Research Article

Phytochemical Evaluation and Antioxidant Activity of *Glycosmis mauritiana* (lam.) Tanaka Leaf Extract

T. Ebenezer¹, *D. Kadamban¹, M.S. Shekhawat¹, S. Justin Packia Jacob²

- 1. Department of Botany, K. M. Centre for P.G. Studies, Pondicherry, India.
- 2. St. Joseph's College of Engineering, Chennai-600119, Tamil Nadu, India.

ABSTRACT

Medicinal plants can provide biologically active molecules and lead structures for the development of modified derivatives with enhanced activity and or reduced toxicity. Ensuring the safety, efficacy and quality of traditional medicine is a priority in as much as it must be respected, preserved and promoted as a branch of medicine. In plants, the medicinal value of the secondary metabolites is due to the presence of chemical substances that produce a definite physiological action on the human body. Research into the isolated plant constituents is of great importance for the development of bioactive substances from ethnic medicine. With the emergence of latest technologies and enhanced knowledge about the isolated plant constituents, characterization and analytical tools, lot of compounds are efficiently isolated from potential plants and have been of great contribution to the drug discovery from ethnic plants. The present investigation is aimed at investigating the phytochemical evaluation, quantification and antioxidant studies of a medicinal plant *Glycosmis mauritiana*. The phytochemical quantification was done for phenolics, tannins and flavanoids. The antioxidant screening was done by FRAP assay and DPPH free radical scavenging assay, using five different solvent extracts. Significant antioxidant activity of ethanolic extracts was recorded in *Glycosmis mauritiana* which can be used as a potent source of natural antioxidant.

Keywords: Antioxidants, DPPH, FRAP assay, *glycosmis mauritiana*, secondary metabolites.

Received 05 August 2013 Received in revised form 23 August 2013 Accepted 25 August 2013

*Address for correspondence:

D. Kadamban Department of Botany, K. M. Centre for P.G. Studies, Pondicherry India. E-mail: kadamband@gmail.com

INTRODUCTION

About 80% of the world population relies on Traditional Medicines for their primary health care needs [1-3]. Attempts to document the plant wealth of India have continued since the landmark publication by [4]. Around 1700 species have been documented for their biological properties and drug action [5] and data is available for approximately 1200 species. Recently, interest has increased considerably in finding naturally occurring antioxidants for use in foods because of their potential in health promotion and disease prevention, and their high safety and consumer acceptability [6]. In Tamil Nadu, crushed leaves and barks of Glycosmis mauritiana (Lam.) Tanaka were externally applied on forehead to cure severe headache by the Irula tribes of the Nilgiris [7]; leaf paste was

externally used for eczema and skin diseases by the Thottianaickan tribes of Semmalai hills [8] and Valaiyans tribes of Piranmalai hills [9]. Keeping the significances of Traditional Medicine in plant-based drug development *Glycosmis mauritiana* of family Rutaceae was selected for the present study.

MATERIALS AND METHODS

1. Collection of plant materials

The plant materials (leaves) of *Glycosmis mauritiana* (**Fig. 1**) were collected from the Madras Christian College Campus, Tambaram, Tamil Nadu and its botanical identity was confirmed at French Institute Herbarium, Pondicherry. The herbarium specimens were deposited in the Department of Botany, Kanchi Mamunivar Centre for Post Graduate Studies, Lawspet, Pondicherry for further reference (Voucher no. DK & TE195).

2. Solvent extraction

The air dried powdered plant material (leaf) was extracted successively in soxhlet extractor with petroleum ether, chloroform,

ethyl acetate ethanol and water. All the solvent extracts were evaporated to remove the final traces of the respective solvents. The extract recovery in different solvents was expressed as per cent of the plant sample dry matter.



Figure 1: Morphology of Glycosmis mauritiana

3. Preliminary phytochemical screening

Phytochemical screening of different solvent extracts was carried out following the methods of [10,11] for the presence of various classes of compounds such as alkaloids, flavanoids, tanins, phenolic compounds, sterols, saponins, resins, glycosides, carbohydrates, amino acids and proteins.

4. Quantitative estimation of phytochemicals

(i) Determination of total phenols, tannins and flavonoid

The total phenolic and tannin content were determined by following the method of [12]. Flavonoid content was determined by [13] method.

5. Pharmacological Studies

(i) *In vitro* antioxidant activity (Ferric Reducing Antioxidant Power (FRAP) assay and free radical scavenging activity on DPPH)

The FRAP activity was measured by using the [14] method and free radical scavenging activity was measured according to the method described by [15].

RESULTS

1. Preliminary phytochemical screening

Preliminary phytochemical screening of leaves revealed the presence of alkaloids, carbohydrates, flavonoids, phenols and tannins in all the tested extracts. Glycosides were present in all the extracts except water. Saponins were present in petroleum ether extract, steroids in ethylacetate and methanol extracts and proteins in ethylacetate and water extracts (**Table 1**).

| Table 1: Preliminary phytochemical screening of | of various leaves extracts of <i>G. mauritiana</i> . |
|---|--|
| | Solvents |

| | borvents | | | | | |
|----------------|-----------------|------------|---------------|----------|-------|--|
| Phytochemicals | Petroleum ether | Chloroform | Ethyl acetate | Methanol | Water | |
| Alkaloids | + | + | + | + | + | |
| Flavonoids | + | + | + | + | + | |
| Saponins | + | + | - | - | - | |
| Phenols | + | + | + | + | + | |
| Steroids | - | - | + | + | - | |
| Tannins | + | + | + | + | + | |
| Carbohydrates | + | + | + | + | + | |
| Protein | + | + | - | + | - | |
| Glycosides | + | + | + | + | - | |

Note: + Present, - Absent

2. Quantitative estimation of phytochemicals

(i) Total flavonoids

Higher amount of total flavonoid content was recorded in leaves, as 13.41 ± 0.34 mg

RE/g in petroleum ether followed by 12.99 \pm 0.39 mg RE/g in ethylacetate, 12.13 \pm 0.20 mg RE/g in methanol, 10.11 \pm 0.44 mg RE/g in water and 8.44 \pm 0.76 in chloroform (**Fig. 2**).



(ii) Total phenols

In leaves, a high content of total phenolics was observed, as $226.03 \pm 0.50 \text{ mg TAE/g in}$ water followed by $122.04 \pm 0.93 \text{ mg TAE/g}$

in methanol, 87. 96 \pm 0.54 mg TAE/g in ethylacetate, 47.64 \pm 0.53 mg TAE/g in chloroform and 27.50 \pm 0.83 mg TAE/g in petroleum ether (**Fig.3**).



ig. 3. Total phenolic content of different solvent extract of *G. mauritiana* leaves

(iii) Total tannins

High tannin content was also observed in leaves, as 38.76 ± 0.44 mg TAE/g in water followed by 34.10 ± 0.83 mg TAE/g in

methanol, 27.01 ± 0.41 mg TAE/g in ethylacetate, 17.88 ± 0.41 mg TAE/g in chloroform and 14.00 ± 0.63 mg TAE/g in petroleum ether (**Fig. 4**).



3. *In vitro* antioxidant activity (i) FRAP Assay

In the FRAP assay, all the extracts showed concentration-dependent activity. In leaves, value maximum was observed in chloroform extract as 60.69 ± 0.89 followed by methanol extract (41.97 ± 0.27) and water extract (33.17 0.38) while ± moderate value was observed in ethyl acetate extract (28.90 ± 0.47) and minimum value was observed in petroleum ether extract (12.19 ± 0.07) as shown in the (Fig.5).





(ii) DPPH scavenging activity In DPPH scavenging activity, all the extracts showed concentration-dependent activity (Fig.6). In petroleum ether extract, the percentage of scavenging activity ranged from 10.63 ± 0.19 to 50.47 ± 0.24. Minimum and maximum activity was observed as 10.63 ± 0.19 at 40 µg and 50.47 ± 0.24 at 200 µg respectively, while moderate percentage activity was observed as 21.69 ± 0.39 at 80 µg, 31.47 ± 0.20 at 120 µg, 39.80 \pm 0.29 at 160 µg concentrations.



Fig. 6. DPPH scavenging activity of different solvent extracts of *G. mauritiana* leaves

In chloroform extract, the percentage of scavenging activity ranged from 19.54 ± 0.26 to 82.67 ± 0.15. Minimum and maximum activity was observed as 19.54 ± 0.26 at 40 ug and 82.67 ± 0.15 at 200 ug respectively, while moderate percentage activity was observed as 38.68 ± 0.36 at 80 μ g, 55.93 ± 0.20 at 120 μ g, 70.45 ± 0.33 at 160 µg concentrations. The percentage of scavenging activity in ethylacetate extract was ranged from 11.91 ± 0.84 to 62.48 ± 0.27. Minimum and maximum activity was observed as 11.91 ± 0.84 at 40 µg and 62.48 ± 0.27 at 200 µg respectively, while moderate percentage activity was observed as 25.21 ± 0.31 at 80 µg, 32.29 ± 0.31 at 120 μ g, 48.42 ± 0.26 at 160 μ g concentrations.

The percentage of scavenging activity in methanol extract was ranged from 8.59 ± 0.29 to 36.70 ± 0.52. Minimum and maximum activity was observed as 8.59 ± 0.29 at 40 μ g and 6.70 \pm 0.52 at 200 μ g respectively, while moderate percentage activity was observed as 15.73 ± 0.78 at 80 μ g, 23.83 ± 0.26 at 120 μ g, 40.64 ± 0.27 at 160 µg concentrations. In water extract, the percentage of scavenging activity ranged from 4.26 ± 0.29 to 36.70 ± 0.52. Minimum and maximum activity was observed as 4.26 \pm 0.29 at 40 µg and 36.70 \pm 0.52 at 200 µg respectively, while moderate percentage activity was observed as 10.21 ± 0.49 at 80 μ g, 17.21 ± 0.49 at 120 μ g, 27.14 ± 0.34 at 160 μg concentrations.

DISCUSSION

Medicinal plants have invariably been a rich source of modern drug discoveries. Many drugs in use today were either obtained from plants or developed using their chemical structure as templates [16, 17]. They play an important role in health care dynamics throughout the world for health security. Plants have traditionally been used as a source of medicine in India by indigenous people of different ethnic groups inhabiting various terrains for the control of various ailments afflicting human and their domestic animals [18]. The medicinal values of the plants are due to the bioactive constituents that produce definite physiological action on the human body [19]. The medicinal properties of plants could be based on the antioxidant, antimicrobial and antipyretic effects of the phytochemicals in them [20, 21]. In this context, an important medicinal plant *Glycosmis mauritiana* was chosen to investigate phytochemicals and antioxidant activity.

In plants, the medicinal value of the secondary metabolites is due to the presence of chemical substances that produce a definite physiological action on the human body [22-27]. Preliminary phytochemical screening of leaves of G. *mauritiana* revealed the presence of various phytoconstituents classes of such as alkaloids, carbohydrates, flavonoids,

glycosides, phenols, proteins, saponins, steroids and tannins in leaves (Table 1). Quantification of phytochemicals revealed the copious presence of alkaloids. flavonoids and total phenolic contents. However, the total content of phenols, flavonoids and tannins were found to be very much higher than the non-polar constituents like steroids. These reports indicate that total phenolics, flavonoid and responsible tannin content are for antioxidant activity of the plant as reported earlier by [28] and anti-hypertension activity by [29-30] Reported alkaloids, flavonoids and terpenoids from leaves and reported fruits and [31] alkaloids. carbohydrates, flavonoids, terpenoids and tannins from roots of allied species, *Glycosmis pentaphylla*.

Antioxidants are known to eliminate oxidative stress by scavenging free radicals and protect biological macromolecules from their toxic effect [32]. Therefore, in recent years evaluating plant antioxidant activity and their free radical quenching ability is considered to be an important task in pharmacological studies.

The FRAP assay measures the reducing potential of an antioxidant reacting with a ferric tripyridyltriazine (Fe³⁺-TPTZ) complex producing a colored ferrous tripyridyltriazine (Fe²⁺-TPTZ) [33]. Generally, the reducing properties are associated with the presence of compounds which exert their action by breaking the free radical chain by donating a hydrogen atom [34]. Frap assay treats the antioxidants in the sample as a reductant in a redox-linked colorimetric reaction [35]. In the present study, the trend for ferric ion reducing activities of G. mauritiana leaf extract showed the increased absorbance, due to the formation of the Fe²⁺-TPTZ complex with increasing concentration. In a related report, the water and ethanol extracts of sumac (Rhus coriaria L.) showed increased ferric reducing power with the increased concentration as standard antioxidants [36]. Hence they should be able to donate electrons to free radicals stable in the actual biological and food system.

The reactivity of ethanol extract of *G. mauritiana* of leaves was analyzed with the

stable free radical, DPPH that picks up one electron in the presence of a free radical scavenger, the absorption decreases and the resulting discoloration is stoichiometrically related to the number of electrons gained [37]. In the present study, the ethanol extract exhibited concentration dependent scavenging activity comparable with BHT. Similar observation was also reported in various *Phyllanthus* species such as *P. amarus, P. maderaspatensis, P. urinaria* and *P. virgatus* [38], *Costus pictus* [39], *Maranta arundanacea* [40].

The ethanolic extracts of *G. mauritana* were found to be an effective scavenger of DPPH and also possessed a good reducing power and FRAP activity. Earlier reports on the antioxidant activity of *G. mauritana* are very scanty. Therefore, it is very interesting to work on antioxidant property in the present study. The high antioxidant activity of G. *mauritana* leaf enhanced the potential interest for improving the efficacy of different products as neutraceutical and pharmacological agents. The consumption of this plant may play a role in preventing human diseases in which free radicals are involved such as cancer, cardiovascular disease and aging. However, further investigations on phenols, flavonoids and their in vivo antioxidant activity, and the different antioxidant mechanism are warranted for drug development.

CONCLUSION

Medicinal plants have invariably been a rich source of modern drug discoveries. Medicinal plants can provide biologically active molecules and lead structures for the development of modified derivatives with enhanced activity and or reduced toxicity. In this context *G. mauritiana* was chosen to investigate phytochemicals, their antioxidant property in order to prove its traditional usages by different tribals of India for various ailments such as inflammatory, hepatic damage, analgesics and antioxidant activity in vitro and in vivo studies. The results of this study indicate that the leaves of G. mauritiana contain phytochemicals of potent pharmacological values. *G. mauritiana* is very rich in variety of secondary metabolites, which might be responsible for various pharmacological properties.

REFERENCES

- 1. Farnsworth NR, Soejarto DD. Global importance of medicinal plants. In: Akerele OVH, Synge H. The Conservation of Medicinal Plants. (eds.) Cambridge (UK): Cambridge University Press; 1991. 25-51.
- 2. Pei SJ. Ethnobotanical approaches of traditional medicine studies: some experience from Asia. Pharm. Biol. 2001; 39:74-79.
- WHO. Traditional Medicine Strategy 2002– 2005. Geneva: World Health Organization; 2002.
- 4. Watt GA. Dictionary of the Economic Products of India. 6 vols. 1889-1886.
- FRLHT. Conserving a National Resource: Need for a National Policy and National Programme on Medicinal Plants Conservation. Unpublished background paper for National Consultation on Formulating a Medicinal Plants Conservation Policy in India. Bangalore: Medplant Conservatory Society; 1996.
- Gorinstein S, Yamamoto K, Katrich E, Leontowicz H, Lojek A, Leontowicz M, Ciz M, Goshev I, Shalev U, Trakhtenberg S. Antioxidative properties of Jaffa sweeties and grapefruit and their influence on lipid metabolism and plasma antioxidative potential in rats. Biosci. Biotechnol. Biochem. 2003; 67:907–910.
- 7. Murugesan M, Balasubramaniam V, Arthi H. Ethno Medical Knowledge of Plants Used by Irula Tribes, Chengal Combai, the Nilgiris, Tamilnadu. Ancient Science of Life. 2005; 24(4):2-4.
- 8. Ganesan S, Venkateshan G, Banumathy N. Medicinal plants used by ethnic group Thottaianaickans of Semmalai hills (Reserved Forest), Tiruchirappalli district, Tamil Nadu. Indian J. Trad. Knowledge. 2006; 5(2):245-252.
- Sandhya B, Thomas S, Isabel W, Shenbagarathai R. Ethnomedicinal plants used by the Valaiyan community of Piranmalai Hills (Reserved Forest), Tamil Nadu, India.- A pilot study. Afr. J. Trad. CAM. 2006; 3(1):101-114.
- 10.Harborne JB. Phytochemical methods.3rd ed. London: Chapman and Hall; 1984.
- 11.Kokate CK, Purohit AP, Gokhale SB. Pharmacognosy, 3rd ed. Pune; Nirali Prakashan; 1995.
- 12.Siddhuraju P, Becker K. Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstic ktree (*Moringa oleifera* Lam.) leaves. J. Agric. Food. Chem. 2003; 51:2144-2155.

- 13.Zhishen J, Mengcheng T, Jianming W. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. Food Chemistry. 1997; 64:555–559.
- 14.Benzie IEF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay. Anal. Biochem. 1996; 239:70-76.
- 15.Blois MS. Antioxidant determinations by the use of a stable free radical. Nature. 1958; 181:1199–1200.
- 16.Ajaiyeoba EO, Abiodun OO, Falade MO, Ogbole NO, Ashidi JS, Happi CT, Akinboye, DO. *In vitro* cytotoxicity studies of 20 plants used in Nigerian antimalarial ethnomedicine. Phytomedicine. 2006; 13:295-298.
- 17.Afolabi F, Afolabi OJ. Phytochemical Constituents of Some Medicinal Plants in South West, Nigeria. IOSR Journal of Applied Chemistry. 2013; 4(1): 76-78.
- 18.Phanghal M, Arya V, Yadav S, Kumar S, Yadav JP. Indigenous knowledge of medicinal plants used by Saperas community of Khetawas, Jhajjar District, Haryana, India. Journal of Ethnobiology and Ethnomedicine. 2010; 28(6):4.
- 19.Akinmoladun EO, Ibukun, Dan-Ologe IA. Phytochemical constituents and antioxidant properties of extracts from the leaves of *Chromolaena odorata*. Scientific Research and Essays, Academic Journals. 2007; 2 (6):191-194.
- 20.Cowman MM. Plant products as antimicrobial agents. Clin. Microbiol. Rev. 1999; 12:561-582.
- 21.Adesokan AA,Yakubu MT, Owoyele BV, Akanji MA, Soladoye A, Lawal OK. Effect of administration of aqueous and ethanolic extracts of *Enantia chlorantha* stem bark of brewer's yeast-induced pyresis in rats. Afri. J. Biochem. Res. 2008; 2(7):165-169.
- 22.Trivedi PC. Medicinal plant, a traditional Knowledge. I. K. International Pvt. Ltd; 2006.
- 23.Jeeva S, Kiruba S, Mishra BP, Venugopal N, Das SSM, Sukumaran S. Weeds of Kanyakumari district and their value in rural life. Indian J. Tradit. Knowledge. 2006; 4:501-509.
- 24.Raja ARD, Jeeva S, Prakash JW, Johnson M, Irudayaraj V. Antibacterial activity of selected ethnomedicinal plants from South India. Asian Pac. J. Trop. Med. 2011; 4(4): 375-378.
- 25.Nisar M, Ali S, Qaisar M. Preliminary Phytochemical Screening of Flowers, Leaves, Bark, Stem and Roots of *Rhododendron arboretum*. Middle-East Journal of Scientific Research. 2011; 10(4):472-476.

- 26. Tirupathi RG, Suresh BK, Ujwal KJ, Sujana P, Veerabhadr R, Sreedhar AS. Anti-microbial principles of selected remedial plants from Southern India. Asian Pac. J. Trop. Biomed. 2011; (4):298-305.
- 27.Maobe MAG, Gatebe E, Gitu L, Rotich H. Preliminary Phytochemical Screening of Eight Selected Medicinal Herbs Used for the Treatment of Diabetes, Malaria and Pneumonia in Kisii Region, Southwest Kenya. European Journal of Applied Sciences. 2013; 5(1):01-06.
- 28. Shivakumar BS, Ramaiah M, Hema MR, Vijay Kumar M, Vaidya VP. Quantitative Determination of Total Content of Phenol, Flavonoid and Tannin in Leaf Extract of *Barlaria Buxifolia* Linn. Am. J. PharmTech Res. 2012; 2(5):417-422.
- 29.Nugroho AE, Malik A, Pramono S. Total phenolic and flavonoid contents, and *in vitro* antihypertension activity of purified extract of Indonesian cashew leave (*Anacardium occidentale* L.). International Food Research Journal. 2013; 20(1):299-305.
- 30.Kantamreddi VSSN, Lakshmi YN, Kasapu VVVS. Preliminary phytochemical analysis of some important Indian plant species. International Journal of Pharma and Bio Sciences. 2010; 1(4):351-357.
- 31.Arora N, Ranawat MS, Arora P. Antihelmintic Activity of Methanolic Extract of Leaves of *Glycosmis pentaphylla*. Journal of Advanced Pharmacy Education & Research. 2010; 1(5):234-237.
- 32.Chander R, Khanna AK, Raj K, Rastogi AK. Antioxidant and Lipid Lowering Activities of Indian Black Tea. Indian Journal of Clinical Biochemistry2005; 20(1):153-159.
- 33.Duh PD, Tu YY, Yen GC. Antioxidant activity of the aqueous extract of harn jyur (*Chrysanthemum morifolium* Ramat). Lebensmittel-Wissenschaft and Technologie. 1999; 32:269-277.
- 34.Guo C, Yang J, Wei J, Li Y, Xu J, Jiang Y. Antioxidant activities of peel, pulp, and seed fractions as determined by FRAP assay. Nutr. Res. 2003; 23:1719-1726.
- 35.Benzie IEF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": The FRAP assay. Anal. Biochem. 1996; 239:70-76.
- 36.Ercan B, Ekrem K. Evaluation of reducing power and radical scavenging activities of water and ethanol extracts from sumac (*Rhus coriaria* L.). Food Research International. 2011; 4:2217–2221.
- 37.Silva CG, Herdeiro RS, Malhias CJ, Panek, AD, Silveira CS, Rodrigues VP et al. Evaluation of

antioxidant activity of Brazilian plants. Pharmacol. Res. 2005; 52:229-233.

- 38.Kumaran A, Karunakaran JR. Antioxidant and free radical scavenging activity of an aqueous extract of Coleus aromaticus. Food Chem. 2006; 97:109-114.
- 39. Jayasri MA, Radha A, Mathew TL. α-amylase and α-glucosidase inhibitory activity of *Costus pictus* D. Don in the management of diabetes. J. Herb. Med. Toxicol. 2009; 3(1):91-94.
- 40.Nishaa S, Vishnupriya M, Sasikumar JM, Hephzibah P, Christabel, Gopalakrishnan V K. Antioxidant activity of ethanolic extract of *Maranta arundinacea* .L tuberous rhizomes. Asian J. Pharm Clin. Res. 2012; 5(4):85-88.