

# Serotypes and Serogroups Implicated in Bacterial Meningitis in Confirmed Cases across Northern Ghana

Enoch Weikem Weyori\*, Nkrumah Bernard, Braimah Baba Abubakari, Adadow Yidaana, Adatsi Richard Kojo, Sylvester Mensah, Valentine Koyiri Cheba

Department of Microbiology, Public Health and Reference Laboratory, Sunyani, Ghana

## Research Article

**Received:** 25-Sep-2023,  
Manuscript No. JMB-23-114662;  
**Editor assigned:** 28-Sep-2023,  
PreQC No. JMB-23-114662(PQ);  
**Reviewed:** 12-Oct-2023, QC No.  
JMB-23-114662; **Revised:** 19-Oct-  
2023, Manuscript No. JMB-23-  
114662(R); **Published:** 26-Oct-  
2023, DOI: 10.4172/2320-  
3528.12.4.001

**\*For Correspondence:**

Enoch Weikem Weyori,  
Department of Microbiology,  
Public Health and Reference  
Laboratory, Sunyani, Ghana  
**E-mail:** eweyori@gmail.com

**Citation:** Weyori EW, et al.  
Serotypes and Serogroups  
Implicated in Bacterial Meningitis  
in Confirmed Cases across  
Northern Ghana. RRJ Microbiol  
Biotechnol. 2023;12:001.

**Copyright:** © 2023 Weyori EW, et  
al. This is an open-access article  
distributed under the terms of the  
Creative Commons Attribution  
License, which permits  
unrestricted use, distribution, and

## ABSTRACT

**Background:** Bacterial meningitis is a dangerous infection that can kill children and adults alike. An estimated 1.2 million instances of bacterial meningitis are anticipated to occur globally each year, despite the discovery of polysaccharide and conjugate vaccines in recent years. The study aimed to find out which bacteria isolates are linked to meningitis and to compare the culture method to the RT-PCR method.

**Methods:** This study examined data on bacterial meningitis from the Ghana health service public health division from 2015 to 2019. Patient's information was collected from case-based forms held at the tamale public health and reference laboratory. The data from the case-based forms was transcribed into a pre-designed microsoft excel template. For analysis, the data was cleaned and loaded into Minitab version 18.

**Results:** There were 2446 CSM cases documented in all, 34.4% were confirmed. Males (52.7%) were more suspected than females (47.3%). Age group 15 years-44 years were affected most (37.5%). The predominant pathogens were *Neisseria meningitidis* (W135) and *Streptococcus pneumoniae* (St.1) with NMX, St.14, St.19, and St.12F/12A/12B as new emerging strains. Notwithstanding that, low NPV (72.6%), NLR (0.69), and sensitivity (31.8%) of culture method for detected pathogens of bacterial meningitis were found to produce a statistically significant false negatives compared to the gold standard (RT-PCR).

**Conclusion:** Emergence of new strains of bacterial meningitis and the false negative results chained out by the culture method is alarming.

**Keywords:** Bacterial meningitis; *Streptococcus pneumoniae*; *Neisseria meningitidis*; *Haemophilus influenzae*; Sickness; Bacteria

reproduction in any medium,  
provided the original author and  
source are credited.

## INTRODUCTION

Meningitis is a devastating disease with a high case fatality rate and leading to serious long-term complications. It causes up to 20% of deaths in victims and accounts for extreme neurological sequelae (influencing 12%-15% of survivors) in non-industrial nations. Even though several bacteria might cause the sickness, three isolates (*Streptococcus pneumoniae* (Sp), *Neisseria meningitidis* (Nm), and *Haemophilus influenzae* type b (Hib)) contribute to a significant proportion accounting for more than one million mortalities every year, principally among kids and adults. In the countries from Ethiopia to Senegal, where incidence rates of the disease are significantly greater, an epidemic of bacterial meningitis disease is a major public health hazard. Meningococcal infections, responsible for severe annual outbreaks of meningitis, are the most feared, with an incidence rate as high as 1000 cases per 100000 population unlike *Streptococcus pneumoniae* that contributes approximate 17 cases per 100000 population annually with a CFR exceeding 73% in children under five years <sup>[1]</sup>. Though *Haemophilus influenzae* meningitis is rare in populations of adolescents and adults, rates of meningitis due to Hib are highest in children less than five years of age, with an estimated incidence rate of 31 cases per 100000 accounting for higher mortality rates than found in *Neisseria meningitidis*.

Notwithstanding that, an additional concern is related to the recent change in the epidemiological pattern of meningococcal outbreaks due to the emergence of serogroup W135 as an epidemic strain as well as the serotypes of *Streptococcus pneumoniae* in populations of developing countries due to the enhancement of the PCR assay technique. The examination of bacterial meningitis relies upon the blood and CSF tests conducted by rapid RDT test, culture and PCR methods. Observational anti-microbial treatment is to be started quickly founded on the clinical discoveries and rapid test results. For a successful treatment of bacterial meningitis, the microorganisms and their anti-toxin weakness examples ought to be quickly identified <sup>[2]</sup>. While the CSF culture is the highest quality level in the finding of bacterial meningitis, the low bacterial development rates especially in the patients who have gotten anti-microbial treatment before the lumbar cut (LP) required the advancement of new test methods. Nucleic corrosive enhancement tests, for example, the PCR can distinguish limited quantities of microbe DNA freely from the development of the microorganism causing the disease.

In the light of these revelations, implementation of the PCR assay for diagnosis and surveillance of bacterial meningitis in Ghana, Niger, and Burkina Faso, among the most stricken countries, is an important milestone for the meningitis national control programmes. This paper reports on the bacterial isolates implicated in the meningitis as well as benefits of the PCR assay compared with culture diagnostic technique across the meningitis belt in Ghana.

## MATERIALS AND METHODS

### Study design

The research constitutes a retrospective institutional cross-sectional study conducted within the northern zone of Ghana taking into account data from 2015 to 2019.

### Study setting

The source of data for the study included the zonal public health laboratory, tamale under the Ghana health service, northern regional directorate. ZPHL is a reference laboratory for bacterial meningitis in Ghana. It also serves as the reference public health laboratory for the northern zone with respect to diseases of public health importance.

### Study population

The study population was made up of all suspected, probable, and confirmed cases of bacterial meningitis. All cases from upper east, upper west, savannah, north-east, and northern regions were part of the geographical composition of the study.

### Clinical case definition

An illness with sudden onset of fever ( $>38.5^{\circ}\text{C}$  rectal or  $>38.0^{\circ}\text{C}$  axillary) with one or more of the following symptoms: Neck stiffness, altered consciousness, additional meningeal sign, or petechial or purpureal rash (GHS, 2019). In patients less than one year, suspect meningitis when fever accompanied by bulging fontanelle (GHS, 2019).

### Laboratory criteria for diagnosis

Within 72 hours of clinical manifestation, positive CSF antigen detection or positive culture at the district or facility level was validated by PCR test (GHS, 2019).

**Case classification:** Suspected case is one that meets the clinical case definition; probable case is one that meets the clinical case definition and has turbid CSF (with or without positive gram stain) or is part of an ongoing epidemic with an epidemiological link to a confirmed case; and confirmed case is one that has laboratory confirmation.

**Sample size determination:** From 2015 to 2019, all probable instances of bacterial meningitis were included in the study, hence no sample size was calculated.

**Data analysis:** Data was retrieved from the case surveillance forms and entered into a microsoft excel spreadsheet. For analysis, the data was cleaned, validated, and exported to minitab version 18. A descriptive analysis was carried out and the results were reported in frequency tables. Statistical significance testing was done using the paired samples sign test with the sensitivity and specificity of culture method determined; Positive Predictive Value (PPV) and Negative Predictive Value (NPV) were calculated. Likelihood ratios for PPV and NPV were also estimated to determine the accuracy of the culture method.

**Inclusion criteria:** The study included all patients who met the case definition criteria for bacterial meningitis as determined by the disease control and surveillance unit of the Ghana health service.

**Exclusion criteria:** All patients with inadequately filled case investigation forms, cases that were not having samples accompanying the case investigation forms and cases that had no culture and PCR results.

**Patient and public involvement:** The study had no patient or public participation because it was a retrospective study.

### RESULTS

Age 15 years-44 years had majority of suspect and confirmed cases across the trend years; males had greater proportion for suspected (1288; 52.7%) and confirmed (466; 36.2%) cases compared to females with 47.3% of suspected cases and 32.5% of confirmed cases [3-5]. Most suspected cases were from northern (993; 40.6%) and upper west (993; 40.6%) regions with majority confirmed cases from northern region (427; 43.0%) (Table 1).

**Table 1.** Demographic characteristics of cases.

| Variable     | Level      | Negative (n=1604) | Positive (n=842) | Total (n=2446) |
|--------------|------------|-------------------|------------------|----------------|
| Age category | <5         | 316(71.5)         | 126(28.5)        | 442(18.1)      |
|              | 5-14       | 358(53.2)         | 315(46.8)        | 673(27.5)      |
|              | 15-44      | 665(67.8)         | 316(32.2)        | 981(40.1)      |
|              | 45-59      | 136(74.7)         | 46(25.3)         | 182(7.4)       |
|              | 60+        | 129(76.8)         | 39(23.2)         | 168(6.9)       |
| Sex          | Male       | 822(63.8)         | 466(36.2)        | 1288(52.7)     |
|              | Female     | 782(67.5)         | 376(32.5)        | 1158(47.3)     |
| Regions      | Northern   | 566(57.0)         | 427(43.0)        | 993(40.6)      |
|              | Upper east | 283(61.5)         | 177(38.5)        | 460(18.8)      |
|              | Upper west | 755(76.0)         | 238(24.0)        | 993(40.6)      |

#### Sex and age distribution

Table 2 indicates majority (466; 55.3%) were males with only 2015 (30; 51.7%) having higher counts for females. Nevertheless, the age categorization saw age 14 years-15 years with the highest positive cases in 2015 (37.9%) to 2017 (37.2%) compared to 15 years-44 years having the highest positivity rate in 2018 (38.4%) to 2019 (44.4%).

**Table 2.** Sex and age distribution of confirmed cases across the years.

| Variables |        | 2015 (n=58) | 2016 (n=151) | 2017 (n=301) | 2018 (n=172) | 2019 (n=160) | Total (n=842) |
|-----------|--------|-------------|--------------|--------------|--------------|--------------|---------------|
| Sex       | Male   | 28(48.3)    | 83(55.0)     | 173(57.5)    | 102(59.3)    | 80(50.0)     | 466(55.3)     |
|           | Female | 30(51.7)    | 68(45.0)     | 128(42.5)    | 70(40.7)     | 80(50.0)     | 376(44.7)     |
| Age       | <5     | 9(15.5)     | 26(17.2)     | 56(18.6%)    | 24(14.0%)    | 13(8.0)      | 128(15.2)     |
|           | 5-14   | 22(37.9)    | 52(34.4)     | 112(37.2%)   | 65(37.8%)    | 54(33.8)     | 305(36.2)     |
|           | 15-44  | 21(36.2)    | 51(33.8)     | 97(32.2%)    | 66(38.4%)    | 71(44.4)     | 306(36.3)     |
|           | 45-59  | 4(6.9)      | 8(5.3)       | 14(4.7%)     | 10(5.7%)     | 10(6.3)      | 46(5.5)       |
|           | 60+    | 2(3.4)      | 4(2.6)       | 22(7.3%)     | 7(4.1%)      | 12(7.5)      | 45(5.3)       |

#### Isolates implicated in bacterial meningitis

Table 3 shows the real time RT-PCR confirmed bacterial causative agents of meningitis over the years studied. It was denoted that, from the 842 cases, *Streptococcus pneumonia* (Spn) accounted for 53.0% (446) of all confirmed

cases of bacterial meningitis trailed by *Neisseria meningitidis* (Nm) (43.7%; 368) and *Haemophilus influenzae* (3.3%; 28) from 2015 to 2019. Except for 2015 and 2016 where *Neisseria meningitidis* was equal and more as causative agent of bacterial meningitis, *Streptococcus pneumoniae* was implicated in most of the cases with *Haemophilus influenzae* accounting for a small proportion of the cases.

**Table 3.** RT-PCR pathogens detected over the trend years.

| RT-PCR results                  | 2015<br>(n=58) | 2016<br>(n=151) | 2017<br>(n=301) | 2018<br>(n=172) | 2019<br>(n=160) | Total<br>(n=842) |
|---------------------------------|----------------|-----------------|-----------------|-----------------|-----------------|------------------|
| <i>Neisseria meningitidis</i>   | 28(48.3)       | 97(64.2)        | 125(41.5)       | 62(36.0)        | 56(35.0)        | 368(43.7)        |
| <i>Streptococcus pneumoniae</i> | 28(48.3)       | 51(33.8)        | 166(55.1)       | 102(59.3)       | 99(61.9)        | 446(53.0)        |
| <i>Haemophilus influenzae</i>   | 2(3.4)         | 3(2.0)          | 10(3.3)         | 8(4.7)          | 5(3.1)          | 28(3.3)          |

### Serogroups of *Neisseria meningitidis*

Table 4 indicates the cross tabulation of the serogroups of *Neisseria meningitidis* causing bacteria meningitis from 2015 to 2019. Of the 368 cases of *Neisseria meningitidis*, serogroup NmW accounted for about 84.0% (309) of the cases followed by serogroups NmX (11.7%; 43), NmNG (2.7%; 10), NmC (1.1%; 4) and NmB (0.5%; 2) notwithstanding that, indications shows that 2017 had the highest frequency of *Neisseria meningitidis* cases with 125 representing 34.0 percent whiles 2015 recorded the least frequency of cases with 28 (7.6%).

**Table 4.** RT-PCR results of *Neisseria meningitidis* serogroups.

| Isolates (Nm) | Years          |                |                 |                |                | Total<br>(n=368) |
|---------------|----------------|----------------|-----------------|----------------|----------------|------------------|
|               | 2015<br>(n=28) | 2016<br>(n=97) | 2017<br>(n=125) | 2018<br>(n=62) | 2019<br>(n=56) |                  |
| NmB           | 0(0.0)         | 0(0.0)         | 1(50.0)         | 1(50.0)        | 0(0.0)         | 2(0.5)           |
| NmC           | 0(0.0)         | 1(25.0)        | 3(75.0)         | 0(0.0)         | 0(0.0)         | 4(1.1)           |
| NmW           | 28(9.1)        | 95(30.7)       | 119(38.5)       | 39(12.6)       | 28(9.1)        | 309(84.0)        |
| NmX           | 0(0.0)         | 1(2.3)         | 2(4.7)          | 17(39.5)       | 23(53.5)       | 43(11.7)         |
| NG            | 0(0.0)         | 0(0.0)         | 0(0.0)          | 5(50.0)        | 5(50.0)        | 10(2.7)          |

### Serotypes of *Streptococcus pneumoniae* (2015-2019)

Table 5 relates to the cross tabulation of polymerase chain reaction results for serotypes of *Streptococcus pneumoniae* causes of meningitis over the five years period under review. In indication, Spn Serotype 1 (St.1) recorded the majority of cases representing almost half of the cases (49.6%). Furthermore, relative to Spn cases recorded over the period, it was realized that St.18C/18B/18A/18F had the least counts of 1 (0.2%) case. Other cases like St.12F/12A/12B/44/4 and St.14 had frequencies of 45 (10.1%) and 21(4.7%) respectively. It was worth noting that non-typeable cases of *Streptococcus pneumoniae* cases had a significant frequency of 61 (13.7%). Notwithstanding, the year that recorded the highest frequency of Spn cases was 2017 with 166 (37.2%) whiles 2015 recorded the least with 28 (6.3%).

**Table 5.** RT-PCR results serotypes against the trend years.

| RT-PCR results      | Years       |             |              |              |             | Total (446) |
|---------------------|-------------|-------------|--------------|--------------|-------------|-------------|
|                     | 2015 (n=28) | 2016 (n=51) | 2017 (n=166) | 2018 (n=102) | 2019 (n=99) |             |
| St.1                | 17(7.7)     | 31(14.0)    | 87(39.4)     | 56(25.3)     | 30(13.6)    | 221(49.6)   |
| St.11A/11D          | 0(0.0)      | 0(0.0)      | 0(0.0)       | 2(40.0)      | 3(60.0)     | 5(1.1)      |
| St.12F/12A          | 0(0.0)      | 0(0.0)      | 2(50.0)      | 2(50.0)      | 0(0.0)      | 4(0.9)      |
| St.12F/12A/12B      | 0(0.0)      | 0(0.0)      | 0(0.0)       | 2(14.3)      | 12(85.7)    | 14(3.1)     |
| St.12F/12A/12B/44/4 | 0(0.0)      | 10(22.2)    | 21(46.7)     | 6(13.3)      | 8(17.8)     | 45(10.1)    |
| St.14               | 1(4.8)      | 0(0.0)      | 4(19.0)      | 2(9.5)       | 14(66.7)    | 21(4.7)     |
| St.15A/15F          | 0(0.0)      | 0(0.0)      | 2(100.0)     | 0(0.0)       | 0(0.0)      | 2(0.4)      |
| St.18C/18B/18A/18F  | 0(0.0)      | 0(0.0)      | 1(100.0)     | 0(0.0)       | 0(0.0)      | 1(0.2)      |
| St.19A              | 0(0.0)      | 0(0.0)      | 2(16.7)      | 0(0.0)       | 10(83.3)    | 12(2.7)     |
| St.19F              | 0(0.0)      | 0(0.0)      | 1(100.0)     | 0(0.0)       | 0(0.0)      | 1(0.2)      |
| St.23F              | 1(5.3)      | 0(0.0)      | 18(94.7)     | 0(0.0)       | 0(0.0)      | 19(4.3)     |
| St.3                | 1(5.9)      | 0(0.0)      | 8(47.1)      | 3(17.6)      | 5(29.4)     | 17(3.8)     |
| St.33F/33A/37       | 0(0.0)      | 0(0.0)      | 3(50.0)      | 1(16.7)      | 2(33.3)     | 6(1.3)      |
| St.4                | 0(0.0)      | 0(0.0)      | 1(50.0)      | 1(50.0)      | 0(0.0)      | 2(0.4)      |
| St.5                | 8(61.5)     | 1(7.7)      | 4(30.8)      | 0(0.0)       | 0(0.0)      | 13(2.9)     |
| St.6A/6B            | 0(0.0)      | 0(0.0)      | 0(0.0)       | 2(100.0)     | 0(0.0)      | 2(0.4)      |
| NT                  | 0(0.0)      | 9(14.8)     | 12(19.7)     | 25(41.0)     | 15(24.5)    | 61(13.7)    |

**Comparison of PCR and culture results outcome for equality**

The hypothesis was therefore developed as  $H_0$ : There is no statistically significant difference between the culture test results and that of the PCR test results,  $H_1$ : There is a statistically significant difference between the culture test results and that of the PCR test results.

A total of 2446 patients were examined using the two methods, pathogens (11.1%) were both isolated in the CSF culture and detected in the RT-PCR method. Twenty-four percent were detected by RT-PCR method but not detected by the CSF culture method. There was 0.2% of the samples in which the CSF culture was positive although the RT-PCR was negative (Table 6).

**Table 6.** Comparison of the CSF culture and the RT-PCR results.

| Methods     |          |            | RT-PCR test |          | PPV  |
|-------------|----------|------------|-------------|----------|------|
|             |          |            | Positive    | Negative |      |
| CSF culture | Positive | count      | 272         | 5        | 277  |
|             |          | % of N     | 11.1        | 0.2      | 65.6 |
|             | Negative | count      | 570         | 1599     | 842  |
|             |          | % of N     | 23.3        | 65.3     | 34.4 |
| Total       |          | count      | 2169        | 277      | 2446 |
|             |          | % of total | 88.7        | 11.3     | 100  |

Table 7 represents the sensitivity and specificity of the CSF culture test method. In order to assess the performance of CSF culture, results were compared with RT-PCR as a gold standard. Accordingly, the diagnostic accuracy of CSF culture including sensitivity (31.8%), specificity (98.2%), positive predictive value (90.5%), and negative predictive value (72.6%). Notably, the positive predictive likelihood ratio of CSF culture was 17.7 while the negative likelihood ratio was 0.69.

**Table 7.** Diagnostic accuracy comparing PCR and culture for CSF samples.

| Diagnostics test              | Cerebro spinal fluid    |                  |
|-------------------------------|-------------------------|------------------|
|                               | RT-PCR positives        | RT-PCR negatives |
| Using RT-PCR as gold standard |                         |                  |
| Culture positives             | 272                     | 5                |
| Culture negatives             | 570                     | 1599             |
| Results                       | 95% confidence interval |                  |
| Sensitivity (%)               | 31.8                    | 22.7-53.4        |
| Specificity (%)               | 98.2                    | 71.2-100.0       |
| Positive predictive value (%) | 90.5                    | 79.5-100.0       |
| Negative predictive value (%) | 72.6                    | 62.3-95.6        |
| Positive likelihood ratio     | 17.7                    | 10.8-29.4        |
| Negative likelihood ratio     | 0.69                    | 0.2-2.7          |

Majority (1823; 74.5%) of results remain symmetric while few (570; 23.3%) are parallel for RT-PCR and culture results. Also, 0.2% of positive culture results tested negative for RT-PCR with a significant p-value ( $p < 0.00$ ). This suggests rejection of  $H_0$  indicative of a statistically significant difference between results chained out by the culture and RT-PCR methods (Table 8).

**Table 8.** Paired sample sign test for difference in test outcomes.

| Method verification   | Measurement                   | Frequencies<br>(n=2446) | Percent<br>(%) |
|---|-------------------------------|-------------------------|----------------|
| Culture results for signed values-PCR results for signed values | Negative differences          | 570                     | 23.3           |
|   | Positive differences          | 5                       | 0.2            |
|   | Ties                          | 1871                    | 76.5           |
|   | Z-value                       | -22.596                 |                |
|   | Asymptomatic. Sig. (2-tailed) | 0                       |                |

## DISCUSSION

The number of confirmed cases were a 34.4% of the total cases suspected over the period. *Neisseria meningitidis* (43.7%) and *Streptococcus pneumonia* (53.0%) were the most predominant pathogens or isolates associated with bacterial meningitis in the three northern regions contributing approximately 95% of confirmed cases. In addition, the pathogens of *Haemophilus influenzae* were 3.3%. Across the trend years, notably were *Streptococcus pneumoniae* with highest positivity rates whereas *Haemophilus influenzae* had the lowest positivity rates. This similarity of pathogens across the trend years could be as a result of the introduction of vaccines to control some strains of the pathogens (*Neisseria meningitidis* W135) [6]. These findings are in well match with CDC, which stated

the most common strains of bacterial meningitis seen in west Africa over the past decades are basically *Streptococcus pneumoniae*, *Neisseria meningitidis*, *Haemophilus influenzae*, and group B *Streptococcus*.

Notwithstanding that, *Neisseria meningitidis* had common strains within the five-year period to be B (0.5%), C (1.1%), W135 (84.0%), and X (11.7%). It was also worth noting that the most common strain of this group is the W135, whereas NmX is identified as the new strain springing up in the meningitis belt of Ghana. Some strains of *Neisseria meningitidis* that could not be grouped by the laboratory due to limited capacity for testing. The results assent to nyarko who uncovered that *Neisseria meningitidis* W135 serogroup which was once rare in Ghana, with only four cases reported in 2004 has now become a common cause of outbreaks in the region [18]. These results also relate and conforms to that of CDC, which stated that the most prevalent and common strains of *Neisseria meningitidis* in Africa is the serogroup A, B, C, W135, X, and Y [7-9]. Contrary to previous findings that uncovered 80%-85% of all of *Neisseria meningitidis* in the African Meningitis belt are due to serogroup A, a different trend is observed in this study where *Neisseria meningitidis* W 135 constituted up to 84.0% of Nm isolates.

*Streptococcus pneumoniae* in recent times have experienced spikes in number of cases and accounted for 53.0% of the total confirmed cases across the five-year period under review with serotype 1 (St.1) accounting for majority (49.6%) cases. Notwithstanding that, some of the strains (Serotypes) of *Streptococcus pneumoniae* (13.7%) were non-typeable. The evidence implies that approximately 70.0% of *Streptococcus pneumoniae* are accounted for by the most common serotypes (St.1, St.12F/12A, St.12F/12A/12B/44/4, and St.14). The emergence of new strains is evident in St.12F/12A/12B, St.14, and St.19. This findings of this study agrees with other researchers who observed a similar increase in incidence of *Streptococcus pneumoniae* in Ghana and other African countries. This suggest that if controls and prevention efforts need to be improved such as mass vaccination campaigns to target predominant strains like *Neisseria meningitidis*, W135, and *Streptococcus pneumoniae* St.1. *Haemophilus influenzae* (3.3%) with 60.7% of the confirmed strains being *Haemophilus influenzae* B. These finding are in congruent with studies which denoted that after the introduction of a conjugated Hib vaccine, the incidence of Hib infections has decreased by 90%. Currently Hib is responsible for 6.7% of all bacterial meningitis cases.

Diagnosis and identification of the isolates causing the bacterial meningitis in the cerebrospinal fluid and the early initiation of the appropriate treatment is the most critical stage in the management of the disease. It is therefore critical to get the best results in order to detect and manage confirmed cases across the country. The RT-PCR remains the gold standard in Ghana but supplemented by CSF culture for immediate management of cases at the peripheral levels [40]. Studies have shown that the latex agglutination test has a very low sensitivity especially in the patients who have received antibiotic treatment before the lumbar puncture and limits the use of his method. Delays in the diagnosis and treatment are avoided through the routine use of CSF culture methods in the patients under the suspicion of bacterial meningitis. The study of the comparison of the culture to the gold standard of RT-PCR method revealed that there is a statistically significant ( $z=-22.596$ ;  $p<0.05$ ) difference between the CSF culture results and RT-PCR methods chained out. In comparison, there was a positive agreement of 11.1% whiles positive disagreement of RT-PCR was 23.3%. Positive disagreement for CSF culture was 0.2% in the overall samples tested. Findings from the results showed that the CSF culture results were not in accordance with those of RT-PCR.

## CONCLUSION

Although some performance characteristics were in good correlation with RT-PCR results such us specificity (98.2%) and positive predictive value (90.5%). Meanwhile, the sensitivity (31.8%) and negative predictive value (72.6%) of



CSF culture were low, which means that majority of the CSF culture results were false negative. The findings are favorably similar with the results of other studies who found specificity of CSF culture not to reflect the true percentage, because the presence of fastidious bacteria, delay in CSF culturing, or consumption of antibiotics before lumbar puncture, could result in altering the outcomes. These outcomes can be attributed to the procedures undertaken before lumbar puncture is performed. Literature states that RT-PCR is more accurate and reliable, especially among patients who have used antibiotics before lumbar puncture. Although some performance characteristics were in good standing by culture results such as the level of agreements of CSF culture with RT-PCR results, the difference was clearly ascertained by the signed rank test.

## DECLARATIONS

### Funding

The research was supported entirely by the authors, there were no funds from any external or internal sources. The study resources were gathered from the research team.

### Competing interests

The authors have no relevant financial or non-financial interests to disclose.

### Ethics approval

The study sought permission and approval from the Ghana health service to assess data from the tamale public health and reference laboratory for bacterial meningitis from 2015 to 2019 respectively. Approval was given from the committee on human research, publication and ethics with ethics number, CHRPE/AP/469/20.

## REFERENCES

1. Mazamay S, et al. An overview of bacterial meningitis epidemics in africa from 1928 to 2018 with a focus on epidemics outside the belt. *BMC Infect Dis.* 2021;21:1-13.
2. Collard JM, et al. Epidemiological changes in meningococcal meningitis in niger from 2008 to 2011 and the impact of vaccination. *BMC Infect Dis.* 2013;13:576.
3. Baspinar EO, et al. Comparison of culture and PCR methods in the diagnosis of bacterial meningitis. *Brazilian J Microbiol.* 2017;48:232-236.
4. Minasyan H. Sepsis: Mechanisms of bacterial injury to the patient. *Scand J Trauma Resusc Emerg Med.* 2019;27:1-22.
5. Karvouniaris M, et al. Current perspectives on the diagnosis and management of healthcare-associated ventriculitis and meningitis. *Infect Drug Resist.* 2022;15:697-721.
6. Maukonen J, et al. Methodologies for the characterization of microbes in industrial environments: A review. *J Ind Microbiol Biotechnol.* 2003;30:327-356.

7. Amidu N, et al. Diagnosis of bacterial meningitis in ghana: Polymerase chain reaction versus latex agglutination methods. PLoS One. 2019;14:e0210812.
8. Sidikou F, et al. Polymerase chain reaction assay and bacterial meningitis surveillance in remote areas, niger. Emerg Infect Dis. 2003;9:1486-1488.
9. Châtelet IP, et al. Bacterial meningitis in burkina faso: Surveillance using field based polymerase chain reaction testing. Clin Infect Dis. 2005;40:17-25.
10. Scarborough M, et al. Corticosteroids for bacterial meningitis in adults in sub-saharan africa. N Engl J Med. 2007;357:2441-2450.