vermicomposting biotechnology: An eco-loving approach for recycling of solid organic wastes into valuable biofertilizers. *J Biofertil Biopestici.* 2012;2:113.
8. Densilin DM, et al. Effect of individual and combined application of biofertilizers, inorganic fertilizer and vermicompost on the biochemical constituents of chilli (Ns-701). *J Biofertil Biopestici.* 2011;2:106.
9. Jhala YK, et al. Efficacy testing of acetobacter and azospirillum isolates on maize cv. GM-3. *J Fertil Pestic.* 2016;7:164.
10. Pandiarajan G, et al. Exploration of different azospirillum strains from various crop soils of srivilliputtur taluk. *J Biofertil Biopestici.* 2012;3:117.
11. Abd-El-Karem Y, et al. Effect of cyanophycin metabolism in recombinant sinorhizobium (ensifer) meliloti 1021 on the symbiosis with alfalfa (*Medicago sativa*). *J Microb Biochem Technol.* 2016;8:375-381.
12. Vanek SJ, et al. Pore-Size and water activity effects on survival of rhizobium tropici in biochar inoculant carriers. *J Microb Biochem Technol.* 2016;8:296-306.
13. Farooqi MS, et al. Genome-wide relative analysis of codon usage bias and codon context pattern in the bacteria *salinibacter ruber*, *chromohalobacter salexigens* and *rhizobium etli*. *Biochem Anal Bioche.* 2016.
14. Abdel-lateif Khalid S, et al. Monitoring of molecular variation among egyptian faba bean rhizobium isolates as response to pesticides stress. *J Bioremed Biodeg.* 2015;6:296.
15. Kefela T, et al. *Paenibacillus polymyxa*, *Bacillus licheniformis* and *Bradyrhizobium japonicum* IRAT FA3 promote faster seed germination rate, growth and disease resistance under pathogenic pressure. *J Plant Biochem Physiol.* 2015;3:145.
16. Elzanaty AM, et al. Molecular and biochemical characterization of some Egyptian genotypes *Rhizobium* (*Vicia Faba*) Isolates. *J Bioengineer & Biomedical Sci.* 2015;5:145
17. Chauhan A. *Sinorhizobium meliloti* bacteria contributing to rehabilitate the toxic environment. *J Bioremed Biodeg.* 2015;6:e164.
18. El-Zanaty AF, et al. Molecular Identification of *Rhizobium* Isolates nodulating faba bean plants in Egyptian soils. *J Bioprocess Biotech.* 2014;5:194
19. Lopes EM, et al. Emulsification properties of bioemulsifiers produced by wild-type and mutant *bradyrhizobium elkanii* strains. *J Bioremed Biodeg.* 2014;5: 245.
20. Messele B and Pant LM. Effects of Inoculation of *Sinorhizobium ciceri* and phosphate solubilizing bacteria on nodulation, yield and nitrogen and phosphorus uptake of chickpea (*Cicer arietinum* L.) in shoa robit area. *J Biofertil Biopestici.* 2012;3:129.
21. El-Halmouch Y, et al. The potential of cell-free cultures of *rhizobium leguminosarum*, *azotobacter chroococcum* and compost tea as biocontrol agents for faba bean broomrape (*Orobanche crenata* Forsk.). *J Plant Pathol Microb.* 2013;4:205.
22. Tersagh I, et al. Aerobic degradation of fluoranthene, benzo(b)fluoranthene and benzo(k)fluoranthene by aerobic heterotrophic bacteria- cyanobacteria interaction in brackish water of bodo creek. *J Pet Environ Biotechnol.* 2016;7:292.
23. Ichor T, et al. Time Dependent Influence of Aerobic Heterotrophic Bacteria-Cyanobacteria Interaction during Biodegradation of Poly Aromatic Hydrocarbons. *J Pet Environ Biotechnol.* 2016;7: 286.
24. Ansari MKA, et al. Evaluation of methyl red tolerant cyanobacteria for simultaneous laccase production and dye decolorization. *Int J Waste Resour.* 2016;6:221.
25. Yunes JS, et al. Identification of the toxic pentapeptide nodularin in a cyanobacterial bloom in a shrimp farm in south american atlantic coast. *Pharm Anal Acta.* 2016;7:479.

26. Anas A, et al. Heavy metals pollution influence the community structure of cyanobacteria in nutrient rich tropical estuary. *Oceanography*. 2015;3:137.
27. Menamo M and Wolde Z. (2015) The potential of cyanobacteria growth in different water sources. *Biochem Physiol*. 2015;4:164.
28. Kilulya KF, et al. Extraction procedures and gcxgc-tofms determination of fatty acids (fas) in cyanobacteria cultures and the effect of growth media iron concentration variation on cellular fas composition. *J Environ Anal Toxicol*. 2015;S7:009.
29. Sakthivel K and Kathiresan K. Cholesterol degradation effect analyzed using marine cyanobacterial species spirulina subsalsa. *J Microb Biochem Technol*. 2015;7:120-123.
30. Rastogi PR and Madamwar D. UV-Induced oxidative stress in cyanobacteria: how life is able to survive? *Biochem Anal Biochem*. 2015;4:173.
31. Yasser ELN and Adli A. Toxicity of single and mixtures of antibiotics to cyanobacteria. *J Environ Anal Toxicol*. 2015;5:274.
32. David Noel S and Rajan MR. Cyanobacteria as a potential source of phycoremediation from textile industry effluent. *J Bioremed Biodeg*. 2014;5:260.
33. Vaithiyalingam SU, et al. Biocalcification mediated remediation of calcium rich ossein effluent by filamentous marine cyanobacteria. *J Bioremed Biodeg*. 2014;5:257.
34. Zagatto PA and Ferrão-Filho AdS. Acute effects of a cylindrospermopsis raciborskii (Cyanobacteria) strain on mouse, daphnia and fish. *J Ecosyst Ecogr*. 2012;2:121.
35. Vijayakumar S and Manoharan C. Treatment of dye industry effluent using free and immobilized cyanobacteria. *J Bioremed Biodeg*. 2012;3:165.
36. Vijayakumar S. potential applications of cyanobacteria in industrial effluents-a review. *J Bioremed Biodeg*. 2012;3:154.
37. El-Sheekh MM, et al. Biodegradation of phenolic and polycyclic aromatic compounds by some algae and cyanobacteria. *J Bioremed Biodegrad*. 2011;2:133.
38. Kannan V, et al. Bioremediation of tannery effluents by filamentous cyanobacteria anabaena flos-aquae west. *Hydrol Current Res*. 2011;2:122.
39. Olubunmi AO and Bernard EO. Bioremediation of soil contaminated with tannery effluent by combined treatment with cow dung and microorganisms isolated from tannery effluent. *J Bioremed Biodeg*. 2016;7:354.
40. Kalayci S. Antimicrobial properties of various non-antibiotic drugs against microorganisms. *J Bioanal Biomed*. 2016;8:e142.
41. Diop MB, et al. Efficiency of neutralized antibacterial culture supernatant from bacteriocinogenic lactic acid bacteria supplemented with salt in control of microorganisms present in Senegalese artisanally handled fish by immersion preservative technology during guedj seafood processing at 10°C and 30°C. *J Food Microbiol Saf Hyg*. 2016;1:102.
42. Nwidi IC and Agunwamba JC. Kinetics of biosorption of three heavy metals by five free microorganisms. *J Bioremed Biodeg*. 2016;7:339.
43. Xenia ME and Refugio RV. Microorganisms metabolism during bioremediation of oil contaminated soils. *J Bioremed Biodeg*. 2016;7:340.
44. Wang R. The influence of microorganisms on the contemporary society. *Med Aromat Plants*. 2016;5:e171.
45. Xiong ZQ. Bioprospecting of uncultured microorganisms: The dawning of antibiotic discovery. *Clin Microbiol*. 2016;5:e132.
46. Nouha K, et al. EPS producing microorganisms from municipal wastewater activated sludge. *J Pet Environ Biotechnol*. 2015;7:255.
47. Owhonka A and Gideon OA. The role of aerobic microorganisms in the biodegradation of petroleum hydrocarbons laboratory contaminated groundwater. *Fermentol Techno*. 2015;4:122.
48. Caruso G. Plastic degrading microorganisms as a tool for bioremediation of plastic contamination in aquatic environments. *J Pollut Eff Cont*. 2015;3:e112.
49. Mustapha MU and Halimoon N. Microorganisms and biosorption of heavy metals in the environment: a review paper. *J Microb Biochem Technol*. 2015;7:253-256.
50. Minic Z. Proteomic studies of the effects of different stress conditions on central carbon metabolism in microorganisms. *J Proteomics Bioinform*. 2015;8:080-090.

51. Kulakovskaya T. Phosphorus storage in microorganisms: Diversity and evolutionary insight. *Biochem Physiol.* 2015;4:e130.
52. Zanini SF, et al. Use of antimicrobials from plants in feed as a control measure for pathogenic microorganisms. *J Microb Biochem Technol.* 2015;7:248-252.
53. Pandey S. Study of preliminary phytochemical screening and antibacterial activity of *tribulus terretris* against selected pathogenic microorganisms. *J Bioanal Biomed.* 2014;S12:001.
54. Ky Karna Radjasa. Marine Fungi: The untapped diversity of marine microorganisms radjasa OK, *J Coast Zone Manag.* 2015;18:e110.
55. Watanabe M and Kawai F. Study on role of microorganisms in depolymerization processes of xenobiotic polymers. *J Environ Anal Chem.* 2014;2:119.
56. Momand L, et al. Antimicrobial effect of *Baccaurea angulata* fruit extracts against human pathogenic microorganisms. *Med Aromat Plants.* 2014;3:172.
57. Sandhya S, et al. Solar light induced photo catalytic disinfection of gram positive and negative microorganisms from water with highly efficient AuTiO<sub>2</sub> nanoparticle. *J Bioprocess Biotech.* 2014;4:176.
58. Shintani H. How to recovery of damaged microorganisms by supplying several sorts of nutrients. *J Bioanal Biomed.* 2014;6:024-028.
59. Samendra PS, et al. Rapid detection technologies for monitoring microorganisms in water. *Biosens J.* 2014;3:109.
60. La Fauci V, et al. The possible role of mobile phones in spreading microorganisms in hospitals. *J Microb Biochem Technol.* 2014;6:334-336.
61. Wainwright M, et al. Recovery of cometary microorganisms from the stratosphere. *Astrobiol Outreach.* 2014;2:110.
62. Chang S, et al. The sheep in wolf's clothing: Vegetable and fruit particles mimicking cells and microorganisms in cytology specimens. *J Cytol Histol.* 2013;5:207.
63. Ramakrishnan B. Fuelling the microorganisms for remediation. *J Bioremed Biodeg.* 2013;4:e139.
64. Shintani H. Rapid assay of airborne microorganisms and bioburden using several procedures. *Pharm Anal Acta.* 2013;4:246.
65. Kulshreshtha S. Genetically engineered microorganisms: A problem solving approach for bioremediation. *J Bioremed Biodeg.* 2013;4:e133.
66. Abd H, et al. Survival of vibrio cholerae inside acanthamoeba and detection of both microorganisms from natural water samples may point out the amoeba as a protozoal host for V. cholerae. *J Bacteriol Parasitol.* 2011;S1-003.
67. Ricke SC. Probiotic gastrointestinal microorganisms: Current and future prospects. *J Prob Health.* 2013;1:e102.
68. Shah MP, et al. Optimization of environmental parameters on microbial degradation of reactive black dye. *J Bioremed Biodeg.* 2013;4:183.
69. Shintani H. Importance considering increased recovery of injured microorganisms to attain reproducible sterilization validation. *Pharmaceut Anal Acta.* 2013;4:210.
70. Goda SK. DNA shuffling and the production of novel enzymes and microorganisms for effective bioremediation and biodegradation process. *J Bioremed Biodeg.* 2012;3:e116.
71. Oladele and Olakunle O. Microorganisms associated with the deterioration of fresh leafy indian spinach in storage. *J Plant Pathol Microbiol.* 2011;2:110.
72. Mesapogu S, et al. Rapid detection and quantification of *Fusarium udum* in soil and plant samples using realtime PCR. *J Plant Pathol Microbiol.* 2011;2:107
73. Chellapandi P, et al. Systems biotechnology: An emerging trend in metabolic engineering of industrial microorganisms. *J Comput Sci Syst Biol.* 2010;3:043-049.
74. Radonic A, et al. Anionic polysaccharides from phototrophic microorganisms exhibit antiviral activities to vaccinia virus. *J Antivir Antiretrovir.* 2010;2:051-055.
75. Chen C, et al. Biological control of *Fusarium* wilt on cotton by use of endophytic bacteria. *Biol Control.* 1995;5:83-91.
76. Park KS and Kloepper JW. Activation of PR-1a promoter by rhizobacteria which induce systemic resistance in tobacco against *Pseudomonas syringae* pv. *tabaci*. *Biol Control.* 2000;18:2-9

77. Chuaboon W, et al. Biological analysis of *Pseudomonas fluorescens* SP007s induced systemic resistance in sweet corn against bacterial leaf streak. In: Proceedings of the 1st Int. Conf. on Corn and Sorghum Research, Chonburi. 2009;206-215.
78. Marleny BC, et al. Suppressiveness of root- knot nematodes mediated by rhizobacteria. *Biological control*. 2008;47:55-59.
79. Kloepper JW and Ryu CM. Bacterial endophytes as elicitors of induced systemic resistance. In: Schulz B, Boyle C, Sieber T (Eds.), *Microbial Root Endophytes*. Springer-Verlag, Heidelberg. 2006;33-51.
80. Lucy M, et al. Applications of free living plant growth-promoting rhizobacteria. *Antonie Van Leeuwenhoek*. 2004;86:1-25
81. Yehia AH, et al. Biological seed treatment to control Fusarium root rot of broad bean. *Egypt J Phytopathol*. 1988;4:59-66.
82. Nelsson EB. Rapid germination of sporangia of *Pythium* species. *Phytopathology*. 1992;77:1108-1112.
83. Ehteshamul-Haque S and Gaffar A. Use of rhizobia in the control of root rot diseases of sunflower, okra, soybean and mung bean. *J Phytopathol*. 1993;138:157-163.
84. Zheng XY and Sinclair JB. The effect of traits of *Bacillus megaterium* on seed and root colonization root rot of soybean. *Biocontrol*. 2000;45:223-243.
85. Lewis JA and Lundsden RD. Biocontrol of damping off greenhouse grown crops caused by *Rhizoctonia solani* with a formulation of *Trichoderma* spp. *Crop Protection*. 2001;22: 49-56.
86. Estevez de Jensen C, et al. Integrated management strategies of bean root rot with *Bacillus subtilis* and *Rhizobium* in Minnesota. *Field Crops Researches*. 2002;74:107-115.
87. Santamarina MP and Josepha R. Influence of temperature and water activity on the antagonism of *Trichoderma harzianum* to *Verticillium* and *Rhizoctonia*. *Crop Protection*. 2006;5:110-121.
88. Hansen B, et al. Approaches to assess the environmental impact of organic farming with particular regard to Denmark. *Agr Eco Env*. 2001;83:11-26.
89. Van Diepenengen AD, et al. Effect of organic versus conventional management on chemical and biological parameters in agricultural soil. *AgrEcosys Environ*. 2006;31:120-135.
90. Lombardi-Boccia G, et al. Nutrients and antioxidant molecules in yellow plums (*Prunus domestica* L.) from conventional and organic productions: a comparative study. *J Agric Food Chem*. 2004;52:90-94
91. Manach C, et al. Polyphenol-rhizobial inoculants had been used in legume fields. *Chem&Eng Tech*. No. 24- Life Science. 2004;1: 217-223.
92. Saber MS M. Clean biotechnology for sustainable farming. *Engineering in Life Science*. 2001;1: 217-223.
93. Ran H, et al. Antioxidative and antimicrobial activities and flavonoids content of organically cultivated vegetables. *Nippon, Shokuhin Kagaku Kaishi*. 2001;48: 246-252.
94. Dinitrios B. Sources of natural phenolic antioxidants. *Trends Food SCI TECH* 2006;17:505-512.
95. Asami DK, et al. Comparison of the total phenolic and ascorbic acid content of freeze-dried and air-dried marionberry, strawberry, and corn grown using conventional, organic, and sustainable agricultural practices. *J Agric Food Chem*. 2003;51: 1237-1241.
96. Dave N, et al. *Trichoderma harzianum* elicits defense response in *Brassica juncea* plantlets. *Int Res J Biological Science*. 2013;2:1-10
97. Shores M, et al. Induced systemic resistance and plant responses to fungal biocontrol agents. *Annu Rev Phytopathol*. 2010;48:21-43.
98. Khan J, et al. Systemic resistance induced by *Trichoderma hamatum* 382 in cucumber against *Phytophthora* crown rot and leaf blight. *Plant Disease*. 2004;88:280-286.
99. Singleton VL and Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am J EnolViticul*. 1965;16:144-158.
100. Blois MS. Antioxidant determinations by the use of a stable free radical. *Nature*. 1958;26:1199-1200.