## **Review Article**

#### Polyelectrolyte Complexes: Based Drug Delivery Devices.

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#### **ABSTRACT**

Polyelectrolyte Complexes (PECs) are the association complexes formed due to the electrostatic interactions between oppositely charged particles. It is formed between cationic(Chitosan) polymers and oppositely charged anionic polymers. It forms amorphous aggregates, held together by reversible ionic/hydrophobic cross-links with predominantly random charge-compensation within the complex. The PECs particle sizes and surface charge can be maintained by varying the concentration of the polymers. IR spectroscopy, NMR, thermal analysis, pKa and X-ray diffraction, are used to evaluate the inter polymer complexation. Based on the structures, PECs can be categorized into different subtypes: soluble, colloidal stable and coacervate complexes. PECs can be used as membranes for coating on films and fibers, for isolation and fractionation of proteins, for isolation of nucleic acid, for binding pharmaceutical products, as supports for catalyst and for preparation of microcapsules for drug delivery. This paper gives a concise review of PECs formation and its application in the drug delivery technology. PECs can be used for the both the deliveries of immediate release (Phenytoin sodium) and as well as in controlled release (Propranolol HCL). PECs can be able to modify the release, improve the stability and character of the drug substances. Hence there is a great prospective in utilizing these PECs in biotechnology and Pharmaceutical technology.

**Keywords:** Complexation, drug delivery devices, macromolecules, polyelectrolyte complexes, polymers.

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#### INTRODUCTION

Polyelectrolyte complexes (PECs) are the association complexes formed between oppositely charged particles (e.g. polymerpolymer, polymer-drug and polymer-drugpolymer). These are formed due to electrostatic interaction between oppositely charged polyions [1]. The term polyelectrolyte denotes a class macromolecular compounds, which when dissolved in a suitable polar solvent (generally water), spontaneously acquires or can be made to acquire a large number of elementary charges distributed along the macromolecular chain [2]. In its uncharged state, a polyelectrolyte behaves like any other macromolecules, but the dissociation of even a small fraction of its ionic (side) groups leads to dramatic changes of its

properties [3]. Polyelectrolyte or polysalt formed. complexes are when macromolecules of opposite charge are allowed to interact. The interaction usually involves a polymeric acid or its salt with a polymeric base or its salt. Depending on a variety of factors, it may cause the system to separate into a dilute phase and a concentrated complex coacervate phase, or it may result in a more-or-less compact precipitate or gel. The complexes can also remain in solution. Electrostatic interactions constitute the main attractive forces, but hydrogen bonding, ion dipole and hydrophobic interactions forces. frequently play a significant role in determining the ultimate structures [2].

Polymer complexation leads to a loss of translational and conformational entropy of the polymer chain, which has to be counterbalanced if complexation is to occur. The loss in entropy (per bond formed) is largest for the first bond formed between the two polymers, but is much smaller for subsequent (neighboring) bonds. The enthalpic change (per bond) due to the interaction of the monomeric units however, is nearly constant, and it is easily understood that at a certain critical chain

(or sequence) length, complexation becomes energetically favorable [6,7]. The short range of these interactions (Vander Waals forces) makes a good sterical fit between the polymers essential if complexation is to occur, leading to very high demands on the polymers chemical structure and tacticity. The complexes formed show a very high degree of ordering and crystal like properties, and have quite compact structures [6].

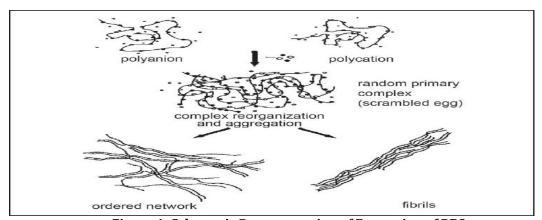


Figure 1: Schematic Representation of Formation of PECs.

The term "scrambled egg structure" is an imaginative description of the complex that is formed after mixing of the two polymers, it is shown in fig1. In such complexes, mainly random change-compensation is found, mostly this situation changes once the complex has been formed [4]. A part from the (quasi-) soluble nonstoichiometric complex (NPEC), most complexation reactions lead to formation of gel particles. The size and composition of such particles depends mainly on the concentration, molecular weights and mixing ratio of the polyelectrolytes. While at high concentrations of the polyelectrolytes, complex formation leads to macroscopic flocculation, the growing of particles can be stopped at a colloidal level in sufficiently diluted systems (C < 0.1 g/ml) [4]. In most cases, the particles have some surface charge as a result of an excess of either one of the polyelectrolytes. Especially, if the ratio of the polyelectrolytes is (exactly) equal to 1, the sign of the surface charge of all particles will be the same, thus

stabilizing the colloidal solution. The (negative) surface charge is responsible for the high biocompatibility of PECs and for the ion selectivity of PECmembranes [8]. Another important feature of PECs is their high swellability in aqueous systems. Most PECs contain well over 85% mass of solvent, comparable to free polymer coils, and if their particle size is kept low. thev provide excellent accessibility to bound moieties [4]. If low molar mass salts are present, the shielding of part of the polymer-bound ions may lower the effective cross-link density of the PEC, thus allowing further swelling. Directly linked to this high swellability is the extremely good permeability for water, gasses and low molar mass electrolytes and organic molecules [9].

# FORMATION OF POLYELECTROLYTE COMPLEXES

Formation Complexion involves 3 steps: [10]

1. Primary complex formation: Secondary binding forces like Coulomb forces are responsible.

- 2. Formation process within intracomplexes: It involves the formation of new bonds and/or correction of the distortion of the polymer chain.
- 3. Intercomplex aggregation process: It involves aggregation of secondary complexes mainly by hydrophobic interactions.

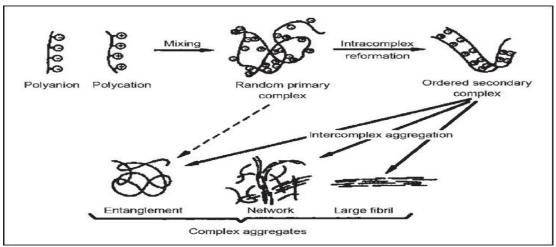


Figure 2: Formation of Polyelectrolyte Complex.

The active components will be encapsulated in the polymer matrix at molecular level. The active substances can be incorporated in to PECs by four ways [12].

- The active substance can be entrapped from the solution during precipitation of the complex, as shown in the fig 2.
- The active substance can be absorbed from the solution and gets incorporated into the already formed complex on contact.
- The active substance may be chemically bound to at least one complex partner and precipitates during complexation.
- ➤ In the last case the active compound itself may act as polyion and form PEC as shown in figure 3.
- The active substance from these PECs will be released either by solution equilibration or by ion exchange mechanism or by charge interaction and slow decomplexation as well as breakdown and dissolution of the complex.

## PECs formed should include the following features:

Amorphous aggregates, held together by reversible ionic/hydrophobic crosslinks with predominantly random charge-compensation within the complex;

- Highly dynamic cross-links, especially when a low molar mass salt and an organic solvent are present in the solution;
- Highly swollen and permeable gel particles (in aqueous solution) forming stable suspensions due to their surface charge.

A variety of PECs can be obtained by changing the chemical structure of component polymers, such as molecular weight, flexibility, functional group structure, charge density, hydrophilic and hydrophobic balance, stereo regularity and compatibility, as well as reaction conditions like pH, ionic strength, concentration, mixing ratio and temperature [11].

#### **Factors affecting the formation of PECs:**

Formation of PECs is influenced by many factors. The formation of PECs is influenced not only by chemical properties like stereochemical fitting, their molecular weight, charge densities, ion site etc, but also by secondary experimental conditions like concentration of polyelectrolytes prior to mixing, their mixing ratio, pH, ionic strength of the solution, mixing order, solvents and temperature etc[4,5]. Changing the ionic strength by addition of modulate the electrostatic salt can interactions in a polyelectrolyte solution [13].

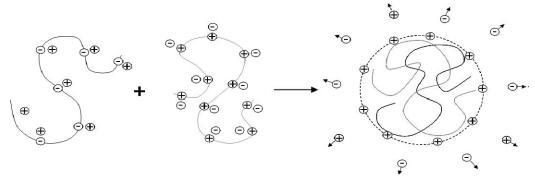


Figure 3: Representation of release of counter ions upon PECs.

**Table 1: Properties of PECs based on Polymer Solution.** 

Polyelectrolyte structure	Solution properties
Molar mass	Polymer concentration
Type of charge group	Ionic strength
Charge density	pH (around the pKa)
Chain architecture	
Hydrophobicity of backbone	Temperature

The electrostatic interactions can be weakened by the addition of inorganic salts into the solutions as shown in (Table1). Thus, an increase of the ionic strength of the solution depresses the complexation between polyions, because of the screening of opposite charges of the macromolecules by low molecular weight ions. By varying the pH environment during PEC formation, degree of ionization of weak polyelectrolyte's can be controlled [14,15]. This was found to affect multilayer properties such as layer thickness, the degree of interpenetration between layers, surface wettability and number of unbound functional groups. Therefore, by choosing the right pH conditions, a platform may be properties found with that advantageous for loading charged small molecules into the film via electrostatic interactions.

#### pH and conductivity study

Polyions possess certain charge when they are present in the aqueous solution. Oppositely charged polyions form an insoluble PEC in the aqueous medium. There is a possible change in the charge that may occur when the PECs interact and form complexes. These interactions and the formation of PECs can be well judged by pH and conductivity studies.

#### Influence of salt on PEC structure

After the changes in ionic strength; swelling deswelling of the PECs occurs immediately, whereas coagulation is a much slower process, dependant on the concentrations of the colloidal particles [58]. Two major effects on the formation of PEC in the presence of salt were found [59]. First, the presence of a very small amount of salt during formation gave a dramatic decrease in level of aggregation, probably due to the less stiff and more coiled structure component. Other studies have shown that the valence (uni- or divalent) of the salt and even the specific ions involved (e.g., K+, Na+, or Li+) seem to be important for the interaction between the polyelectrolyte's [60,61].

## Effect of polymers mixing ratio

The polymers mixing ratio influences the polyelectrolyte complex yield due to the interaction of the two polymers.

## Effect of pH

Polyelectrolyte complexation is a pH sensitive process. At the pH values where the charges are not balanced, a reduction in the interaction between the polymers causes a reduction in the PEC yield.

## Effect of ionic strength

Neutral salts influence the complexation process due to the screening of the charge

groups on the poly electrolytes. Increasing ionic strength results in decreased attraction between the polyions and the tendency to form polyelectrolyte complex.

## **Theoretical aspects of PECs**

Many researchers extensively investigated the properties of the polyelectrolyte and the formation of PECs [14-17]. There have been theories proposed based on electrostatic forces and Flory-Huggins mixing free energies of the polyelectrolytes to explain the mechanism of formation of PECs [18,19]. In general the backbones of the two polymers are not compatible and repel to each other, however, the charge fraction of the polymers determines the type of interaction going to occur between the polymers. When the charge fraction is low, the polymer backbone repulsion (Flory interaction parameter) is dominant and the solution separates in to two phases each containing mostly one of the polymers. At charge fraction, the attractive electrostatic interactions between the polymers dominate and they precipitate to form a complex. In an intermediate range of charge fraction, the equilibrium state can be a meso phase where the two polymers only separate microscopically. Depending on the stoichiometry of the mixture (the relative concentrations, the relative chain lengths and charge densities), one observes mainly two types of complex formations, a macroscopic phase separation between the solvent and the polymers or a partial aggregation of the polymer chains [20].

#### **Characterization of PECs**

The interaction between polymers can be investigated by various methods [21]. Methods like light-scattering infrared spectroscopy (using KBr discs), NMR, thermal analysis, pKa and X-ray powder diffraction are used to evaluate the interpolymer complexation. [22-24]. The different structures of a PEC can be categorized into different subtypes. Three types, soluble are colloidally stable and coacervate complexes, and the type of complex formed is governed by pH, salt, and polymer concentration, and is characterized using turbidimetric or light scattering techniques [54].

Water-soluble PECs Under certain salt conditions, combinations of polyions with

significantly different molecular weights and weak ionic groups in a mixture of nonstoichiometric proportions result in watersoluble PECs (a longer host chain with several small guest chains)[55]. The complex adopts a conformation similar to that of the ladder model, with hydrophilic single-stranded segments and hydrophobic double-stranded segments [56]. presence of a small amount of salt enables rearrangements, which in turn allow the complex to reconfirm to a structure closer to its thermodynamic equilibrium. At slightly higher salt concentrations, a shrinking of the PECs, due to the shielding of polyelectrolyte charges caused by the electrolytes, occurs. A further increase in salt concentration leads to completely complexed, precipitating species; eventually, the precipitates dissolve again and both components exist as free polyelectrolyte's in solution [55-57].

## **Colloidally stable PECs**

PEC formation between strong polyelectrolytes results in highly aggregated or macroscopic flocculated systems [57]. In extremely dilute solutions, the aggregation can be stopped at a colloidal level (with diameters of 10–100nm), and a polydisperse system of nearly spherical particles is usually achieved.

#### **Coacervates**

A coacervate is formed when the mutual binding of opposite polyelectrolytes is of moderate strength as a result of low charge density. The coacervate is a liquid-like, mobile, and reversible structure [62]. The formation of such complexes, for example, from cationic polyacrylamide (CPAM) and sulphonated Kraft lignin, has been investigated, and it was found that the molecular weight of the CPAM was a very important factor for coacervate formation, since a shorter chain can more easily adopt a coiled structure, which will precipitate [63].

## Applications of Drug Delivery Devices with PECs:

PECs have gained much attention in the past few years because of their potential applications. These can be used as membranes[25-28], for coating on films, fibers for isolation and fractionation of

proteins [29,30] for isolation of nucleic acid [31-33] for binding pharmaceutical products as supports for catalyst [34,35] and for preparation of microcapsules for drug delivery [36-37].

Kawashima et al. reported a novel method for the preparation of theophylline granules coated with a PEC of sodium tripolyphosphate and chitosan [37]. The prepared granules containing sodium tripolyphosphate were stirred in an HCL solution of chitosan. During the mixing, the dissolved sodium tripolyphosphate in the granules moved to the surface and reacted with the chitosan, resulting in the formation of a PEC film. The drug-release pattern of the coated granules was of zero-order kinetics and the release rate was significantly reduced compared to that of the original granules.

Shiraishi et al. studied the controlled drug release behaviour of indomethacin by chitosan-PEC [38]. They also optimized the formulation conditions and reported its invivo/in-vitro evaluation studies. PEC of indomethacin prepared by complexation of sodium tripolyphosphate and chitosan. Here the effects of the molecular weights of chitosan hydrolysates on the release and absorption rates of indomethacin from gel beads were examined. The release rates of drug decreased with increasing of molecular weight.

limenez-Kairuz et al. produced and characterized swellable drugpolyelectrolyte matrices (SDPM) carbomer and different basic drugs like atenelol, lidocaine and metoclopramide [39]. The drugs can be loaded in a high proportion on to the polymer and therefore the resulting SDPM material could be diluted with other polymers to modulate delivery properties of SDPM. Matrices of atenolol and lidocaine exhibited vigorous delivery properties with regard to change in proportion of loading drug.

Tapia et al. evaluated the possibility of using mixtures of PECs from both chitosan (CS)-alginate and (CS) carrageenan as prolonged release systems [40]. Different dissolution profiles for diltiazem clorhydrate were obtained by changing the polymer matrix system and the method

used to include these polymers into the formulation (physical mixture or PEC). Drug dissolution profiles from the matrices have been discussed by considering the swelling behavior of the polymers used. It was reported that CS-alginate systems were considered to be better in prolonging the release, when compared to CS-carrageenan systems.

Liao *et al.* developed drug-loaded chitosanalginate fibers by interfacial polyelectrolyte complexation technique [41]. Depending on the component properties, the release time of encapsulated components from these fibers could range from hours to weeks. Dexamethasone was completely released within 2 h, whereas charged compounds such as bovine serum albumin, PDGF-bb, and avidin showed sustained release for 3 w.

Paloma et al. prepared polyionic complexes of CS and poly (acrylic acid) (PAA) in a wide range of copolymer composition and with two kinds of drugs (amoxicillin trihydrate and amoxicillin sodium) [42]. Release from the different complexes was also studied. The swelling behavior of and solute transport in swellable hydrogels were investigated to check the effect of polymer/polymer polymer/drugs and interactions. The electrostatic polymer/polymer interactions took place between the cationic groups from CS and the anionic ones from PAA. The diffusion of amoxicillin trihvdrate was controlled only by the swelling/eroding ratio of the polyionic complexes. The swelling degree of amoxicillin sodium hydrogels was more extensive when compared to the swelling of amoxicillin degree trihvdrate formulations. From the studies it was known that the water uptake was mainly governed by the degree of ionization. Restriction of amoxicillin sodium diffusion could be achieved by polymer/ionized-drug interaction that retards the drug release. Petzold et al. prepared different PECs from poly(diallyl-dimethyl-mmoniumchloride)

poly(diallyl-dimethyl-mmoniumchloride) and two different polyanions and characterized their application as flocculants [50]. The results showed that the most important advantages of PEC were the high velocity of sedimentation and a

very broad range of the optimum flocculation concentration

Win et al. developed PEC gel beads based on phosphorylated chitosan (PCS) controlled release of ibuprofen in oral administration [44]. The PCS gel beads prepared from soluble phosphorylated chitosan by using an ionotropic gelation with counter polyanion, tripolyphosphate (TPP) at pH 4. Surface morphology studies for the prepared beads were done by using SEM. The percentage release of ibuprofen from PCS gel beads was found to be increased as the pH of the dissolution medium increased. The release rate of ibuprofen at pH 7.4 was higher than the release rate at pH 1.4 due to the ionization of phosphate group and higher solubility of ibuprofen at pH 7.4 medium. The ability of the prepared copolymer to be used as drug carrier for colon-specific drug delivery system was estimated using ketoprofen as model drug.

Rolfes *et al.* reported a method of making a solid interpolymer complex for use as a controlled release matrix for oral administration [45]. It involved mixing of two oppositely charged polymers and spray-dried to evaporate the solvent and to prepare solid particles of interpolymer complex. An active agent such as drug can be preferably embedded or encapsulated in the interpolymer complex before spray drying or may be incorporated by suitable means at a later stage.

Mi *et al.* used the enzyme hydrolyzed CS to prepare CS tripolyphosphate and CS polyphosphoric acid gel beads using a polyelectrolyte complexation method for the sustained release of anticancer agent, 6-mercaptopurine[46].

Nandini and Cherng-Ju prepared drug PECs with poly (acrylamido-2-methyl-1propansulfonate sodium-co-methylmethacrylate [47]. They studied and reported that the release kinetics were strongly dependent on the drug solubility rather than on the type of amine in the drug. The release of drugs from the tablets drug-poly(acrylamide-2-methyl-1sulfonate sodium-co-methyl propane methacrylate complex were well described by the dissociation/erosion mechanism.

Albeno et al. obtained a patent preparation of stable water insoluble complexes of poorly soluble compounds molecularly dispersed in water insoluble ionic polymers[48]. The compounds were micro precipitated in the ionic polymers in amorphous form. The complexes according to the present invention significantly increased the bioavailability of poorly soluble therapeutically active compounds. Sabar MH et al. studied variables affecting the formulation of Ketoprofen sustained release oral tablet using Polyelectrolyte complex as a matrix former [43]. It was found that as the strength of the PEC was increased, the release rate of Ketoprofen was found to be increased.

Stefania Ret al. worked on the preparation and characterization of a new type of microparticles based on the complexes between chitosan(CH) and two poly(carboxybetaines) (PCB), in view of their use in medical fields as potential drug carriers.they found that the drug retention capacities were larger in the case of microparticles based on CH-PCB complexes than those based on chitosan[50].

Taira Y et al. concluded by their findings that a low molecular weight chitosan (MW 38kDa) is an effective inhibitor for oxidative stress in various LMW chitosans (MW 10-100kDa) [51]. It was found that the low molecular weight Chitosan-Theophylline anhydrous tablet was a safe and non-toxic extended release tablet with high antioxidant activity.

Sanem AS et al. studied the effect of complexation conditions on xanthan chitosan polyelectrolyte complex gels [52]. For developing hydrogels with desired mechanical and controlled properties the characterization of factors contributing to the cross-linking density of xanthan-chitosan network were important. The results obtained from Swelling degree (SD) and Differential Scanning Colorimetry (DSC) showed that the cross-linking density of xanthanchitosan network was dependent on the complexation conditions. There complete cross-linking of the hydrogel capsules at all the conditions, when the initial xanthan solution concentration was at 1.5 % (w/v). The increase in the xanthan

concentration affected the degree of swelling of the hydrogel at two different chitosan concentrations. The effect of chitosan solution pH on the degree of swelling is more pronounced at 0.7% (w/v) than at 1.0% (w/v) chitosan concentration. The conformational changes of chitosan polymer chains, which are dependent on solution pН, were critical determining the crosslinked network structure which affects the SDs of the gels. It was concluded that pH and concentration effects on the xanthan-chitosan network properties are depended on each other, which can be modulated by changing controllable operationally parameters. concentration and especially xanthan chitosan solution pH.

Rishabha M et al. worked on the preparation. characterization and evaluation of chitosan- gum arabic Coacervates as excipients in Fast dissolving/ disintegrating dosage form.[53] The study revealed that the presence and interaction of two different polymer solutions may lead to formation of a characteristic composite which can enhance the release of drug from the dosage form. proved that formulating fast disintegrating tablets using Chitosan-gum arabic coacervates can give a dosage form

that can be used for the treatment of chronic disorders at the initial stage of chronic attack.

Baloglu E *et al.* evaluated the performance of swellable polymers (chitosan and carrageenan) in the form of layered matrix tablets to provide controlled therapeutic effect of Metaprolol tartrate for twice daily administration[64]. They found that the carrageenan three layered matrix tablet formulation is an effective and promising drug delivery system for twice daily administration of Metaprolol tartrate.

Ala F. Eftaiha *et al.* formulated Metronidazole with chitosan and xanthan gum hydrophilic polymers has proven to be a well representative example of a sustained release solid dosage form preparation [65]. The formulation was found to attain prolonged

*in vitro* drug release, strong bioadhesivity, when tested on a sheep duodenum, and a high *in vivo* bioavailability thus it was found to be superior in all aspects of *in vitro-in vivo* analysis.

There are a number of pharmaceutical applications of polyelectrolytes, such as in controlled release systems are listed in table 2, for the enzyme and cell support, for different types of tissue reconstitution etc.

Table 2: List of API formulated along with PECs.

API	Cationic	Anionic	Formulation	References
Metaprolol tartrate	chitosan	carrageenan	CR	64
Phenytoin sodium	chitosan	Gum arabic	FDT	54
Glipizide	chitosan	Xanthan gum	SR	69
Phenytoin sodium	chitosan	Sodium alginate	FDT	53
Ketoprofen	chitosan	Carbopol- 940	SR	50
Diltiazem HCL	chitosan	Gum ghatti	SR	68
Diltiazem HCL	chitosan	Sodium alginate Sodium CMC Carbopol 940	Hydrogel	67
Metronidazole	chitosan	Xanthan gum	SR	65
Propronolol HCL	chitosan	Xanthan gum	CR	66

CR: Controlled release, SR: Sustained release, FDT: Fast disintegrating Tablets.

#### CONCLUSION

A broad research is going on, in the area of polyelectrolyte complexes (PECs). Most of the PECs are formed with cationic Chitosan (natural polymer) i.e. with ionic polymers. There is a great prospective in utilizing these PECs in biotechnology and Pharmaceutical technology. PECs have multiple applications according to the ionic interactions, when the polymers are combined. It is used for sustaining and controlling the release of drug and also for fast disintegrating dosage forms. By their capacity to entrap the drug at molecular level, PECs can be able to modify the release, improve the stability and character of the drug substances, which ensures their great potential in designing novel drug delivery devices.

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## REFERENCES

- 1. Krishnendu R, Hai-Quan M, Shau-Ku H, Kam WL. Oral gene delivery with chitosan DNA nanoparticles generates immunologic protection in a murine model of peanut allergy. Nature Med 1999;5:387-91.
- Mark HF, Bikales NM, Overberger CG, Menges G. Encyclopedia of polymer science. 2 nd ed, Vol. 2. NY: John Wiley and Sons, A Willey Interscience Publishers; 1987. p. 739.
- 3. Encyclopedia of polymer science and technology. 2 nd ed, Vol. 2. Chichester: Wiley; 1988. p. 739-829.
- 4. Philipp B, Doutzenberg H, Linow KJ, Koetz J, Dowydoff W. Polyelectrolyte recent development and open problems. Prog Polym Sci 1989; 14:91.
- 5. Kabanov VA, Zezin AB, Ragacheva VB, Guskina N, Goethals EJ, Valde MV. Properties of polyelectrolyte complexes containing poly (N-tertbutylaziridine). Makromol Chem 1986;187:1151.
- 6. Brinkhuis RH, Schouten AJ. Thin-film behaviour of poly(methyl methacrylates): Characteristics of the poly (methyl methacrylate) monolayer stereocomplexation process. Macromol 1992; 25:2732-8.
- 7. Kabanov VA, Papisov IM. Formation of complexes between complimentary synthetic polymers and oligomers in dilute solution review. Polym Sci 1979;21:261-307.
- 8. Kikuchi Y, Kubota N. Structure properties and Na+ transport membrane of polyelectrolyte

- complexes consisting of glycol chitosan and poly (vinyl sulfate). Bull Chem Soc Jpn 1987;60:375
- 9. Smolen VF, Hahman DE. A water membrane hypothesis behavior of hydrated polycation-polyanion salt complexed membranes as appeared lipoidal barriers to solute transport. J Colloid Interface Sci 1973;42:70-8.
- 10. Krone V, Magerstadt M, Walch A, Groner A, Hoffmann D. Pharmacological composition containing polyelectrolyte complexes in microparticulate form and at least on active agent. United States patent 5,700,459, Dec 23, 1997.
- 11. Dumitriu S, Chornet E. Inclusion and release of proteins from polysaccharide based polyion complexes. Adv Drug Deliv Rev 1998;31:223-46.
- 12. Alexander K, Monica OD. Precipitation of oppositely charged polyelectrolytes in salt solutions. J Chem Phys 2004; 20:404-12.
- 13. Joanny JF, Castelnovo M. Polyelectrolyte adsorption and multiplayer formation. In: Decher G. Schlenoff JB, editors. Multilayer Thin Films. Weinheim: Wiley-VCH; 2002. p. 87-97.
- 14. Webster L, Huglin MB, Robb ID. Complex formation between polyelectrolytes in dilute aqueous solution. Polymer 1997; 38:1373-80.
- Webster L, Huglin MB. Observations on complex formation between polyelectrolytes in dilute aqueous solution. Eur Polym J 1997; 339:1173-7.
- 16. Burgess DJ. Practical analysis of complex coacervate systems. J Colloid Interfacial Sci 1990; 140:227-38.
- 17. Overbeek JT, Voorn MJ. Phase separation in polyelectrolyte solutions: Theory of complex coacervation. J Cell Comp Physiol 1957; 49:7-26.
- 18. Xavier C, Jean-Francois J. Adsorption of polyelectrolyte solutions on surface: A Debye-Huckel theory. J Phys II France 1996; 6:1669-86.
- 19. Tsuchida E. Formation of polyelectrolyte complexes and their structures. J Macromol Sci Pure Appl Chem 1994; A31:1-15.
- 20. Kumar V, Yang T, Yang Y. Interpolymer complexation. I. Preparation and characterization of a polyvinyl acetate phthalate-polyvinylpyrrolidone (PVAP-PVP) complex. Int J Pharm 1999; 188:221-32.
- 21. Herbert D, Light scattering studies on polyelectrolyte complexes. Macromolecular Symposia 2001;162:1-22.
- 22. Natalia VP, Nickolay VT. Structure and dynamics of the polyelectrolyte complex formation. Macromol 1997; 30:4897-904.

- 23. Anlar Hapan Y, Goven O, GA, Dalkara T, Hincal A. Formulation and in-vitro-in-vivo evaluation of buccoadhesive morphine sulfate tablets. Pharm Res 1994: 11:231-6.
- 24. Sato H, Maeda M, Nakajima A. Mechanochemistry and permeability of polyelectrolyte complex membranes composed of poly (vinyl alcohol) derivatives. J Appl Polym Sci 1979; 23:1759-67.
- 25. Harris EL, Angal S. Protein purification methods: A practical approach. New York: Oxford University Press; 1993.
- Senuma M, Kuwabara S, Kaeriyama K, Hase F, Shimura Y. Polymer complex from copolymers of acrylonitrile and ionic vinyl benzyl compounds. J Appl Polym Sci 1986; 31:1687-97.
- 27. Yamamoto H, Horita C, Senoo Y, Nishida A, Ohkawa K. Polyion complex fiber and capsule formed by self-assembly of chitosan and gellan at solution interfaces. Macromol Chem Phys 2000; 201:84-92.
- 28. Dubin PL, Gao J, Mattison K. Protein purification by selective phase separation with polyelectrolytes. Sep Purif Methods 1994; 23:1-16.
- 29. Hirouki Y, Takeshi K. Adsorption of BSA on cross-linked chitosan: The equilibrium isotherm. Chem Eng Japan 1989; 41:B11-5.
- 30. Atkinson JG. Precipitation of nucleic acids with polyethyleneimine and the chromatography of nucleic acids on immobilized polyethyleneimine. Biochem Biophys Acta 1973; 308:41-52.
- 31. Jendrisak J. In: Burgerss R, editors Protein purification: Micro to macro. New York: Alan R Liss Inc; 1987. p. 75-97.
- 32. Cordes RM, Sima WB, Glatz CE. Precipitation of nucleic acids with poly (ethyleneimine). Biotechnol Prog 1990; 6:283-5.
- 33. Chen J, Jo S, Park K. Polysaccharide hydrogels for protein drug delivery. Carbohydr Polym 1995; 28:69-76.
- 34. Dautzenberg H, Kotz J, Linow KJ, Philipp B, Rother G. Static light scattering of polyelectrolyte complex solutions. In: Dubin P, Bock J, Davis R, Schulz DN, Thies C, editors. Macromolecular complexes in chemistry and biology. Berlin: Springer Verlag; 1994. p. 119-33.
- 35. Murakami R, Takashima R. Mechanical properties of the capsules of chitosan-soy globulin polyelectrolyte complex. Food Hydrocolloids 2003; 17:885-8.
- Artur B, David H. Carrageenan-oligochitosan microcapsules: Optimization of the formation process. Colloids Surf B: Biointerfaces 2001; 21:285-98.

- 37. Kawashima Y, Handa T, Kasai A, Takenaka H, Lin SY, Ando Y. Novel method for the preparation of controlled-release theophylline granules coated with a polyelectrolyte complex of sodium polyphosphate-chitosan. J Pharm Sci 1985; 74:264-8.
- 38. Shiraishi S, Imai T, Otagiri M. Controlled release of Indomethacin by chitosan polyelectrolyte complex: Optimization and *in vivo/in vitro* evaluation. J Control Release 1993; 25:217-25.
- 39. Jimenez-Kairuz AF, Llabot JM, Allemandi DA, Manzo RH. Swellable drug-polyelectrolyte matrices (SDPM): Characterization and delivery properties. Int J Pharm 2005; 288:87-99.
- 40. Tapia C, Escobar Z, Costa E, Sapag-Hagar J, Valenzuela F, Basualto C, *et al.* Comparative studies on polyelectrolyte complexes and mixtures of chitosan-alginate and chitosan-carrageenan as prolonged diltiazem clorhydrate release systems. Eur J Pharm Biopharm 2004; 57:65-75.
- 41. Liao I-C, Wan ACA, Yim EK, Leong KW. Controlled release from fibers of polyelectrolyte complexes. J Control Release 2005; 104:347-58.
- 42. Paloma M, de la T, Yewande E, Guillermo T, Susana T. Release of amoxicillin from polyionic complexes of chitosan and poly (acrylic acid): Study of polymer/polymer and polymer/drug interactions within the network structure. Biomaterials 2003; 24:1499-506.
- 43. M.H. Sabar, L.H.Samein and Hayder B.Sahib Some variables Affecting the Formulation of Ketoprofen Sustained Release Oral Tablet using Polyelectrolyte Complex as a Matrix Former. Journal of Pharmacy and Allied Health Sciences 2011; 1:1-15.
- 44. Win PP, Shin-ya Y, Hong KJ, Kajiuchi T. Formulation and characterization of pH sensitive drug carrier based on phosphorylated chitosan (PCS). Carbohydr Polym 2003; 53:305-10.
- 45. Rolfes H, Van Der Merve TL, Truter PA. Method of making controlled release particles of complexed polymers. United States Patent 6,221,399, 2001.
- 46. Mi FL, Shyu SS, Kuan CY, Lee ST, Lu KT, Jang SF. Chitosan-Polyelectrolyte complexation for the preparation of gel beads and controlled release of anticancer drug: I: Effect of phosphorous polyelectrolyte complex and enzymatic hydrolysis of polymer. J Appl Polym Sci 1999; 74:1868-79.
- 47. Nandini K, Cherng-ju K. Drug release from drug-polyanion complex tablets:

- Poly(acrylamido-2-methyl-1-propanesulfonate sodium-co-methyl methacrylate). J Control Release 1999; 57:141-50.
- 48. Albiono AA, Phuapradit W, Sandhu HK, ShahNH.Stable Complexes of porly soluble Compounds in ionic Polymers.United states Patents 6,221,399,2001.
- 49. Petzold G, Nebel A, Buchhammer H-M, Lunkwitz K. Preparation and characterization of different polyelectrolyte complexes and their application as flocculants. Colloid Polym Sci 1998; 276:125-30.
- 50. Stefania R, Silvia V and Cristina DV New Drug Delivery Systems Based on Polyelectrolyte Complexes. Rev.Roum.Chim 2010, 55(10), 659-666.
- 51. Taira Y, Makoto A, Yuko K, Toshiyuki H, Junzo H, Nobuyuki K and Hisao Tomida. Useful Extend Release Chitosan Tablets with High Anto oxidant Activity. Pharmaceutics 2010, 2,245-257.
- 52. Sanem Argin- Soysal, Peter Kofinas, Y. Martin Lo. Effect of complexation conditions on xanthan-chitosan polyelectrolyte complex gels. Food Hydrocolloids 2009; 23: 202–209.
- 53. Rishabha M, Pranati S. Preparation, Characterization and Application of Chitosan-Alginate based Polyelectrolyte Complex as fast Disintegrating Drug Delivery Carrier. Polimery Medycynie 2011; T.41, Nr 3.
- 54. Tsuchida, E. and K. Abe Interactions between macromolecules in solution and intermacromolecular complexes. Advances in Polymer Science. H.-J. Cantow, G. Dall'Asta et al., Springer-Verlag Berlin Heidelberg, Germany. 1982; 45: 1-130.
- 55. Kabanov, V. Fundamentals of polyelectrolyte complexes in solution and the bulk. Multilayer Thin Films. G. Decher and J. B. Schlenoff, Wiley-VCH Verlag GmbH, Weinheim, Germany 2003; 47-86.
- 56. Kabanov, V. A. and A. B. Zezin. Soluble interpolymeric complexes as a new class of synthetic polyelectrolytes. Pure Appl. Chem. 1984; 56(3): 343-354.
- 57. Thunemann AF, Muller M, Dautzenberg H, Joanny JF and Lowen H Polyelectrolyte complexes. Polyelectrolytes with defined molecular architecture II. M.Schmidt, Springer-Verlag Berlin Heidelberg, Germany. 2004; 166: 113-171.
- 58. Dautzenberg H. and Rother G. Response of polyelectrolyte complexes to subsequent addition of sodium chloride: Time-dependent static light scattering studies. Macromol. Chem. Phys. 2003; 205(1): 114-121.
- 59. Dautzenberg H. Polyelectrolyte complex formation in highly aggregating systems. 1.

- Effect of salt: Polyelectrolyte complex formation in the presence of NaCl. Macromolecules 1997; 30(25): 7810-7815.
- 60. Michaels AS, Mir L. and Schneider NS. A conductometric study of polycation polyanion reactions in dilute aqueous solution. J. Phys. Chem. 1965; 69(5): 1447-1455.
- 61. Sukhishvili SA, Kharlampieva E and Izumrudov V. Where polyelectrolyte multilayers and polyelectrolyte complexes meet. Macromolecules 2006; 39(26): 8873-8881.
- 62. Biesheuvel PM. and Cohen Stuart MA Cylindrical cell model for the electrostatic free energy of polyelectrolyte complexes. Langmuir 2004; 20(11): 4764-4770.
- 63. Vanerek, A. van de Ven and TGM. Coacervate complex formation between cationic polyacrylamide and anionic sulfonated kraft lignin. A Colloid Surface 2006; 273: 55-62.
- 64. Baloglu E and Senyigit T. A Design and Evaluation of Layered Matrix Tablet Formulations of Metoprolol Tartrate. AAPS Pharm Sci Tech. 2010; 11(2):563-573.
- 65. Alaa FE, Nidal QI, Rashid S, Mayyas M. AI Remawi, Munther R.AI Shami, Tawfiq AA and Badwan AA. Bioadhesive Controlled Metronidazole Release Matrix Based on Chitosan and Xanthan Gum Mar. Drugs 2010; 8:1716-1730.
- 66. Thawatchai P. Effect of particle size of chitosan on drug release from Layered Matrix System comprising Chitosan and xanthan gum. Thai Pharm Health Sci J 2008; 3(1): 1-11.
- 67. Saleem MA, Azharuiddin Sk, Sadat A, Patil CC. Studies on different Chitosan Polyelectrolyte complex hydrogels for modified release of Diltiazem hydrochloride. International J of Pharmacy and Pharmac Sci 2010; 2(4): 64-67.
- 68. Mahesh RM, Jagadeeswara RD, Moin A and Shiva kumar HG. Formulation of sustained release matrix tablet using Chitosan/Ghatti gum polyelectrolyte complex. Der pharmacia Lettre 2011;3(2): 119-128
- 69. Ramanji RT, Dhachinamoorthi D, Chandrasekhar KB. Independent release behaviour of Glipizide matrix release tablets containing chitosan and xanthan gum. International J of Pharm and Biomed Res 2010; 1(2):64-70.