

Research and Reviews: Journal of Zoological Sciences

Biofuel generation from Microalgae

Sri Avinash Kandula*

Pydah college of Engineering, JNTU, Kakinada, Andhra Pradesh, India

Review Article

Received: 10/07/2016

Revised: 21/07/2016

Accepted: 26/07/2016

*For Correspondence

Sri Avinash Kandula, Pydah college of Engineering,
JNTU, Kakinada, Andhra Pradesh, India. Tel:
8332946664

E-mail: sriavinash.kandula@gmail.com

Keywords: Atlantic salmon, Pacific salmon,
Anadromous, Alevins, Chinook, Spawning, Inter
breeding, Catching capacity

ABSTRACT

Exhausting stores and soaring costs of petroleum and oil have overwhelming impact on the economy of numerous created and creating countries and constrained analysts, government and elected organizations to select advancement of interchange energizes. As of late, there is restored enthusiasm for the field of algal biofuel creation inferable from its capacity to develop in non-agricultural able waste area and metropolitan wastewater/farming spill over water. Earlier decade was credited in growing new open society/reactor outline for improved microalga biomass creation, proficient gathering and pre-treatment frameworks for business algal biofuel generation.

INTRODUCTION

Draining fossil fuel saves, an unnatural weather change and expanded worldwide interest for vitality pushed the world to a phase where finding another renewable green option fuel is unavoidable. In no time the United States and China are the biggest overall oil shoppers (>10 million barrel for each day) trailed by Japan (4.7) and India (3) (US Energy Information Administration). It is assessed that by the year 2020, there will be 60 fold increments in the interest for oil around the world. The fossil oil disclosure has declined relentlessly (from 55 billion barrel for every year to 10 billion barrel) following 1960's and extractions from whimsical common sources (profound wells, oil shale and tar sands) are troublesome and significantly more costly. The critical advancement has been made in enhancing innovations for natural transformation/biofuel generation however capital expenses for biofuel offices are generally high. Albeit business force is building, however this developing industry is as yet confronting numerous difficulties. In any case, issues like change in comprehension of the key standards of biofuel proficiency, its scale up innovation improvement and how it will change the vehicles area must be tended to.

Culturing systems of green growth

Microalgae are phototrophic living beings that can build their biomass utilizing light, carbon dioxide, water, and inorganic salts [1-8]. The ideal development temperature fluctuates from 15-30 °C and unsettling is required to avoid algal biomass settlement to the base. Breath amid the night hours causes some a player in the biomass misfortune delivered amid day time. The microalgae for the most part develop on appended surfaces yet it keep the gas and light penetrance to lower level when it exceed subsequently bringing about the passing of fundamental layer, in this way for large scale manufacturing more often than not suspension societies are favored [9-15]. The anticipated yield of algal biomass from pilot scale to fermenter scale is for the most part wrongly computed on the grounds that in high thickness society, basic green growth can't catch all approaching light to change over it into biomass, thus the particular development rate drops in contrast with low

thickness societies [16-20]. The particular development rate of exponentially developing societies can be accomplished in heterotrophic societies under upgraded states of temperature, light force, blending and CO₂ supply. Utilizing such frameworks the photosynthetic efficiencies of ~ 7% might be accomplished however it results in expanded bioreactor upkeep costs consequently making the procedure financially unviable. Green growth is by and large developed in variable open-society frameworks or controlled shut society frameworks and both having their own particular focal points and disservices. The subtle element of every framework is examined underneath.

Land based open society frameworks

Land based open culture framework operation is restricted to zones where ease water is accessible because of the low profundity and huge surface region and water misfortune through vanishing can turn into a noteworthy issue. Marine waters and wastewaters can serve as great matches for this framework, as natural and maintainability issues would forestall expansive open lake development utilizing consumable water [21-30]. There is as of now some experience on vast scale generation utilizing these sorts of frameworks, either in pilot extends mostly subsidized by the legislature, in wastewater treatment plants, where it is utilized as a part of optional or tertiary treatment of sewage, or in business scale algal development for the wellbeing nourishment market.

Shallow unstirred lakes

Shallow and unstirred lakes are the least complex of the area based open society framework for green growth development [31-40]. Their sizes fluctuate from few m² to 2,500,000 m² and use CO₂ as carbon source. Albeit open-society frameworks are anything but difficult to work, less costly and have vast creation limit yet utilizes more vitality and don't permit control of temperature and lighting conditions [41-50]. It is all the more effortlessly inclined to attack by other green growth and defilement from microscopic organisms. Moderate dissemination of supplements, dead and living green growth sedimentation, buoyancy and restricted use of accessible daylight are some different issues that add to the hopelessness of the open-society frameworks.

Circular/raceway lakes

The race-away lakes have attempted to minimize the constraints by utilizing mechanical fomenters to give air circulation. In mechanical fomenters, arm move in a round movement, and an oar wheel cause the flow of water through slender lake. The gas air pockets can be blown and some portion of this gas is utilized to as carbon source and rest give medium tumult [51-60]. A predetermined number of animal varieties can be kept up in an open framework, and thus the locally-happening strain is ideal in an open framework. The open air business creation of microalgae was accomplished in *Arthrospira*, *Chlorella* and *Dunaliella* genera species simply because they demonstrate high development in particular medium (fundamental or profoundly saline) furthermore indicate diminished pollution issues. Such frameworks are less vitality serious, simple to work, shoddy and stronger than shut frameworks [61-70].

Closed frameworks

The motivation behind the mass algal society and nearby climate conditions may settle on the decision of framework self-evident. Be that as it may, the primary correlation between the two frameworks is basically cost and profitability [71-82]. Prior shut frameworks were made by covering the lake with a nursery. While this brought about littler frameworks however numerous issues connected with lake frameworks were handled. It permits more species to be developed and broaden their developing season freely. Open lakes require bigger and more development regions to accomplish the same efficiency. The low blending rate of open lakes intensifies the self-shading impact because of cell fixation and the physical structure of open lakes avert appropriate air circulation, creating a low medium CO₂ halfway weight, therefore constraining the profitability rate per unit of territory and volume [83-90].

Photobioreactors

These primarily include photoautotrophic creation utilizing characteristic or artificial lighting, albeit ordinary blended bioreactor can be utilized to culture some microalgae species heterotrophic ally at high densities oblivious. Photobioreactor then again, are frameworks that are adaptable and can be advanced as per the natural and physiological attributes of the species being developed. In this way, minimizing the tainting and offering better control over society conditions [91-95].

'Major Bag' frameworks

Most likely the longest utilized shut society frameworks for mass society of microalgae are the 'enormous pack' frameworks by and large utilized as a part of aquaculture butcheries to sustain larval fish, scavengers, mollusks or rotifers. Albeit broadly utilized these frameworks are infamous for the precariousness of the way of life. This unsteadiness likely happens in light of the fact that blending in these sacks is uneven, prompting the development of the cells in unmixed ranges, which thusly prompts the cell passing, particularly if the way of life is not axenic (microbes free). To accomplish sensibly dependable societies, it is key to keep up axenic conditions, an element that is not vital for the tubular photobioreactors [95-100].

Offshore culture of macroalgae

Macroalgae are long multi-cell green growth (measure in inches/feet) regularly developed in open lakes and seas (as kelp e.g. goliath kelp plant). Macroalgae can be developed in seaward frameworks (i.e. green growth society in the vast sea). Macroalgae hold guarantee as a crude material for fuel since they deliver more biomass per unit region every year. The primary species applicable to biofuel generation are chlorophyta (family Ulva and Caulerpa), red green growth (Gigartinales, Halymeniales and Palmariales) and chestnut green growth (request of Fucales, Laminariales and Tilopteridales). They can be developed on sewage, city waste water or agribusiness or homestead spill over water. In Florida researcher made Algal Turf Scrubber (ATS) in shallow waterways having nylon netting on which filamentous green growth can frame provinces. Contemplates on Algal Turf Scrubber (ATS) has uncovered that the green growth can catch around 60-90% of N(nitrogen) and 70-100% of K(potassium) from fertilizer effluents overflow water accordingly lessening eutrophication of water bodies. On reaping, the macroalgae can be utilized as natural manure.

Collecting and preparing of green growth

Subsequent to refined in open or shut frameworks, the algal biomass should be collected for further preparing. In any case, gathering microalgae cells is entirely testing. The microalgae can't be effortlessly collected as naturally visible plants, and in this way the subsequent oil extraction is more muddled. In addition, algal societies are exceptionally weaken, more often than not around 1% for autotrophic development up to 10% for heterotrophic development, along these lines making dewatering an important stride preceding biomass use. In green growth reaping, dewatering is most capital and vitality concentrated (-30% of aggregate cost) step. Collecting from bioreactor is less costly when contrasted with open lake framework in light of the fact that the biomass efficiency can normal 13 times more than the open lake. In open culture the biomass yield is around 0.5-1.0 gL⁻¹, while in shut framework it stretch around 5-10 gL⁻¹. Gravity settlement, filtration and centrifugation are the usually utilized collecting technique. Now and again flocculation step or flocculation-flotation is added to help the collecting. The decision of gathering procedure is represented by microalgal species utilized and last item.

Pretreatment technique

Microalgae pretreatment is essential for partition of its diverse helpful segments, which are further handled to various sorts of biofuel. Sugar of algal biomass is changed over either to ethanol or biogas utilizing aging. The biomass is prepared in three successive strides hydrolysis, fermentation and vitality era. Out of these strides hydrolysis is regularly seen as a rate restricting stride. The inflexible cell divider and film make them impervious to biodegradation or show slower biodegradation amid maturation. For algal oil reaping expellers/squeezes, dissolvable extraction and supercritical CO₂ are customarily utilized. For green growth having high oil content expeller/pressers are utilized which mechanically crack the algal cell (70-75% recuperation).

Artificial light can be given by any customary light source, for example, tungsten or fluorescent globules. Low warmth era, the specificity of the wavelength of transmitted light, low power utilization and, permitting the confinement of light to photosynthetic dynamic radiation, the influence of various wavelengths and intensities on these microorganisms has prompted the utilization of LEDs. A late study demonstrated that diverse wavelengths may have a significant influence on biomass and lipid profitability, and also on the lipid profile. A strain of *Nannochloropsis* demonstrated a higher development rate, lipid profitability and distinctive lipid profile under blue light (470 nm) when contrasted and development under red (680 nm) or green (550 nm).

REFERENCES

1. Pérez L. Biofuels from Microalgae, A Promising Alternative. *Pharm Anal Chem Open Access* 2016;2:e103
2. dos Santos RG, et al. Thermochemical Liquefaction of Swine Manure as Feedstock for the Production of a Potential Biofuel. *Innov Ener Res* 2015;4:125.
3. Hong JW, et al. Mass Cultivation from a Korean Raceway Pond System of Indigenous Microalgae as Potential Biofuel Feedstock. *Oil Gas Res* 2016;2:108.
4. Silva LMS. Microbial Production of Short Chain Alkanes: A Future Biofuel. *Adv Genet Eng* 2015;4:136.
5. Wysocka J, et al. The use of Alcohols and their Compounds as Biofuel and Gasoline Blends. *J Civil Environ Eng* 2015;5:187.
6. Slaughter G and Kulkarni T. Enzymatic Glucose Biofuel Cell and its Application. *J Biochip Tissue Chip* 2015;5:110.
7. Stephen S, et al. Tracking Interfacial Adsorption/Desorption Phenomena in Polypropylene/Biofuel Media using Trace Cr³⁺/Cr⁶⁺ and As³⁺/As⁵⁺-A Study by Liquid Chromatography-plasma Mass Spectrometry. *J Pet Environ Biotechnol* 2015;6:239.
8. Saini KJ. Biofuel: A Ray of Hope for Sustainable Future. *J Pet Environ Biotechnol* 2015;6:229.
9. Saldivar RP et al. Algae Biofuels Production Processes, Carbon Dioxide Fixation and Biorefinery Concept. *J Pet Environ Biotechnol* 2014;5:185.
10. Swain KC. Biofuel Production in India: Potential, Prospectus and Technology. *J Fundam Renewable Energy Appl* 2014;4:129.
11. Sticklen MB, et al. Towards Cellulosic Biofuels Evolution: Using the Petro-Industry Model. *Adv Crop Sci Tech* 2014;2:131.
12. Hasan R, et al. Bioremediation of Swine Wastewater and Biofuel Potential by using *Chlorella vulgaris*, *Chlamydomonas reinhardtii*, and *Chlamydomonas debaryana*. *J Pet Environ Biotechnol* 2014;5:175.
13. Kurosawa K, et al. Triacylglycerol Production from Corn Stover Using a Xylose-Fermenting *Rhodococcus opacus* Strain for Lignocellulosic Biofuels. *J Microb Biochem Technol* 2014;6:254-259.
14. Gabriel Morales et al. Advanced Biofuels from Lignocellulosic Biomass. *J Adv Chem Eng* 2014;4:e101.
15. Dhaman Y and Roy P. Challenges and Generations of Biofuels: Will Algae Fuel the World? *Ferment Technol* 2013;2:119.
16. Yang ST and Liu X. Metabolic Process Engineering for Biochemicals and Biofuels Production. *J Microb Biochem Technol* 2014;6:e116.
17. Bhatt SM. Developments in Cellulase Activity Improvements Intended Towards Biofuel Production. *J Bacteriol Parasitol* 2013;4:e120.
18. Sekhon KK and Rahman PKSM. Synthetic Biology: A Promising Technology for Biofuel Production. *J Phylogenetics Evol Biol* 2013;4:e121.
19. Borole AP. Biofuel Cells and Bioelectrochemical Systems. *J Microb Biochem Technol* 2013;S6:e001.
20. Kapazoglou A, et al. Biofuels Get in the Fast Lane: Developments in Plant Feedstock Production and Processing. *Adv Crop Sci Tech* 2013;1:117.

21. Ibrahim E, et al. Molecular Cloning and Expression of Cellulase and Polygalacturonase Genes in *E. coli* as a Promising Application for Biofuel Production. *J Phylogenetics Evol Biol* 2013;4:147
22. Jessup RW. Seeded-Yet-Sterile' Perennial Biofuel Feedstocks. *Adv Crop Sci Tech* 2013;1:e102.
23. Sticklen MB. Co-Production of High-Value Recombinant Biobased Matter in Bioenergy Crops for Expediting the Cellulosic Biofuels Agenda. *Adv Crop Sci Tech* 2013;1:e101.
24. Ragauskas AJ. Do-Able Biofuels. *J Pet Environ Biotechnol* 2012;3:e105.
25. Nag A. Open Access Research in Biological Networks Will Facilitate Advances in Network-Based Paradigms for Biomedicine and Biofuel Production. *J Phys Chem Biophys* 2012;2:e106.
26. Ball MRB, et al. The "Some Sense" of Biofuels. *J Pet Environ Biotechnol* 2012;3:e107.
27. Sameera V, et al. Current Strategies Involved in Biofuel Production from Plants and Algae. *J Microbial Biochem Technol* 2011;R1:002.
28. Zhang B and Shahbazi A. Recent Developments in Pretreatment Technologies for Production of Lignocellulosic Biofuels. *J Phylogenetics Evol Biol* 2011;2:108.
29. Iyovo GD, et al. Sustainable Bioenergy Bioprocessing: Biomethane Production, Digestate as Biofertilizer and as Supplemental Feed in Algae Cultivation to Promote Algae Biofuel Commercialization. *J Microbial Biochem Technol* 2010;2:100-106.
30. Iyovo GD, et al. Poultry Manure Digestate Enhancement of *Chlorella Vulgaris* Biomass Under Mixotrophic Condition for Biofuel Production. *J Microbial Biochem Technol* 2010;2:051-057.
31. Christaki E, et al. Phycobiliproteins: A New Perspective in Natural Pigments Derived from Microalgae. *J Oceanogr Mar Res* 2016;4:e114.
32. Mehta P, et al. Growth and Tolerability of Healthy Term Infants Fed a New Formula Supplemented with DHA from *Schizochytrium* sp Microalgae. *J Vasc Med Surg* 2016;4: 267.
33. Pérez L. Biofuels from Microalgae, A Promising Alternative. *Pharm Anal Chem Open Access* 2016;2:e103.
34. Benmoussa M. Algomics for the Development of a Sustainable Microalgae Biorefinery. *Single Cell Biol* 2016;5:132.
35. Sarpal AS, et al. Investigation of Biodiesel Potential of Biomasses of Microalgae *Chlorella*, *Spirulina* and *Tetraselmis* by NMR and GC-MS Techniques. *J Biotechnol Biomater* 2016;6:220.
36. Hong JW, et al. Mass Cultivation from a Korean Raceway Pond System of Indigenous Microalgae as Potential Biofuel Feedstock. *Oil Gas Res* 2016;2:108.
37. Taucher J, et al. Cell Disruption and Pressurized Liquid Extraction of Carotenoids from Microalgae. *J Thermodyn Catal* 2016;7:158.
38. Iturriaga R. Photo Adaptation Response of Microalgae to Environmental Changes. *Oceanography* 2015;3:e113.
39. Ramirez-Merida LG, et al. Microalgae as Nanofactory for Production of Antimicrobial Molecules. *J Nanomed Nanotechnol* 2015;S6:004.
40. Suantika G, et al. Performance of Zero Water Discharge (ZWD) System with Nitrifying Bacteria and Microalgae *Chaetoceros calcitrans* Components in Super Intensive White Shrimp (*Litopenaeus vannamei*) Culture. *J Aquac Res Development* 2015;6:359.
41. Hattab MA and Ghaly A. Microalgae Oil Extraction Pretreatment Methods: Critical Review and Comparative Analysis. *J Fundam Renewable Energy Appl* 2015;5:172.

42. Sharma M, et al. Microalgae as Future Fuel: Real Opportunities and Challenges. *J Thermodyn Catal* 2015;6:139.
43. Al hattab M, et al. Microalgae Harvesting Methods for Industrial Production of Biodiesel: Critical Review and Comparative Analysis. *J Fundam Renewable Energy Appl* 2015;5:154.
44. Sibi G. Biosorption of Arsenic by Living and Dried Biomass of Fresh Water Microalgae - Potentials and Equilibrium Studies. *J Bioremed Biodeg* 2014;5:249.
45. Sanmukh S, et al. Bioactive Compounds Derived from Microalgae Showing Antimicrobial Activities. *J Aquac Res Development* 2014;5:224
46. Steudel B. Microalgae in Ecology: Ecosystem Functioning Experiments. *Oceanography* 2014;2:122.
47. Christaki E. Microalgae as a Potential New Generation of Material for Various Innovative Products. *Oceanography* 2014;1:e106.
48. Raja R, et al. Biomass from Microalgae: An Overview. *Oceanography* 2014;2:118.
49. Jakob G, et al. Surveying a Diverse Pool of Microalgae as a Bioresource for Future Biotechnological Applications. *J Phylogenetics Evol Biol* 2013;4:153.
50. Botana LM, et al. Warm Seawater Microalgae: Growth and Toxic Profile of *Ostreopsis* Spp. From European Coasts. *Oceanography* 2013;1:104.
51. Fargione J, et al. Land clearing and the biofuel carbon debt. *Science*. 2008;29:1235-8.
52. Hu Q, et al. Microalgal triacylglycerols as feedstocks for biofuel production: perspectives and advances. *The Plant Journal*. 2008;1;54:621-39.
53. Crutzen PJ, et al. N₂O release from agro-biofuel production negates global warming reduction by replacing fossil fuels. *Atmospheric chemistry and physics*. 2008;29;8:389-95.
54. Kumar P, et al. Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial & engineering chemistry research*. 2009;20;48:3713-29.
55. Rabaey K, et al. Biofuel cells select for microbial consortia that self-mediate electron transfer. *Applied and environmental microbiology*. 2004;1;70:5373-82.
56. Calabrese Barton S et al. Enzymatic biofuel cells for implantable and microscale devices. *Chemical reviews*. 2004;13;104:4867-86.
57. Chen F and Dixon RA. Lignin modification improves fermentable sugar yields for biofuel production. *Nature biotechnology*. 2007;1;25:759-61.
58. Sims RE, et al. An overview of second generation biofuel technologies. *Bioresource technology*. 2010;31;101:1570-80.
59. Heller A. Miniature biofuel cells. *Physical Chemistry Chemical Physics*. 2004;6(2):209-16.
60. Bullen RA, et al. Biofuel cells and their development. *Biosensors and Bioelectronics*. 2006;15;41-45.
61. Pittman JK, et al. The potential of sustainable algal biofuel production using wastewater resources. *Bioresource technology*. 2011;102:17-25.
62. Peterson AA, et al. Thermochemical biofuel production in hydrothermal media: a review of sub-and supercritical water technologies. *Energy & Environmental Science*. 2008;1:32-65.
63. Lal RA. World crop residues production and implications of its use as a biofuel. *Environment International*. 2005;31:575-84.
64. Demirbas A. Biofuels sources, biofuel policy, biofuel economy and global biofuel projections. *Energy conversion and management*. 2008;49:2106-16.

65. Chen T et al. A miniature biofuel cell. *Journal of the American Chemical Society*. 2001;123:8630-1.
66. Heaton EA, et al. Meeting US biofuel goals with less land: the potential of *Miscanthus*. *Global Change Biology*. 2008;14:2000-14.
67. Rabaey K, et al. Microbial phenazine production enhances electron transfer in biofuel cells. *Environmental science & technology*. 2005;39:3401-8.
68. Foidl N, et al. as a source for the production of biofuel in Nicaragua. *Bioresource Technology*. 1996;58:77-82.
69. Dürre P. Biobutanol: an attractive biofuel. *Biotechnology journal*. 2007;2:1525-34.
70. Radakovits R, et al. Genetic engineering of algae for enhanced biofuel production. *Eukaryotic cell*. 2010;9:486-501.
71. Cherubini F, et al. Energy-and greenhouse gas-based LCA of biofuel and bioenergy systems: Key issues, ranges and recommendations. *Resources, conservation and recycling*. 2009;53:434-47.
72. Katz E, et al. A non-compartmentalized glucose|O₂ biofuel cell by bioengineered electrode surfaces. *Journal of Electroanalytical Chemistry*. 1999;479:64-8.
73. Amin S. Review on biofuel oil and gas production processes from microalgae. *Energy conversion and management*. 2009;50:1834-40.
74. Park JB, et al. Wastewater treatment high rate algal ponds for biofuel production. *Bioresource technology*. 2011;102:35-42.
75. Davis F and Higson SP. Biofuel cells-recent advances and applications. *Biosensors and Bioelectronics*. 2007;22:1224-35.
76. Kim J, et al. Challenges in biocatalysis for enzyme-based biofuel cells. *Biotechnology advances*. 2006;24:296-308.
77. Mano N, et al. Characteristics of a miniature compartment-less glucose-O₂ biofuel cell and its operation in a living plant. *Journal of the American Chemical Society*. 2003;125:6588-94.
78. Minter SD, et al. Enzyme-based biofuel cells. *Current opinion in biotechnology*. 2007;18:228-34.
79. Sticklen MB. Plant genetic engineering for biofuel production: towards affordable cellulosic ethanol. *Nature Reviews Genetics*. 2008;9:433-43.
80. Gibbs HK, et al. Carbon payback times for crop-based biofuel expansion in the tropics: the effects of changing yield and technology. *Environmental Research Letters*. 2008;3:034001.
81. Chisti Y. Biodiesel from microalgae. *Biotechnology advances*. 2007;25:294-306.
82. Spolaore P, et al. Commercial applications of microalgae. *Journal of bioscience and bioengineering*. 2006;101:87-96.
83. Mata TM, et al. Microalgae for biodiesel production and other applications: a review. *Renewable and sustainable energy reviews*. 2010;14:217-32.
84. Hillebrand H, et al. Biovolume calculation for pelagic and benthic microalgae. *Journal of phycology*. 1999;35:403-24.
85. Brennan L and Owende P. Biofuels from microalgae-a review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and sustainable energy reviews*. 2010;14:557-77.
86. Rodolfi L, et al. Microalgae for oil: Strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. *Biotechnology and bioengineering*. 2009;102:100-12.

87. Chisti Y. Biodiesel from microalgae beats bioethanol. *Trends in biotechnology*. 2008;26:126-31.
88. Schenk PM, et al. Second generation biofuels: high-efficiency microalgae for biodiesel production. *Bioenergy research*. 2008;1:20-43.
89. Brown MR, et al. Nutritional properties of microalgae for mariculture. *Aquaculture*. 1997;151:315-31.
90. Pulz O and Gross W. Valuable products from biotechnology of microalgae. *Applied microbiology and biotechnology*. 2004;65:635-48.
91. Gouveia L and Oliveira AC. Microalgae as a raw material for biofuels production. *Journal of industrial microbiology & biotechnology*. 2009;36:269-74.
92. Borowitzka MA. Commercial production of microalgae: ponds, tanks, tubes and fermenters. *Journal of biotechnology*. 1999;70:313-21.
93. Becker EW. Micro-algae as a source of protein. *Biotechnology advances*. 2007;25:207-10.
94. Lee JY, et al. Comparison of several methods for effective lipid extraction from microalgae. *Bioresource technology*. 2010;101:S75-7.
95. Lardon L, et al. Life-cycle assessment of biodiesel production from microalgae. *Environmental science & technology*. 2009;43:6475-81.
96. Benemann JR. CO₂ mitigation with microalgae systems. *Energy conversion and management*. 1997;38:S475-9.
97. Sialve B, et al. Anaerobic digestion of microalgae as a necessary step to make microalgal biodiesel sustainable. *Biotechnology advances*. 2009;27:409-16.
98. Chen CY, et al. Cultivation, photobioreactor design and harvesting of microalgae for biodiesel production: a critical review. *Bioresource technology*. 2011;102:71-81.
99. Lorenz RT and Cysewski GR. Commercial potential for *Haematococcus* microalgae as a natural source of astaxanthin. *Trends in biotechnology*. 2000;18:160-7.
100. Li Y, et al. Biofuels from microalgae. *Biotechnology progress*. 2008;24:815-20.