Research & Reviews: Journal of Pharmaceutics and Nanotechnology

Nanostructured Lipid Carriers as a Drug Carrier Spandana Peddinti*

Department of Pharmaceutics, St. Anns college of Pharmacy, Vizianagaram, Andhra Pradesh, India

Review Article

Received: 04/07/2016 Accepted: 12/08/2016 Published: 31/08/2016

*For Correspondence

Spandana Peddinti, B pharmacy, St. Anns college of Pharmacy, Vizianagaram, Andhra Pradesh, E-mail:

spandanapeddinti@gmail.com

E-mail: spandanapeddinti@gmail.com

Keywords:	Drug;	Lipid
nanoparticles;	Lipid	carriers
(NLCs); Drug release		

Nanostructured lipid carriers (NLCs) are drug-delivery systems and they are made out of solid and fluid lipids as a core matrix. It was demonstrated that NLCs uncover a few points of interest for medication treatment conventional carriers, including increased solubility, the ability to enhance storage stability, improved permeability and bioavailability, prolonged half-life, reduced adverse effect, and tissue-targeted delivery. NLCs have pulled in expanding consideration as of late. This survey depicts developments in drug delivery utilizing NLCs procedures. The structures, preparation methods, and physicochemical characterization of NLCs are deliberately clarified in this review. The capability of NLCs to be utilized for different administration routes is highlighted. Uncommon consideration is paid to parenteral injection and topical delivery since these are the most widely routes for exploring NLCs. Pertinent issues for the introduction of NLCs with business sector, including pharmaceutical and cosmetic applications, are talked about. The related licenses of NLCs for drug delivery are additionally checked on. At last, the future advancement and current impediments waiting be resolved are elucidated.

ABSTRACT

INTRODUCTION

In the very recent years, it has ended up apparent that the advancement of novel medications is deficient for ensuring progress in medication treatment. Exciting experimental information got in vitro is regularly trailed by disappointing results in the in vivo or clinical circumstance. Overwhelming purposes behind this disappointment are the deficient drug concentration in the body as a result of quick metabolism system, the high medication poisonous quality as a result of extensive distribution, poor drug dissolvability in formulations, and high change or inter subject variability of plasma medication or drug levels ^[1-11]. A promising way to deal with beating this issue is the improvement of feasible drug-delivery systems. In the past decades, a few methodologies have been studied to develop or create nanosized drug- carrier systems ^[12-18]. These drug delivery systems are essentially divided into two groups: polymeric nanoparticles and lipid nanoparticles ^[19-25]. Polymeric nanosystems are strong colloidal particles comprising of non-biodegradable synthetic polymers or biodegradable macromolecular materials from synthetic, semisynthetic or normal assets. The disadvantages of polymeric nanoparticles are the cytotoxicity of polymers and the absence of reasonable large scale production techniques ^[26,27]. Attributable to the normal and organic beginnings of the materials, the toxicological danger connected with lipid nanoparticles is significantly less than the risk connected with polymeric nanoparticles.

Lipid nanoparticles made up of solid matrix (Solid Lipid Nanoparticles-SLNs) are arisen from oil-in-water nanoemulsions formed by changing liquid oil with a solid lipid. The original of SLNs was produced at the beginning of 1990 ^[28,29]. The benefits of SLNs are the utilization of physiological lipids, the evasion of organic solvents, and this can be applicable to large scale production ^[30,31]. As drug delivery systems, SLNs can enhance bioavailability, protect sensitive drugs from a rigorous environment, and control drug-release attributes ^[32-35]. By and by, SLNs shows few drawbacks as drug carriers including an eccentric gelation inclination, polymorphic move, and low incorporation because of the crystalline structure of solid lipids. At the turn of the thousand years, nanostructured lipid carriers (NLCs) were designed to resolve, at times, the issues raised by SLNs. NLCs are designed by controlling the blending of solid lipids with liquid oil, prompting uncommon nanostructures in the matrix. The potential disadvantages of SLNs, for example, restricted drug loading and drug expulsion during storage, can be avoided from by the new era. In this review we might want to demonstrate the present development of NLCs for drug

delivery and the focusing on application. The different sorts of NLCs for drugs s and the possible delivery routes are likewise portrayed in the present survey.

STRUCTURES AND PREPARATION METHODS OF NLCs

Materials for NLCs

The fundamental ingredients for NLCs incorporate lipids, water, and emulsifiers. Both strong and fluid lipids are incorporated into NLCs for developing the inner cores. The solid lipids normally utilized for NLCs incorporate glyceryl behenate (Com-pritol® 888 ATO), glyceryl palmitostearate (Precirol® ATO5), unsaturated fats (e.g. stearic corrosive), triglycerides (e.g. tristearin), steroids (e.g. cholesterol), and waxes (e.g. cetyl palmitate). These lipids are in a strong state at room temperature. They would liquefy at higher temperatures (e.g. >80°C) amid the readiness procedure. Fluid oils ordinarily utilized for NLCs comprise of edible oils from regular sources. The medium chain triglycerides, for example, Miglyol® 812, are frequently used as the constituents of fluid lipids due to their comparative structures to Compritol® [35-38]. Other sleek segments, for example, paraffin oil, 2-octyl dodecanol, propylene glycol dicaprylocaprate (Labrafac®), isopropyl myristate and squalene are incorporated also. Then again, the unsaturated fats, for example, oleic corrosive, linoleic corrosive, and decanoic corrosive, are incorporated into NLCs for their worth as having sleek parts and as being infiltration enhancers of topical delivery. As a rule, these lipids are as of now affirmed by European and American administrative powers for clinical applications and for their "for the most part perceived as sheltered" (GRAS) status. There is a requirement for novel and biocompatible oils that are financially savvy, non-bothering, and fit for being sanitized before application. Vitamin E (α -tocopherol) and different tocols have been researched as materials for nanoemulsions [39-42]. Tocols can serve as a decision of oils for NLCs in light of their strength, simplicity of creation on an extensive scale, and great dissolvability in lipophilic drugs [43]. NLCs created utilizing characteristic oils from plants are likewise presently mainstream. Averina et al. [44] have utilized Siberian pine seed oil and fish oil from Baikal Lake as the fluid oils since they indicate satisfactory physical and substance solidness to NLCs.

There are three techniques transcendently used to for the preparation of NLCs; hot homogenization, cold homogenization, and microemulsion. Hot homogenization is performed at temperatures above the melting point of the lipids. At to start with, lipid phase and aqueous phase are arranged independently. The lipid phase comprises of solid and liquid lipids and additionally lipophilic emulsifiers, while the aqueous phase comprises of double-distilled water and hydrophilic emulsifiers. Both phases are warmed separately to a high temperature for a determined time. The aqueous phase is added to the lipid phase and blended. The blend can be homogenized by a high-shear homogenizer. At times, the blend can be further treated utilizing a water-bath or probe-type sonicator to get the littler and that's just the beginning customary size dissemination. The built up high-temperature high-weight homogenization system may cause degradation of heat-sensitive drugs. Accordingly an enhanced procedure is expected to minimize the chemical instability. A straightforward strategy is the decrease of the warming temperature. Hung et al. [18] have decreased the handling temperature from 85°C to 60°C. It is found that 32% of vitamin E is corrupted in NLCs arranged utilizing the routine method following a 90-day storage period. Then again, no degradation is identified in NLCs arranged in the lower-temperature condition. A comparative result is watched on account of α -carotene. In the cold homogenization method, the lipid melt is cooled and the solid lipid is ground to lipid microparticles. The microparticles are dispersed in a cold emulsifier solution for yielding a pre-suspension. Thusly, the suspension is homogenized at or below room temperature. The cavitation power is sufficiently high to break the microparticles directly to NLCs. This procedure can maintain a strategic distance from the liquefying of the lipids and accordingly minimize loss of hydrophilic drugs to the aqueous phase [45-51]. Notwithstanding, it ought to be advised that the molecule size may not accomplish the nanosized range because of the absence of hot treatment. A transparent and thermodynamically stable dispersion, supposed microemulsion, can be formed when the softened lipids, emulsifiers, and water are blended in a right proportion. The further expansion of the microemulsion to water prompts precipitation of the lipid phase forming fine particles [51-58]. Large scale production of lipid nanoparticles by the microemulsion strategy seems possible for the pharmaceutical industry. Since dilute nanoparticle dispersion is delivered, some of the time the products should be concentrated by ultrafiltration or lyophilization [59-66].

DRUG DELIVERY BY NLCs

The most prominent use of NLCs is that of a drug nanocarrier. NLCs have been designed to convey the drug by various application routes, including parenteral infusion, topical skin delivery, oral administration, ocular delivery, and pulmonary inhalation ^[66-74]. Among them, the routes of injection and the skin are the most examined pathways for NLCs. The accompanying is the introduction and description of the drug administration by NLCs categorized by different routes. The novel application of NLCs for gene delivery is also described ^[74-80].

CURRENT AND FUTURE DEVELOPMENTS

The selection of vehicles is vital for drug delivery to apply greatest activity and cause minimal antagonistic effects. Some novel nanocarriers are considered to load drugs for treatment ^[66-70]. Among them, NLCs have increased much enthusiasm in recent years in view of the fulfilled drug carrier potency and safety. This review states recent advances in drug delivery by NLCs ^[81-87]. Notwithstanding intravenous administration, topical and oral routes are conceivable pathways for drug delivery from NLCs. Downsides of clinically utilized vehicles can be resolved by utilizing lipid nanoparticles ^[87-95]. It is normal that the utility of NLCs in essential research and the clinical setting will be broader later on due to dire needs to find new therapies, for example, treatments for cancer, neurodegenerative disease, and inflammation. Numerous examinations have analyzed lipid nanoparticles are designed for less adverse effects, longer half-life, and higher bioavailability compared to conventional carriers. Be that as it may, just a couple NLCs have been utilized as a part of current clinical practice ^[95-98]. The cosmetic products are the most ordinarily utilized NLCs available in the market. Additionally, clinical trials exploring NLCs for drugs are constrained. It is proposed that more results in animal and clinical studies will empower future utilization of drug therapy using the lipid nanocarriers.

Although most of the ingredients used for composing NLCs are biodegradable, the conceivable lethality of nanoparticles still can't be ignored in the improvement of NLCs ^[98-100]. Nanomaterials are thought to have moreserious adverse effects on organisms than materials of a bigger size because of their small size and comparing higher surface areas. Data with respect to the health concerns of materials at the nano-level is still restricted. Intravenous infusion and topical delivery are the main routes for drug administration by NLCs. The effort to develop alternative routes and to treat different diseases with NLCs should be kept on broadening their applications. Permeation by means of the gastrointestinal tract and BBB might be a future pattern. Combination of two therapeutically active agents to be included in a single nanosystem is another thought for future improvement. Albeit a few advantages of NLCs for drug delivery are illustrated, the systems for upgraded viability are not completely caught on. Consequently, these systems ought to be further investigated with the goal of elucidation and efficacy enhancement.

REFERENCES

- 1. Heli H. Electrochemical studies of vitamin k3 and its interaction with human serum albumin using a carbon nanoparticles-modified electrode. J Nanomater Mol Nanotechnol. 2013;2:7.
- 2. Bajaj L and Sekhon BS. Nanocarriers based oral insulin delivery. J Nanomater Mol Nanotechnol. 2014;3:1.
- 3. Hwang TL, et al. Development and evaluation of perfluorocarbon nanobubbles for apomorphine delivery. J Pharm Sci. 2009;98:3735-47.
- 4. Menaa F, et al. Dietary intake of (-)-epigallocatechin-3-gallate against aging and cancers: nanoencapsulation of multi-rings still requires new rounds! J Nanomater Mol Nanotechnol. 2013;2:7.
- 5. Lokesh BVS and Kumar PV. Enhanced cytotoxic effect of chemically conjugated polymeric sirolimus against ht-29 colon cancer and a-549 lung cancer cell lines. J Pharm Drug Deliv Res. 2015;4:2.
- 6. Ajore R, et al. Effect of humidity on structural distortion and conductance of DNA nanowire. J Nanomater Mol Nanotechnol. 2013;2:7.
- 7. Habibzade S, et al. Effect of nano-zno on decay resistance and artificial weathering of wood polymer composite. J Nanomater Mol Nanotechnol. 2014;3:3.
- 8. Xavier S, et al. Effect of neodymium substitution on structural and magnetic properties of cobalt ferrite nanoparticles. J Nanomater Mol Nanotechnol. 2013;2:7.
- 9. Forny L, et al. Contact angle assessment of hydrophobic silica nanoparticles related to the mechanisms of dry water formation. Langmuir. 2010;26: 2333-8.
- 10. He S, et al. Towards methanol electro-oxidation: comparative study on imidazolium and guanidinium ionic liquids supported pt nanocrystals on carbon nanotubes. J Nanomater Mol Nanotechnol. 2013;2:7
- 11. Hu FQ, et al. Preparation and characteristics of monostearin nanostructured lipid carriers. Int J Pharm. 2006;314:83-9.
- 12. AL-Thabaiti SA, et al. Synthesis of poly(vinyl alcohol)-silver nano-composites and effect of ctab on their morphology. J Nanomater Mol Nanotechnol. 2013;2:7.
- 13. Liye X, et al. Fabrication of Cu hollow microspheres by liquid reduction method. J Nanomater Mol Nanotechnol. 2014;3:3.
- 14. Loomba L and Sekhon BS. Calcium phosphate nanoparticles and their biomedical potential. J Nanomater Mol Nanotechnol. 2015;4:1.

- 15. Wan G, et al. Hierarchical ZnO nanostructures derived from zn-al layered double hydroxides and their photocatalytic activity. J Nanomater Mol Nanotechnol. 2014;3:4.
- 16. Castelli F, et al. Characterization of indomethacin-loaded lipid nanoparticles by differential scanning calorimetry. Int J Pharm. 2005;304:231-8.
- 17. Kumar D, et al. Some mechanical properties of carbon nanotubes heterojunctions. J Nanomater Mol Nanotechnol. 2014;3:3
- 18. Hung LC, et al. An improved method for the preparations of nanostructured lipid carriers containing heat sensitive bioactives. Colloids Surf B: Biointerf. 2011;87:180-186.
- 19. Faheim AS, et al. Effect of Zn substitution on the characterization of cobalt ferrite nano particles prepared co-precipitation method. J Nanomater Mol Nanotechnol. 2014;4:1.
- 20. Sato K. Crystallization behaviour of fats and lipids a review. Chem Eng Sci. 2001;5:2255-2265.
- 21. Vaze OS. Pharmaceutical nanocarriers (liposomes and micelles) in cancer therapy. J Nanomed Nanotechnol. 2016;7:e138
- 22. Álvarez-Bautista A, et al. Poly(N–Isopropylacrylamide–Co–Acrylic Acid) smart nanocarriers for drug release: a study of theophylline delivery. J Mol Genet Med. 2015;9:196
- 23. Dall'Agnol FF and den Engelsen D. Field emission simulations of carbon nanotubes and graphene with an atomic model. J Nanomater Mol Nanotechnol. 2014;3:4.
- 24. Fadel M, et al. Antitumor efficiency of doxorubicin loaded in liposomes and poly ethylene glycol coated ferrofluid nanoparticles. J Nanomater Mol Nanotechnol. 2015;4:1
- 25. Kumar R and Lal S. Synthesis of organic nanoparticles and their applications in drug delivery and food nanotechnology: a review. J Nanomater Mol Nanotechnol. 2014;3:4.
- 26. Gajbhiye KR, et al. Targeted brain delivery of bioactive molecules using nanocarriers. J Bioequiv Availab. 2015;7:112-122
- 27. Olson JL, et al. Neuroprotective effect of photoactive quantum dots in progressive retinal photoreceptor degeneration. J Nanomater Mol Nanotechnol. 2013;2:4.
- 28. Li C, et al. Electronic Theory of Ultrafast Spin Dynamics in NiO. J Nanomater Mol Nanotechnol. 2014;3:4.
- 29. Ramdani L, et al. Multifunctional curcumin-nanocarriers based on host-guest interactions for alzheimer disease diagnostic. J Nanomed Nanotechnol. 2015;6:270.
- 30. Suresh Sagadevan. Preparation, structural and electrical properties of tin oxide nanoparticles. J Nanomater Mol Nanotechnol. 2015;4:1.
- 31. Misra R, et al. Design considerations for chemotherapeutic drug nanocarriers. Pharm Anal Acta. 2014;5:279.
- 32. Gambino D. Metal-Organic frameworks in nanotherapeutics: development of novel drug nanocarriers for conventional and nuclear oncology. J Nanomed Biotherapeut Discov. 2012;2:e120.
- 33. López T, et al. Occlusion of INTERFERON® and COPAXONE® on SBA-15 silica reservoirs for their use in the treatment of demyelization diseases. J Nanomater Mol Nanotechnol. 2013;2:4.
- 34. Patil J. Encapsulation technology: opportunity to develop novel drug delivery systems. J Pharmacovigil. 2016;4:e157
- 35. Koushik OS, et al. Nano drug delivery systems to overcome cancer drug resistance a review. J Nanomed Nanotechnol. 2016;7:378
- 36. Mikheev KG, et al. Laser bleaching of carbon nanotubes suspension in n,ndimethylformamide. J Nanomater Mol Nanotechnol. 2013;2:4.
- 37. Gopi S. Effective drug delivery system of biopolymers based on nanomaterials and hydrogels a review. Drug Des. 2016;5:129
- 38. Otles S and Yalcin B. Food chemistry and nanoscience. J Nanomater Mol Nanotechnol. 2013;2:4.
- 39. Nirmala MJ and Nagarajan R. Microemulsions as potent drug delivery systems. J Nanomed Nanotechnol. 2016;7:e139.
- 40. Mandal B. Personalized nanotheranotics for cancer. J Biotechnol Biomater. 2016;6:e127.
- 41. Zaman HH. Addressing solubility through nano based drug delivery systems. J Nanomed Nanotechnol. 2016;7:376.
- 42. Hassan AZA and Mahmoud AWM. Hydrothermal synthesis of nano crystals (a.m.) zeolite using variable temperature programs. J Nanomater Mol Nanotechnol. 2015;4:4.

- 43. Dudhipala N, et al. Amoxycillin trihydrate floating-bioadhesive drug delivery system for eradication of helicobacter pylori: preparation, in vitro and ex vivo evaluation. J Bioequiv Availab. 2016;8:118-124.
- 44. Averina ES, et al. Nanostructured lipid carriers (NLC) on the basis of Siberian pine (Pinus sibirica) seed oil. Pharmazie. 2010;65:25-31.
- 45. Mohsen R, et al. design, synthesis, characterization and toxicity studies of poly (n-iso-propylacrylamide-colucifer yellow) particles for drug delivery applications. J Nanomed Nanotechnol. 2016;7:363.
- 46. Brijesh KV, et al. Physicochemical characterization and in-vitro dissolution enhancement of bicalutamidehp-β-cd complex. J Pharm Drug Deliv Res. 2015;3:2.
- 47. Raza A, et al. In-situ synthesis, characterization and application of co0.5zn0.5fe2o4 nanoparticles assisted with green laser to kill s. enterica in water. J Nanomater Mol Nanotechnol. 2016;5:2.
- 48. Yari A, et al. Sensing element based on a new nanoparticle to develop a carbon past electrode for highly sensitive determination of ag+ in aqueous solutions. J Nanomater Mol Nanotechnol. 2015;4:4.
- 49. AbouAitah KEA, et al. Mesoporous silica materials in drug delivery system: ph/glutathione- responsive release of poorly water-soluble pro-drug quercetin from two and three-dimensional pore-structure nanoparticles. J Nanomed Nanotechnol. 2016;7:360.

50. Ferreira H, et al. Deformable liposomes for the transdermal delivery of piroxicam. J Pharm Drug Deliv Res. 2015;4:4.

- 51. Satya Krishna HP, et al. Solubility and dissolution enhancement of candesartan cilexetil by liquisolid compacts. J Pharm Drug Deliv Res. 2013;2:2.
- 52. Ramani T, et al. Synthesis, characterization of phosphine, phosphine oxide and amine stabilized platinum nanoparticles in organic medium. J Nanomater Mol Nanotechnol. 2015;4:4.
- 53. Van Tilburg CWJ. Spinal analgesic drug delivery for ehlers-danlos hypermobility type chronic pain treatment: a case report. J Pain Relief. 2016;5:235.
- 54. Ranjna CD, et al. Inhibiting human lactate dehydrogenase-c for male fertility control; initial hits. J Pharm Drug Deliv Res. 2014;3:2.
- 55. Colone M, Kaliappan S, et al. Redox-active microcapsules as drug delivery system in breast cancer cells and spheroids. J Mol Genet Med. 2016;10:200.
- 56. Solomon AO, et al. Making drugs safer: improving drug delivery and reducing side-effect of drugs on the human biochemical system. J Pharm Drug Deliv Res. 2015;4:4.
- 57. Shanmugan P and Bandameedi R. Chronotherapeutic drug delivery systems. J Drug Metab Toxicol. 2015;6:194
- 58. Orji JI, et al. Physicochemical properties of co-precipitate of plantain peel cellulose and gelatin. J Pharm Drug Deliv Res. 2015;4:4.
- 59. Wang X and Lu W. Active targeting liposomes: promising approach for tumor-targeted therapy. J Bioequiv Availab. 2016;8:013-014.
- 60. Isabel S and Paula G. Encapsulation of fluoroquinolones in 1-palmitoyl-2-myristoyl-phosphatidylcholine: cholesterol liposomes. J Pharm Drug Deliv Res. 2013;2:1.
- 61. Lopes CM and Soares C. Transdermal drug delivery systems activated by physical stimuli: techniques and applications. Drug Des. 2015;4:e129.
- 62. Teeranachaideekul V, et al. Encapsulation of ascorbyl palmitate in nanostructured lipid carriers (NLC)effects of formulation parameters on physicochemical stability. Int J Pharm. 2007;340:198-206.
- 63. Sivaramakrishnan R, et al. Glucocorticoid entrapment into lipid carriers-characterization by parelectric spectroscopy and influence on dermal uptake. J Control Release. 2004;97:493-502.
- 64. Scott D and Bae Y. Block copolymer crosslinked nanoassemblies co-entrapping hydrophobic drugs and lipophilic polymer additives. J Pharm Drug Deliv Res. 2013;2:2.
- 65. Zhang P, et al. Novel nanostructured lipid-dextran sulfate hybrid carriers overcome tumor multidrug resistance of mitoxantrone hydrochloride. Nanomed-Nanotechnol Biol Med. 2012;8:185-93.
- 66. Wissing SA, et al. Structural characterization of Q10-loaded solid lipid nanoparticles by NMR spectroscopy. Pharm Res. 2004;21:400-5.
- 67. Salehi M, et al. An alternative way to prepare biocompatible nanotags with increased reproducibility of results. J Nanomater Mol Nanotechnol. 2016;5:2.
- 68. Wu XC, et al. On further understanding of interaction of pristine carbon nanotubes with hemoglobin, serum and H2S. J Nanomater Mol Nanotechnol. 2013;2:2.

- 69. Gazák R, et al. Silybin and silymarin-new and emerging applications in medicine. Curr Med Chem. 2007;14:315-338.
- 70. Schubert MA, et al. Structural investigations on lipid nanoparticles containing high amounts of lecithin. Eur J Pharm Biopharm. 2006;27:226-236.
- 71. Mohamed Idrees RY and Khalid A. Comparative modeling of serotonin receptors 5ht2a and 5ht2c and insilico investigation of their potential as off-target to ethinylestradiol. J Pharm Drug Deliv Res. 2013;2:2.
- 72. Jia L, et al. Nanostructured lipid carriers for parenteral delivery of silybin: biodistribution and pharmacokinetic studies. Colloids Surf B: Biointerf. 2010;80:213-218.
- 73. Vashist SK. Nanomaterials-based health care and bioanalytical applications: trend and prospects. J Nanomater Mol Nanotechnol. 2013;2:2.
- 74. Saupe A, et al. Structural investigations on nanoemulsions, solid lipid nanoparticles and nanostructured lipid carriers by cryo-field emission scanning electron microscopy and Raman spectroscopy. Int J Pharm. 2006;314:56-62.
- 75. Olson JL, et al. Intraocular biocompatibility of gold-nanoparticles. J Nanomater Mol Nanotechnol. 2013;2:2.
- 76. Pan SY, et al. Bifendate treatment attenuates hepatic steatosis in cholesterol/bile salt- and high-fat dietinduced hypercholesterolemia in mice. Eur J Pharmacol. 2006;552:170-175.
- 77. Wang H, et al. Hydrothermal growth of aligned zno nanorods along the seeds prepared by magnetron sputtering and its applications in quantum dots sensitized photovoltaic cells. J Nanomater Mol Nanotechnol. 2013;2:2
- 78. Lombardi Borgia S, et al. Lipid nanoparticles for skin penetration enhancement-correlation to drug localization within the particle matrix as determined by fluorescence and parelectric spectroscopy. J Control Release. 2005;110:151-163.
- 79. Hu W, et al. Fibroblast behavior on PMMAEA and PMMAEA-collagen films and nanofibers. J Nanomater Mol Nanotechnol. 2013;2:5.
- 80. Feng F, et al. Preparation, characterization, and biodistribution of nanostructured lipid carriers for parenteral delivery of bifendate. J Microencapsul. 2011; 28:280-285.
- 81. Wiley TS, et al. H1R antagonists for brain inflammation and anxiety: targeted treatment for autism spectrum disorders. J Pharm Drug Deliv Res. 2015;4:3.
- 82. Joshi M and Patravale V. Nanostructured lipid carrier (NLC) based gel of celecoxib. Int J Pharm. 2008;346:124-132.
- 83. Nair AK, et al. Development and comparative assessment of hydrocolloid based against wax based gastro retentive bilayered floating tablet designs of atorvastatin calcium using qbd approach. J Pharm Drug Deliv Res. 2015;4:3.
- 84. Azarnova TO, et al. Effects of the nanostructured complex of biologically active compounds on the freeradical processes and the liver state of the chicken cross "Shaver 2000". J Nanomater Mol Nanotechnol. 2013;2:5.
- 85. Escher M, et al. Pharmacokinetic and pharmacodynamics properties of buprenorphine after a single intravenous administration in healthy volunteers: a randomized, double-blind, placebo-controlled, crossover study. Clin Ther. 2007;29:1620-1631.
- 86. Joshi RR and Devarajan PV. Anionic self-micro-emulsifying drug delivery system (smedds) of docetaxel for circulation longevity. J Pharm Drug Deliv Res. 2015;4:3.
- 87. Mahipalreddy D, et al. Preparation and evaluation of ketoprofen enteric coated mini tablets for prevention of chronic inflammatory disease. J Pharm Drug Deliv Res. 2015;4:2.
- 88. Ghashghaei S and Emtiazi G. Production of hydroxyapatite nanoparticles using tricalcium-phosphate by alkanindiges illinoisensis. J Nanomater Mol Nanotechnol. 2013;2:5.
- 89. Yuan H, et al. Preparation and characteristics of nanostructured lipid carriers for control-releasing progesterone by melt-emulsification. Colloids Surf B: Biointerf. 2007;60:174-179.
- 90. Dey B, et al. Comparative evaluation of hypoglycemic potentials of eucalyptus spp. leaf extracts and their encapsulations for controlled delivery. J Pharm Drug Deliv Res. 2013;3:2.
- 91. Wang JJ, et al. Lipid nanoparticles with different oil/fatty ester ratios as carriers of buprenorphine and its prodrugs for injection. Eur J Pharm Sci. 2009;38:138-146.
- 92. Efentakis M and Siamidi A. Design and evaluation of a multi-layer tablet system based on dextran. J Pharm Drug Deliv Res. 2014;3:2.

- 93. Ma Q, et al. Preparation of CdSe quantum dot sensitized solar cells based on improved successive ionic layer absorption and reaction method. J Nanomater Mol Nanotechnol. 2013;2:7.
- 94. Marcato PD and Durán N. New aspects of nanoparmaceutical delivery systems. J Nanosci Nanotechnol. 2008;8:2216-2229.
- 95. Humayoon R, et al. Quality control testing and equivalence of doxycycline hyclate (100 mg) capsule brands under biowaiver conditions. J Pharm Drug Deliv Res. 2014;3:2.
- 96. Ning Y, et al. A novel biosensor for detection of salmonella typhimurium carrying ssec gene based on the secondary quenching effect of carbon nanotubes. J Nanomater Mol Nanotechnol. 2013;2:5.
- 97. Xu X, et al. Anti-inflammatory activity of injectable dexamethasone acetate-loaded nanostructured lipid carriers. Drug Deliv. 2011;18:485-492.
- 98. Nabid MR, et al. Synthesis of nonionic dendrimer-like star block copolymers based on PCL and PEG as stabilizer for gold nanoparticles. J Nanomater Mol Nanotechnol. 2013;2:7.
- 99. Chattopadhyay N, et al. Solid lipid nanoparticles enhance the delivery of the HIV protease inhibitor, atazanavir, by a human brain endothelial cell line. Pharm Res. 2008;25:2262-2271.
- 100. Ma L, et al. Silver sulfide nanoparticles as photothermal transducing agents for cancer treatment. J Nanomater Mol Nanotechnol. 2016;5:2.