

Prevalence And Associated Risk Factors Of Bovine Trypanosomiasis In And Around Itang District Of Gambella Region, Western Ethiopia

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ABSTRACT

A protozoan disease of animals causing considerable impact on cattle production and trypanosomiasis is the most known by causing decrease in productivity in the tropical and subtropical countries of Africa. A prevalence of bovine trypanosomiasis was conducted from January 2019 to May 2019 using cross-sectional study design. Random blood sample (n=203) collection from ear-vein of cattle was done. Direct microscopic examination, buffy coat and thin smears were done following sample collection. The total prevalence of bovine trypanosomiasis in Itang district was 17.7% (36/203). Among each village, the prevalence was recorded 39.2% (20/51), 16% (8/49), 14% (7/50) and 1.9% (1/53) in Pilual, Biljakoak, Makod and Lare by decreasing order respectively. *T. congolense* and *T. Brucei* were the ultimate and lowest documented trypanosome species with 44.4% and 16.7% prevalence respectively. The prevalence of bovine trypanosomiasis infection between the male and female was statistically non-significant ($p>0.05$), but there was a significant difference ($p<0.05$) in the prevalence of trypanosomiasis amongst different age groups of bovine with 3.8% (4/105) and 32.6% (32/98) in adult and old respectively. This study also indicated that animal with poor body conditions were significantly ($p<0.05$) affected with trypanosomiasis. Bovine trypanosomiasis is the main constraints that reduce

milk production, decreasing body condition and, even death of cattle. Hence there should be an integrated tsetse and trypanosomiasis control action to diminish the direct and possible loss of livestock in study areas.

INTRODUCTION

The Ethiopian country is known by huge livestock population. The cattle production is heading by double digit and according to reported data “livestock population is assumed to be 59 million cattle, 35 million sheep, 31 million goats, 2.3 million camels, more than 9 million equides (donkey, horse and mule) and 38 million Poultry”^[1] and heading the other African countries. Cattle are important animals for poor communities to teaching their students, as well as other cash income farming system ^[2] and offer food item, products and hooves for cash income. Despite there are many constraints that will affect the target result of these purposes. Tsetse transmitted animal trypanomiasis encourages animal product and production victims in the country. For instance dozens of Cattle, a huge number of sheep and goats, many equides and camels are at risk of contracting tsetse borne trypanosomiasis annually^[3]. Therefore, this is an indication of the importance of *trypanosomosis* in cattle production and productivity^[4].

Agricultural development and renovation of animal’s diseases are among move in parallel manor. In some areas of the country (river basins of Abay, Omo, Ghibe and Baro ^[5]), limiting livestock productivity and agricultural development is due to high trypanosomiasis burden distributed in the areas. Trypanosomiasis is caused by a unicellular protozoan parasitic disease found in circulatory and many tissues of vertebrates ^[6] and causing constraints to agricultural developments. According to reports from Ethiopia, cattle, sheep, goats, camels and horses are infected by *T. Congolese*, *T. vivax*, *T. Brucei*, *T. evansi* and *T. equiperdium* species respectively ^[7]. Vector distribution, its virulence and species of animals are influencing factors for the epidemiology of trypanosomiasis ^[8]. The tsetse flies infest animal’s mechanically ^[4] by sucking blood containing infectious trypanosomiasis and fly over non-infected animals to transfer paracitemia. Tabanus species vectors are the primary transmitter of *T.vivax* mechanically ^[9].

Trypanosome development and its life cycle are complete in both tsetse fly vectors and the mammalian host. Different forms of transformation are found in trypanosomes during their development ^[10]. Most tsetse transmission is cyclical and begins when blood from trypanosomes infected animals are ingested by the fly ^[11]. The surface coat glycoprotein of trypanosome loosed and to become infective they must get their surface coat again. The metacyclic form is the process by which infective stages of trypanosomes is released from salivary gland tsetse fly ^[9].

The diagnostic methods to be used for identification of trypanosomiasis are different and the easiest is based on direct microscopic examinations that afford the best means of detecting *Trypanosoma* species ^[12]. The other sophisticated method is demonstration of antibody (serological) in conjunction with clinical observation. Tsetse control methods by trap and insecticide treated livestock, which has little direct damage to the environment ^[13] is very effective if applied properly in appropriate circumstances. A few studies were conducted on prevalence of

bovine trypanosome in the Itang district of Ethiopia. Therefore, the objectives of this study were to determine the prevalence of bovine trypanosomiasis in four areas of Itang district and to assess the major risk factors associated with the disease in the study areas.

MATERIALS AND METHODS

Sample collection and sample size

The set-up that includes locations, sex, age, breed (bovine; local breed) and body conditions of the animals were prepared before starting the collection of samples. The animals' number mandatory for this study was calculated to take 384 based on calculation given by [14]. But for the sake of this study, 203 animals were selected based on clinical sign and history of animals complained from the animal's owner came to the veterinary clinics. The ears of the target cattle were prepared thoroughly for blood sample collection by following the procedures for ear vein puncture [15].

Buffy coat technique

The hematocrit tubes were filled with the blood samples collected from the suspected cattle and centrifuged, and then the tubes were cut 1mm below the buffy coat including the top most layers of the RBC's. The contents of the upper layer of the tube were poured on the cover slide, covered with cover slip and examined using microscope to determine the movements of the trypanosomes species under 40x objective lens. Further, the species of the trypanosomes were identified based on their pattern of movement in wet film preparation [16].

Thin blood smear

This study employed a thin blood smear protocol indicated in the Cherenet et al [17]. A drop of blood from study animals were placed on the cover slide and gently spread with another slide by forming an angle of 45° with the first one. The slides were then immersed in the Giemsa stain for 50min after air dried and fixed with methyl alcohol. Giemsa stained slides were washed with distilled water and dried by putting on the rack at upright position. The species of the trypanosomes were identified based on their different morphological characteristics (size, shape, position, location and size) of the kinoplast under microscope with oil immersion.

Data management and statistical analysis

The analysis of the data was done using the SPSS (version 20.0), a software program for social science. The percentages and chi-square (χ^2) outputs were used to measure prevalence and the association between the risk factors, respectively. Ninety five percent (95%) confidence interval and a p-value<0.05 were used for the interpretation of the output.

Ethical Approval

The study was approved by Addis Ababa University, College of Veterinary Medicine and Agriculture Research Ethics Review Committee (Reference number VM/ERC/O110/2017). Animal was handled humanely and blood sample was collected in suitable handling methods.

RESULTS AND DISCUSSION

The result of bovine trypanosomiasis prevalent in study area was 17.7% (36/203). The result obtained from each district was determined and found to be 39.2% (20/51), 14% (7/50), 16.3% (8/49) and 1.9% (1/53) in Pilual, Makod, Biljakoak and Lare areas respectively. *T. congolense* was found dominant with proportion of 7.8% (16/203) followed by *T. vivax* 7.7% (14/203) and *Trypanosoma brucei* 2.8% (6/203) in the areas. The highest 39.2% (20/51) and lowest 1.9% (1/53) prevalent of bovine trypanosomiasis was recorded in Pilual and Lare respectively (table 1). There were significant difference ($p < 0.05$) recorded between prevalence of trypanosomiasis and peasant association.

Table 1: Overall prevalence of bovine trypanomiasis at peasant association level of Itang district.

No	kebele	No- of animal examined	<i>T. congolense</i>	<i>T. vivax</i>	<i>T. Brucei</i>	Prevalence (%)	χ^2	p-Value
1	Pilual	51	9	8	3	39.2	25.7	0
2	Makod	50	3	3	1	14		
3	Biljakok	49	4	3	1	16.3		
4	Lare	53	0	0	1	1.9		
	Total	203	16	14	6	17.7	25.7	0

Prevalence of trypanosomiasis was recorded 32.6% (32/98) in adult and 3.8% (4/105) in young age of cattle respectively (table 2 and fig 2). The association between trypanosomes prevalent and the age group of animals was shown significant difference ($p < 0.05$). The prevalence of bovine trypanosomiasis in female and male was 18.3% (20/109) and 17% (16/94) respectively, with no association with the prevalence ($p > 0.05$) among sex groups.

Table 2: Prevalence of bovine Trypanomiasis based on associated risk factors effects on reproductive physiology.

Factor	Category	Number of examined	Number of positive	Prevalence (%)	χ^2	p-value
Age	Young	105	4	3.80%	28.9	0
	Adult	98	32	32.60%		
Sex	Male	94	16	17.00%	0.61	0.47
	Female	109	20	18.30%		
Body	Poor	18	5	27.78%	9.2	0.01

condition	Medium	88	22	25%		
	Good	97	9	9.20%		
Study sites	Pilual	51	20	39.20%	25.7	0
	Makod	50	7	14%		
	Biljakok	49	8	16%		
	Lare	53	1	1.90%		

Poor body conditioned animals resulted the highest prevalence (27.78% (5/18) (fig 1)) than medium and good body condition and the association was statistically significant difference ($p < 0.05$).

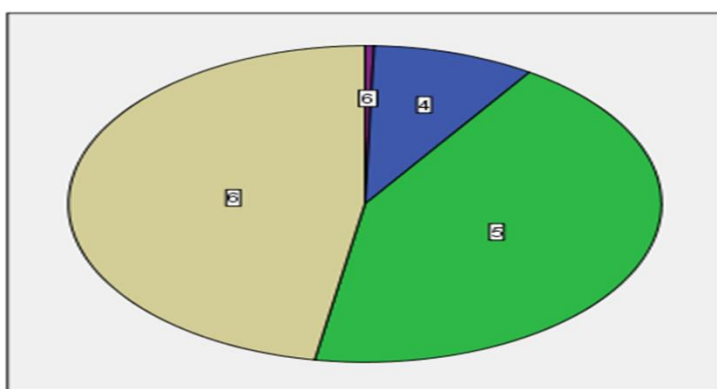


Figure 1: Prevalence of bovine trypanosomiasis according to body condition score(4=good; 5=medium; 6'=poor and 6=very good).

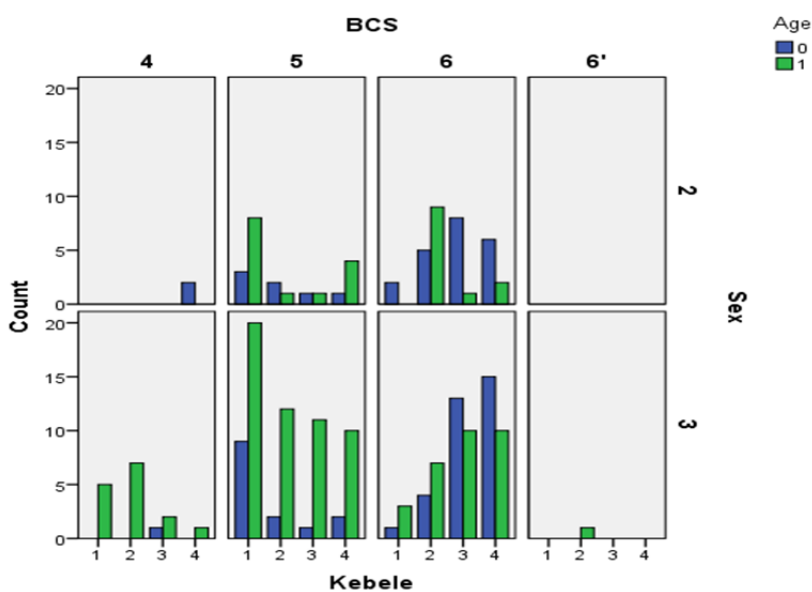


Figure 2: Prevalence of bovine trypanosomiasis according to age group (green- adult and blue color young).

DISCUSSION

The current result revealed that there are over estimated prevalence of bovine trypanosomiasis in the Itang district. From a total of 203 examined cattle, 36 (17.7%) were positive for trypanosome parasite. This result is higher than the result reported by Olani and Bekele ^[18] 7.8% in Ialo-kile district; Fedesa et al. ^[19] 7.1% in Darimu district and Miruk et al. ^[20] and Kitila et al. ^[21] 7.4% in Yayo district Iluababora Zone of Ethiopia in bovine respectively. In addition, the current study is much greater than the value reported by Teka ^[22], Dawit ^[23] and Feyisa et al. ^[24] who reported that the prevalence of 3.7% in Abaya district, 4.9% in Arbaminch and 4.4% in Didessa district of Ethiopia respectively. This difference may be associated with agro-ecological, breed, management system, distribution of vectors and control and prevention strategies implemented in the different studies. In addition the study areas are very near to the grasses and forests very favorable for the survival of tsetse flies.

From 36 cattle tested positive for trypanosomiasis, 44.4% of the predominant species of trypanosome were *T. congolense* which is in agreement with the work of Olani and Bekele ^[18] and Biyazen et al. ^[25] who reported that the major transmitters of *T. congolense* is the cyclical vector groups of *Glossina* species. It might also be attributable to the high number of genetics of *T. congolense* relative to *T. vivax* and *T. brucei*, which is highly susceptible to treatment while the problem of drug resistance are higher in *T. congolense*. The other justification may be the predilection sites of *T. congolense* in blood, while *T. vivax* and *T. Brucei* invade tissue which shows different pathotypes. The current result is by far lower than the work of Aki et al. ^[26] demonstrated *T. congolense* proportional prevalence of 75.86% and *T. vivax* proportional prevalence of 24.14% on cattle *trypanosomosis* in Pawe district, Benishangul Gumuz Regional State, Western Ethiopia. Bayisa et al. ^[27] demonstrated *T. congolense* proportional prevalence of 85% during his research on prevalence of bovine *trypanosomosis* in Asossa district, Benishangul Gumuz Regional State, Western Ethiopia. The highest prevalence reported in these studies may be the higher prevalence of savannah grassland; difference in geography, vector distribution, animal husbandry and management practice.

The highest prevalent of trypanosomiasis in female animals shown in present finding in consistence with Lelisa et al. ^[28], Takile et al. ^[29], Adale and Yasin ^[30] and Mulaw et al. ^[31] who did similar investigation at Mandura District Northwest Ethiopia, Guto Gida District of East Wollega Zone, Wolaita Zone Kindo Koish District and Assosa area respectively. However, this study result is not agree with study done by Teka et al. ^[32] in selected villages of Arbaminch and Gemedra in and around Nekemte areas of East Wollega Zone. The higher prevalence reported in female animals may be by stresses of pregnancy and disturbances encountered in female animals which makes them susceptible for infection due to decreased immunity.

The higher prevalence of bovine trypanosomiasis reported in poor body conditioned animals may be due to blood sucking trypanosomes, which results in progressive emaciation of the infected animals. The poor body conditioned animal may also be associated with other pathogens and nutrition deficiency which might make the animal more susceptible to trypanosomiasis. Nevertheless, non-infected under good condition have well developed immune status that can respond to any foreign protein better than those non infected cattle with poor condition ^[33]. This finding is also in line with that of Bitew et al. ^[34], Teka et al. ^[22] and Feyisa et al. ^[24], Gona et al. ^[35] and Yalew and

fantahun [36] who reported that prevalence of trypanosomiasis was significantly associated with poor body condition in cattle. The finding is also in agreement with the reports of by Dinka and Mulugeta [37], Bekele and Nasir [38] and Mulatu et al. [39] in Ethiopia, who reported high prevalence of trypanosomiasis in poor body conditioned cattle. The current prevalence of trypanosomiasis in young 4(3.8%) and adult 32(32.6%) cattle examined with statistically non-significant difference ($P=0.215$) is in agreement with the study conducted by Dagnachew and Shibeshi [40] in Anger valley of East Wollega Zone and Teka et al. [22] at selected areas of Arbaminich areas. Nevertheless, it's different from study conducted by Alemayehu et al. [41] in Chena Wereda and Bishaw et al. [42] at Wembera district of West Gojam. This may be due to the fact that most of the young animals in the study area were confined to house and they don't have access to grazing lands where the vectors usually prevail.

CONCLUSION

Bovine trypanosomiasis is the most important livestock constraint in the study area. The major species of trypanosome encountered were *T. Congolese* followed by *T.vivax* and *T. Brucei*. The disease is mainly affecting young and poor body conditioned animals. Therefore, an integrated tsetse and trypanosomiasis control action should be strengthened in the area in ordered to minimize direct and potential loss of livestock due to this disease. The laboratory facility should be fulfilled to enable for proper use of drugs with respect to the susceptible species of trypanosome at the study areas and awareness creation in rural communities about the parasite, vector and their impact are forwarded as recommendation for stakeholders and responsible bodies of the study district.

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CONFLICT OF INTEREST

All authors have no potential conflicts of interest.

REFERENCES

1. Federal democratic republic Ethiopia Central Statistical Agency (CSA). Agricultural survey livestock and livestock characteristics. IHSN .2008;08:1-188.
2. Ayana, et al. A cross-sectional study on the prevalence of bovine Trypanosomosis in Amhara region. Northwest Ethiopia. Livestock Research for Rural Development.2012;24(8).
3. Tsetse and Trypanosomiasis prevention and control strategies. Ministry of Agriculture and Rural Development of Government of Ethiopia .MoARD. Ahmaric version.2004.

4. Endalew D, et al. Study on the Prevalence of Trypanosoma Species Causing Bovine Trypanosomosis in South Achefer District. Northern Ethiopia. International Journal of Research Studies in Biosciences.2019;7(9):37-43.
5. Shimelis D, et al. Epidemiology of Tsetse Transmitted Trypanosomosis in Abay (Blue Nile) basin of Northwest Ethiopia. Rev.Elev.Vet.pays.Trop.2005;58(3):151-157.
6. Tesfaye M. Report of Trypanosome infection rate. in G.morsitans and G.tachninoïdes in Didessa valley from July to September.Bedelle.2002.
7. Abebe G.Trypanosomosis in Ethiopia.J.Biol.science.2005;4:75-121.
8. Urquhart, et al. Veterinary parasitology 2nd ed. Black Well publishing.1996;212-217.
9. Aiello E, et al .Tsetse transmitted Trypanosomiasis in Merck veterinary manual. eighth edition .Merck and Co.. Inc.1998.
10. Seifert SHH. Tropical animal health; second edition. Dordrecht Kluwer Academic publisher.1996;78-170.
11. World Organization for Animal Health (2012).Manual of Diagnostic Tests and Vaccines for Terrestrial Animals. OIE. Paris.2.4.18.
12. Wondewosen T, et al. Prevalence study of trypanosomosis and Tsetse density in selected villages of Arbaminch Ethiopia. J.Vet. med.Anim.Heal.2012;4(3):36-41.
13. Vale GA. Development of baits for Tsetse flies (Diptera:Glossinidae)in Zimbabwe .J.Med.Entomol.1993;30:831-842.
14. Thrusfield M. Veterinary Epidemiology third edition. Blackwell publishing England.2005;345-543.
15. <https://veteriankey.com/bovine-clinical-procedures/>
16. Juyal PD, et al. Management of surra due to Trypanosomaevansi in India: an overview.In:Infections Diseases of Domestic Animals and Zoonosis in India. Tandon v and Dhawan BN(Ed's) . proceedings of the National Academy of sciences India section B: Biological science.2005;75:109-120.
17. Cherenet T, et al. A comparative longitudinal study of bovine trypanosomosis in Tsetse free and Tsetse infested zone of the Amhara Region. North west Ethiopia. Veterinary parasitology.2006.
18. Olani A, et al. Epidemiological status and vector identification of bovine trypanosomosis in Lalo-kile district of kellem wollega zone. Western Ethiopia. Journal of Veterinary Medicine and Research.2016;3(2):1045.
19. Fedesa H, et al. Study on spatial distribution of tsetse fly and prevalence of bovine trypanosomosis and other risk factors: case study in Darimu district. Iluababora zone. Western Ethiopia. Journal of pharmacy and Alternative medicine.2015;7.

20. Miruk A, et al. Prevalence of bovine trypanosomosis and trypanocidal drug sensitivity studies on *Trypanosoma congolense* Wolaita and Dawero zones of southern Ethiopia. *Veterinary Parasitology*.2008;152:141-147.
21. Kitila G, et al. Epidemiological investigation of bovine trypanosomosis and its vector apparent densities in Yayo District Illuababorazone. Western Oromia. Ethiopia. *Austin journal of veterinary science and animal husbandry*. 2017;4(1):1031.
22. Teka W, et al. Prevalence study of bovine trypanosomosis and tsetse density in selected villages of Arbaminch. *Eth J of Veterinary Medicine and Animal Health*. 2012;4:36-41.
23. Dawit A, et al. Prevalence of bovine trypanosomosis and its associated risk factors in Abaya district. Borena zone. Ethiopia. *Natural sciences*. 2015;13(10):64-70.
24. Feyisa G, et al. Epidemiological status and vector identification of bovine trypanosomosis in Didessa district of Oromia regional state. Ethiopia. *International journal of Nutrition and food science*. 2015;4(3):373-380.
25. Biyazen H, et al. Trypanosomosis: its risk factors and anemia in cattle population of Dale wabera district of kellem Wollega zone. Western Ethiopia. *Journal of veterinary medicine Hindawi Publishing Corporation*.2014.
26. Asmamaw A. Epidemiology of Bovine Trypanosomosis and Apparent Density of Tsetse and Biting flies in selected Districts of Asossa zone of BenishangulGumuz Regional State. Western Ethiopia. *Biomedicine and Nursing*. 2016;2(3):24-31.
27. Bayisa. K, et al. T. Bovine Trypanosomosis in Asossa District. BenishangulGumuz Regional State. Western Report and Opinion.2017;9(3).
28. Lelisa K, et al. Prevalence of Bovine Trypanosomosis and Apparent Density of Tsetse and Other Biting Flies in Mandura District. Northwest Ethiopia. *J of Veterinary Science and Technology*.2015;6:2-5.
29. Takile D, et al. Prevalence of Bovine Trypanosomosis in Guto Gida District of East Wollega Zone. Oromia Regional State. Ethiopia. *Global Journals Inc*.2014;14:5-9.
30. Adale E, et al. Prevalence of bovine trypanosomosis in Wolaita Zone Kindo Koish District of Ethiopia. *African J of Agri Research*. 2013;8:6383-6387.
31. Mulaw S, et al. Study on the Prevalence of Major Trypanosomes Affecting Bovine in Tsetse Infested Asosa District of BenishangulGumuz Regional State. Western Ethiopia. *Global Veterinaria*. 2011;7:330-336.
32. Gemeda F. Prevalence of Bovine Trypanosomosis in and around Nekemte Areas. East Wollega Zone. Ethiopia. *Open Access Library Journal*. 2015;2:1-7.

33. Berhanu M, et al. Mechanical Transmitted bovine trypanosomosis in Tselamitu woreda. Western Tigray .Northern Ethiopia. Agricultural journal.2011;6(1):10-13.
34. Bitew M, et al. Prevalence Of bovine trypanosomosis in selected areas of JabiTehenandistrict.westGojam of Ahmara regional state .north Western Ethiopia .Africa journal of Agricultural Research. 2011;6(1):141-144.
35. Gona Z, et al. Study on prevalence of bovine trypanosomosis and density of its vector in three selected districts of Wolaita zone. Southern Ethiopia. Journal of Veterinary Medicine and Animal Health. 2016; 8(9):128-135.
36. Yalew, et al. Prevalence of bovine trypanosomosis and its associated risk factors in Bambasiworeda. Western Ethiopia. Journal of dairy. Veterinary and Animals research. 2017;5(1):00132.
37. Dinka. A, et al. Study on the Prevalence of Bovine Trypanosomosis. Vector Density and Associated Risk Factors in Assosa District of the BenishangulGumuz Region. West Ethiopia. Europe. J. Appl. Sci. 2016;8(5):319-325.
38. Bekele, et al. Prevalence and host related risk factors of bovine trypanosomosis in Hawa-gelan district. West Wollega zone. Western Ethiopia. Afr J Agri Res. 2011;6:5055-5060.
39. Mulatu. E, et al. Prevalence of Bovine Trypanosomosis and Apparent Density of Tsetse Flies in Eastern Part of Dangur District. North Western Ethiopia. J Vet SciTechnol; 2016;7:347-353.
40. Dagnachew S, et al. Prevalence and vector distributions of bovine trypanosomosis in control (Sibu Sire) and noncontrol (GutoGida) districts bordering upper Anger valley of East Wollega Zone. Western Ethiopia. Ethiopian Vet J. 2011;15:77-86.
41. Alemayehu B, et al. Bovine trypanosomosis: A threat to cattle production in Chena district. Southwest Ethiopia. Open Jof Animal Sc. 2012;2:287-291.
42. Bishaw Y, et al. Prevalence of bovine trypanosomosis in Wemberma district of West Gojjam zone. North West Ethiopia. Ethiopian Vet J. 2012;16:41-48.