**Research Article** 

# GC-MS Analysis and Phytochemical Screening of A Rare Pteridophyte *Nephrolepis Cardifolia* (L.) Presl. from Tirumala Hills

#### K. Rukmini, \*P. Suvarnalatha Devi

Department of Applied Microbiology, Sri Padmavathi Mahila Visvavidyalayam, Tirupathi-517502, Andhra Pradesh, India.

#### ABSTRACT

Plants have been an important source of medicine with qualities for thousands of years. Mainly on traditional remedies such as ferns for their history it has been used as a popular folk medicine. Nephrolepis cardifolia(L.) Presl. of family Nephrolepidaceae found in hilly area of Tirumala hills, Tirupathi, Andhra Pradesh, India. Ethanomedicinally, the genus is important and popularly known as Fishbone fern. The present paper reveals the phytochemical screening and GC-MS analysis of Nephrolepis cardifolia. Preliminary phytochemical studies showed the presence of alkaloids, flavonoids, phenols, tannins and saponins in higher to moderate concentrations. Acetone extract of the plant was analyzed using Gas Chromatography-Mass Spectrometry, while the mass spectra of the compounds found in the extract was matched with the National Institute of Standards and Technologies (NIST) library. Gas chromatography mass spectrometry (GC-MS) analysis revealed the presence of six compounds. The phytoconstituents screened were Neophytadiene and 2,6,10-trimethyl,14,ethylene,2 Hexadecen-1-ol and 3,7,11,15 tetramethyl R-R, hexadecanoic acid & palmitic acid, 9 octadecenoic acid(z) & oleic acid, octadecanoic acid & stearic acid, Stigmast-4-en-3-one & 4-Stigmasten-3-one. The six compounds were identified by comparing their retention time and peak area with that of literature and by interpretation of mass spectra. Hence, at any rate Nephrolepis cardifolia is an attractive material for further research leading to possible drug development.

Keywords: GC-MS analysis, nephrolepis cardifolia, phytoconstituents, tirumala hills

Received 30 Oct 2014 Received in revised form 24 Nov 2014 Accepted 26 Nov 2014

\*Address for correspondence:

P. Suvarnalatha Devi,

Department of Applied Microbiology, Sri Padmavathi Mahila Visvavidyalayam, Tirupathi-517502, Andhra Pradesh, India.

E-mail: drsuvarnapallipati@gmail.com

#### INTRODUCTION

Medicinal plants have been used by human being since ages in traditional medicine due to their therapeutic potential and the search on medicinal plants led to the discovery of novel drug candidates used against diverse diseases. According to the World Health Organisation (WHO) in 2008, more than 80% of the world's population relies on traditional medicine for their primary healthcare needs [1]. Higher plants as of bioactive compounds sources continue to play a dominant role in the maintenance of human health. Reports available on green plants represent a effective reservoir of chemotherapeutants, these are nonphytotoxic, more systematic and easily

biodegradable [2- 4]. Plants are a rich source of secondary metabolites with interesting biological activities. In general these secondary metabolites are an important source with a variety of structural arrangements and properties [5]. Nephrolepis cardifolia (L.) Presl. is commonly known as fishbone fern, ladder fern, sword fern. It has bright green fronds that are from 16-32 inches (40-80 cm) long and about 4 inches (10 cm) wide at their widest point. *N.cardifolia* is commonly cultivated as ornamental fern in Shilong and often the tubers are eaten to quench thirst [6]. Juice of root tubers is taken to treat fever, indigestion, headache, cough, cold and hematuria. Whole plant is used to

cure renal, liver and skin disorder [7]. It is a vigorous grower and colony former, producing small scaly tubers on their roots. In nature, they will grow in a wide variety of situations, growing in the soil or among rocks as well as growing as an epiphyte.

In recent years GC-MS studies have been increasingly applied for the analysis of medicinal plants as this technique has proved to be a reliable method for the analysis of non-polar components and volatile essential oil, fatty acids, lipids and alkaloids [8-9]. GC-MS is the best technique to identify the bioactive components of long chain hydrocarbons, alcohols, acids used in the analysis of the herbal medicines, there are more significant advantages for GC-MS [10]. Thus, GC-MS should be the most preferable tool for the analysis of the volatile chemical compounds in medicines. The medicinal herbal importance of the plant is due to the presence of some special compounds like alkaloids. flavonoids. phenols. tannins and saponins. These active principles usually remain concentrated in the storage organs of the plants viz., roots, leaves, tubers etc., considering all these facts, present investigation is designed to find out phytochemical screening and GC-MS of N.cardifolia which evokes various therapeutic effects. Now we are aimed to isolate the bioactive compounds of N.cardifolia, a rare plant of Tirumala hills.

#### MATERIALS AND METHODS

## Collection and Identification of Plants

The whole plant was collected from the Japalitheertham of Tirumala hills. Tirupati [11]. The plant was identified using a dictionary of the Pteridophytes of India and was authenticated by Dr. K. Madhava Chetty, Assistant Professor, Department of Botany, S.V. University, Tirupati voucher specimen No.2679. The plant was washed 2-3 times with tap water and distilled water to remove the soil and dirt particles then gabbled, pulverized, air dried and subjected to extraction with gradient soxhlet apparatus.

## Phytochemical analysis of different extracts

The Extracts were analyzed for the presence of phytoconstituents such as Alkaloids, Flavonoids, Saponins, Tannins and Phenols. Following standard procedures were used [12 - 14].

#### Mayer's test for Alkaloids

To the acidic solution, Mayer's reagent (Potassiuum mercuric iodide solution) was added. Appearence of cream coloured precipitate indicates the presence of alkaloids.



Plant of Nephrolepis cardifolia (L.) Presl.

#### Ferric chloride test for Tannins

Small quantity of extract was mixed with water and heated on water bath. The mixture was filtered and ferric chloride was added to the filtrate. A dark green solution indicates the presence of tannins.

#### Shinoda's test for Flavonoids

To one ml each of alcoholic extract, a small piece of magnesium ribbon or magnesium foil was added and 3-4 drops of conc.HCL was added,change in colour from red to pink shows the presence of flavonoids.

#### Froth test for Saponins

About 0.2 g of the extract was shaken with 5 ml of distilled water and then heated to boil. Frothing (appearance of creamy mass of small bubbles) shows the presence of saponins.

#### Ellagic test for Phenols

One ml of each of the various extracts dissolved in alcohol and treated with 2-3 ml of 5% neutral ferric chloride solution. Colour change indicates the presence of phenols.

## Preparation of sample for GC-MS analysis

30 gm of the powdered plant material was soaked in 95 % acetone for 12 hrs. The extract was then filtered through Whatman filter paper No.1 along with 2 gm sodium sulphate to remove the sediments and traces of water in the filtrate. Before filtering, the filter paper was made wet with 95 % ethanol along with sodium sulphate. The filtratre was then concentrated by bubbling nitrogen gas into the solution. The extract contained both polar and non-polar phyto-components in the plant material. 2  $\mu$ l of this solution was employed for GC-MS analysis [15].

### **GC-MS** analysis

GC-MS analysis was carried out on a GC clarus 500 Perkin Elmer system and gas chromatograph interfaced to a mass spectrometer (GC-MS) instrument employing the following conditions: Column Elite-5MS fused capillary column (30 x 0.25mm x 0.25m df) composed of 5% Diphenyl / 95% Dimethyl poly siloxane. Helium (99.999%) was used as carrier gas at a constant flow of 1ml per min and injection volume of 2µl was adopted (split ratio of 10:1) The injected sample was detected by Turbo mass gold detector (Perkin Elmer) with the aid of Turbomass 5.2 software. During 36 minute GC extraction process, the oven temperature was programmed from of 110° C with an increase of 10°C/min up to 200° C, then 5°C/min up to 280° C (9 minutes hold). The injector temperature was set at 250°C (mass analyzer). Other parameters involved in the operation of Clarus 500MS, was also standardized (Inlet line temperature: 200°C; Source temperature: 200° C). Mass spectra were taken at 70eV and fragments from 45-450 Da. The MS detection was completed in 36 minutes. The detection employed the NIST (National Institute of Standards and Technology).

### **Compounds identification**

Identification was based on the molecular structure, molecular moss and calculated fragments.

Interpretation on mass spectrum GC-MS was conducted using the database of Standardand National Institute Technology [NIST] having more than 62,000 patterns. The name, molecular weight and structure of the components of the test materials were ascertained. The relative percentage amount of each component was calculated bv comparing its average peak area to the total areas. The spectrum of the unknown component was compared with the spectrum of the component stored in the NIST library.

### **RESULT AND DISCUSSION**

In the present study, the phytochemical screening of Nephrolepis cardifolia qualitatively shown the presence of Alkaloids, Flavonoids, Tannins, Saponins along with Phenols in all the extracts investigated. Pink colour was observed for Flavonoids (Shinoda's test), cream coloured precipitate for Alkaloids, blue colour was observed for tannins and saponins (Table 1). Some of the flavonoids that favour polar solutes entry bind to the bacteria's structural membrane proteins called changes porins, causing in the tridimensional confirmation exposuring the hydrophilic character of the pore, which lead to an easier passage of other polar bioactive compounds via diffusion [16]. Saponins have the property of precipitating and coagulating human RBC [17]. Flavonoids on the other hand are water soluble antioxidants and free radical scavengers, which are capable of preventing oxidative cell damage and have strong anticancer activity [18,19]. Tannins have astringent property, hasten healing of wounds and inflamed mucous membrane.

<b>Table 1: Results of Preliminar</b>	v Dhytochomical Evaluation	of Nonbrolonic cardifolia
Table 1: Results of Preniminar	y Phytochemical Evaluation	ol Nephrolepis caraijolia

Test for Secondary Metabolites	Alkaloids Mayers Test	Flavonoids Shinodas test	Tannins Fecl3 test	Phenols Ellagic test	Saponins Froth test
Acetone	++	+++	+	++	_
Benzene	+++	++	+++	+++	++
Ethylacetate,	+	+	_	++	+
Methanol	_	_	++	++	+++
Ethanol	_	+	++	_	+++

+++ - High Concentration, ++ - Moderate Concentration, + - Low Concentration-Negative

GC-MS is one of the best techniques to identify the constituents of volatile matter, long chain, branched chain hydrocarbons, alcohols, acids, esters etc. The GC-MS identification of the chemical constituents was based on comparison of their mass spectra with NIST and WILEY libraries. Structures were defined by percentage similarity values. They are confirmed by the study of base peaks, retention time (RT), and molecular weight (MW) of the compounds. By the study of GC-MS we identify six major chemical constituents. They are identified as Neo phytadiene and 2, 6, 10-trimethyl, 14-Ethylene, 2 hexadecen-1-ol and 3,7,11,15- tetra methyl-R-R, hexadecanoic acid and palmitic acid, 9-octadecenoic acid(Z) and oleic acid, Octadecanoic acid and

stearic acid. Stigmast-4-en-3-one (Table 2) (Figure 1 & 2). The first compound identified with less retention (18.152 min) was Neophytadiene and 2,6,10 trimethyl 14-ethylene whereas Stigmast-4-en-3-one was the last compound which took longest retention time (26.400 min) to identify. N-Hexadecanoic acid – Palmitic acid (RT) can be an antioxidant, hvpocholesterolemic, nematicide, pesticide, lubricant activities and hemolytic 5alpha is a reductase inhibitors. 9,12-Octadecanoic acid and squalene were identified in the ethanol leaf extract of Aloe vera [20] and Vitex negundo [21]. Our results are in agreement with reports that many plant extracts have been as a source of medicinal plant to cure number of diseases [22].



Figure 1: GC-MS Chromatogram of Nephrolepis cardifolia, acetone extract

S.NO.	<b>Retention time</b>	Name of the compound	Base	Molecular Ion
			peak	Peak
1	18.152	Neo phytadiene and 2,6,10- trimethyl,14-Ethylene (1)	68	278
2	18.594	2 hexadecen-1-ol and 3,7,11,15- tetramethyl-R-R (2)	68	278
3	19.376	hexadecanoic acid and palmitic acid (3)	73	256
4	21.065	9-octadecenoic acid(Z) and oleic acid (4)	55	264
5	21.255	Octadecanoic acid and stearic acid (5)	43	284
6	26.400	Stigmast-4-en-3-one and 4- Stigmasten -3-one (6)	124	412

Table 2: Name of the Chemical compounds identified by GC-MS Analysis



Figure 2: Acetone extract of *Nephrolepis cardifolia* were Identified by GC-MS analysis

#### CONCLUSION

It was concluded that acetone extract of the total plant of Nephrolepis cardifolia possesses various potent bioactive compounds and is recommended as a plant of phyto-pharmaceutical Further subjects importance. are required to explore the potential responsible for compounds the biological activity from Nephrolepis cardifolia for application in drug delivery, nutritional or pharmaceutical studies.

#### ACKNOWLEDGEMENT

The authors are thankful to the University Grants Commission, New Delhi, for financial assistance under UGC Major Research Project and Department of Applied Microbiology, DST-FIST&CURIE, Sri Padmavati Mahila Visvavidyalayam (Women's University), Tirupati, Andhra Pradesh, India for providing Research facilities. The authors are also thankful to authors/ editors/ publishers for all those articles, journals and books from where the literature for this article has been reviewed and discussed.

#### REFERENCES

- Pierangeli G, Vital G, Rivera W. Antimicrobial activity and cytotoxicity of Chromalaena odorata [L.f] king and Robinson and Uncaria perrottetii [A.Rich] Merr Extracts; Journal of Medicinal Plants Research 2009, 3(7), 511-518.
- 2. Vyas GD. Soil fertility Deterioration in crop land due to pesticide. Journal of Indian botanical society 1999, 78,177-178.
- 3. Kaushik JC, Arya Sanjay, Tripathi NN, Arya S. Antifungal properties of some plant extracts against the damping off fungi of forest nurseries. Indian Journal of Forestry 2002,25,359-361.
- Chaman lal,Verma LR. Use of certain bioproducts for insect-pest control. Indian Journal of Traditional knowledge 2006, 5 (1), 79-82.
- de-Fatima A, Modolo LV, Conegero LS, Pili RA, Ferreira CV, Kohn LK. Lactones and their derivatives, biological activities, mechanisms of action and potential leads for drug design. Current Medicinal Chemistry 2006,13,3371-3384.

- 6. Chhetri RB. Trend in Ethnodomestication of some wild plants in Meghalaya, North East India. Indian Journal of Traditional Knowledge 2006,53 (3), 342- 347.
- Dhiman, Anil Kumar. Ethnomedicinal uses of some pteridophytic species in India. Indian Fern journal 1998, 15 (1-2), 61-64.
- 8. Jie MSF, Choi CYC. Journal of the International Federation of Clinical Chemistry 1991, 3, 122.
- Bertz JM, Gay ML, Mossoba MM, Adams S. Journal of AOAC International 1997, 80, 303.
- 10.Sridharan S, Meena V, Kavitha V, Agnel Arul John Nayagam. GC-MS study and phytochemical profiling of mimosa pudica Linn. Journal of Pharmacy Research 2011,4(3),741-742.
- 11.Swamy PM, Thammanna. Ferns and Fern allies of Tirumala hills, 1985.
- 12.Trease GE, Evans WC. Pharmacognosy Brailliar tiridal can Macmillian publishers, 1989.
- 13.Sofowora A. medicinal plants and Traditional medicine in west Africa. john wiley and sons, New York,1982.
- 14.Harbone JB. Phytochemical methods, Chapman and hall ltd.London, 3,1973,135-203.
- 15.Merlin NJ, Parthasarathy V, Manavala R, Kumravel SS. chemical investigation of aerial parts of Gmelina asiatica Linn by GC-MS, Pharmacognosy Research 2009,1(3),152-156.
- 16.Van-Burden TP, Robinson WC, Formation of complexes between protein and tannic acid, Journal of Agriculture and Food Chemistry 1981,1,77-82.
- 17.Okwu DE, Josiah C. Evaluation of the chemical composition of two Nigerian medicinal plants, African Journal of Biotechnology 2006, 5(4),357-361.
- 18.Salah N, Miller NJ, Pagange G, Tijburg L, Bolwell GP, Evans C. Polyphenolic flavonoils as seavenger of aqueous phase radicals as chain breaking antioxidant, Archives of Biochemistry and Biophysics 1995,2, 339-346.
- 19.Del-Rio A, Obdululio BG, Casfillo J, Marin FG,Ortuno A. Uses and Properties of citrus flavonoids, Journal of Agriculture and Food Chemistry 1997,45,4505-4515.
- 20.Arunkumar S, Muthuselvam M. 2009.Analysis of phytochemical constituents and antimicrobial activities of Aloe vera L. against clinical pathogens.

World Journal of Agricultural Sciences 2009, 5(5), 572-576.

- 21.Praveen kumar P, Kumaravel S, Lalitha C. Screening of antioxidant activity ,total phenolics and GC-MS study of vitex negundo. African Journal of Biochemistry Research 2010,4(7),191-195.
- 22.0Kwu DE, Phytochemicals and vitamin content of indigenous spices of Southeastern Nigeria. Journal of Sustain Agriculture and Environment 2004,6(1), 30-37.