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# Review on Natural Excipients in the Formulation of Oral Fast Dissolving Films Mamatha M\*, Deepthi B , Sasidhar R

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# **Review Article**

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ABSTRACT

oral route is the most acceptable from patient compliance. Fast dissolving drug delivery system are used to overcome the problem of difficulty in swallowing tablets or capsules. This review mainly represents the use of natural excipients such as polymers, plasticizers, flavoring agents, colorants and sweeteners for the formulation of oral fast dissolving films was more advantageous over synthetic excipients. Because of patient compliance especially with paediatric and geriatric patients these films are prepared with natural excipients..

# INTRODUCTION

#### **Oral Fast Dissolving films**

"Oral Fast dissolving films drug delivery means a film dissolves or oral drug strip to administer drugs through buccal or sublingual absorption or it absorbs by enterically."

#### Advantages of oral films

- Dose convenience
- Less water requirement
- Zero risk of chocking
- Masking the taste
- Stability Enhancement
- Accurate dosing in comparison to syrup

#### Disadvantages of oral films

- Dose for incorporating into the strip is in between 1-30 mgonly
- A number of technical limitations with use of film strips; Thickness while casting the film. Casting cannot be done by glass petriplates.
- Achieving dose uniformity
- Special equipment should be used for packaging of oral films leads to difficulty in packaging

#### **Natural Excipients**

Natural excipients are preferred on the synthetic and semisynthetic ones because of their lack of toxicity, low cost, soothing action, availability, and nonirritant nature of the excipients.

Natural polymers

Because of cost efficacy and regulatory acceptance natural gums <sup>[1-3]</sup> are the most popular hydrophilic polymers

## **Advantages of Natural Polymers**

As the name indicates they are available in nature [4-13] so that they are Biodegradable in nature, and they are produced by all living organisms.

- All of these plant materials are reiterating sugar polysaccharides [14-25] these are biocompatible and nontoxic
- > When compared to synthetic materials cost of production is less for natural polymers
- > Large quantities of natural polymers are produced due to simple production processes are involved
- Minimum chance of adverse and side effects with natural polymers <sup>[26-36]</sup> when compared with synthetic materials

There is promotion being done by government for the plant production as pharmaceutical excipients, and it withal provides the facilities for bulk production, because of their wide applications like gum and mucilage's in industries In India and homogeneous developing countries. Natural polymers are various plant based materials. Plant-based material serves as an alternative to synthetic products because of different reasons:

- I. Local obtainability
- II. Ecological in nature
- III. Bio-acceptability
- IV. Having renewable source as well as lowest price when compared to synthetic products

#### Guar gum

It is also called guaran, is a galactomannan with high molecular weight of 8,000,000. It is obtained from the Guar plant as an endosperm seed Cyamopsis tetragonoloba (L) Taub. (Syn. Cyamopsis psoralioides). It is free flowing, consummately soluble, neutral polymer which is composed of sugar units and has also been approved for use in food. Guar gum <sup>[37, 38]</sup> and derivatives are used as binders and disintegrate in films and also used as a control-released agent for the drug. It is used in a concentration of 1% w/w as a disintegrant for the preparation of oral films.

#### Mangifera indica gum (MIG)

In various pharmaceutical formulations MIG is used as a disintegrating agent, binder <sup>[39-48]</sup>, suspending agent, and emulsifying agent because of it's non-toxic nature. It is used as a polymer in formulation of oral films.

#### Dehydrated banana Powder (DBP)

Additionally banana is called as plantain [49-58]. DBP is used as a superdisintegrant in the formulation of oral films. It is a very good source of energy due to high carbohydrate content, and it contains potassium, which is responsible for more preponderant brain functioning.

#### Pullulan

Pullulan is a natural and extracellular microbial polysaccharide produced by the fungus-like yeast, Aureobasidium pullulans. Pullulan can be made into very thin films (down to 0.01mm) which also have more tensile strength and can stable over a range of temperatures. Pullulan can be made into films of high tensile strength and low oxygen permeability, are oil and grease resistant. Pullulan films are usually prepared with 5-10% aqueous pullulan solution by rapid evaporation and applied to a smooth surface and dried; it may also involve the use of high temperature and pressure. Pullulan can be mixed with gelatin, amylose and polyvinyl alcohol <sup>[59-67]</sup> for better release of drug.

#### Plasticizers

Plasticizers are the additives that increase the plasticity or fluidity of a material. Plasticizer <sup>[68, 69]</sup> mailnly reduces the brittleness of the strip and imparts flexibility. Based on the compatibility with the polymer plasticizer should be selected and also based on the type of solvent used for casting of the strip.commonly used plasticizers in the formulation of oral films are in the concentration of 0-20% w/w of dry polymer weight.

- Glycerol
- Propylene glycol
- Castor oil
- > polyethylene glycols with low molecular weight
- > Tributyl citrates, Triethyl citrates, Actyl citrates

In comparison to citric acid, tartaric acid and oleic acid Malic acid was found to be better plasticizer as it did not crystallize out after drying of the films. Maltodextrin can also be plasticized as and converted with incorporation of glycerine as well as propylene glycol as plasticizer into oral dissolving film in the concentration range of 16-20% w/w, and found to be more advantageous by using glycerin over propylene glycol as it shows miscibility problems with maltodextrin [<sup>70, 71</sup>] either by using hot melt extrusion or solvent casting methods.

#### Coloring agents

Natural colorants are used in the formulation of oral films. All the colorants used in the formulation of oral fast dissolving films should not exceed 1% w/w as per FD&C. Following are some of the colorants used in the preparation of oral films.

- 1) Caramel also known as burnt sugar prepared by heating water
  - soluble carbohydrates with an accelerator until a black viscid mass is formed
- 2) Cochineal
- 3) Carmine

Other examples for natural colorants include

Riboflavin and Anthocyanins, Paprika Oleoresin, Beet Root Red, Annatto, Curcumin [72-90].

#### Sweetening agents

As the formulation has to be disintegrated in the oral cavity sweeteners become the important part. General concentration of sweeteners are in the concentration range of 3 to 6 % w/w either alone or in combination. Use of natural sugars in oral dissolving films need to be restricted in people who are on diet or in the case of diabetic patients. Because of this reason the artificial sweeteners <sup>[91]</sup> have gained more popularity in food and pharmaceutical preparations. First generation artificial sweeteners are Saccharin, cyclamate and aspartame. Second generation artificial sweeteners are acesulfame-K, sucralose, alitame and neotame. Bitter taste of fast dissolving films of diclofenac and ondansetron were suppressed by Sucralose <sup>[92, 93]</sup> and neotame respectively.

#### Flavoring agents

In oral fast dissolving films the concentration of flavors added is up to 10% w/w. Flavor <sup>[94-98]</sup> selection mainly depends on the type of drug to be incorporated in the **formulation (Table 1)**.

Flavors recommended	Basic taste
Wild cherry, walnut, chocolate	Bitter
Citrus flavor, licorice, root beer, raspberry.	Sour
Vanilla, fruit and berry	Sweet
Butterscotch, maple, apricot.	Salt

Table 1: Flavors for taste Masking

## CONCLUSION

Because of their rapid disintegration, improved dissolution properties principally with paediatric <sup>[99-102]</sup> and geriatric patients oral fast dissolving films are considered as most promising and important drug delivery systems. Natural excipients usage in the preparation of oral fast dissolving films is preferable due to less expensive, biodegradable, and ecofriendly nature rather than synthetic excipients.

#### REFERENCES

- 1. Shanmugam S. Oral Films: A Look Back. Clin Pharmacol Biopharm. 2016;5:124.
- 2. Ali MS et al. Formulation and Evaluation of Fast Dissolving Oral Films of Diazepam. J Pharmacovigilance 2016;4:210.
- 3. Ogaji IJ et al. Advances in Natural Polymers as Pharmaceutical Excipients. Pharm Anal Acta. 2012;3:146.
- 4. Parrott EL. Pharmaceutical Technology: Fundamental Pharmaceutics. 1971. Burgess Publishing
- Company, Minneapolis. IPECFED. The world unites for safer medicines. 2011.
  Russell R. Synthetic excipient challenge all-natural organics offer advantages/challenges to developer and formulators. Pharmaceutical Technology. 2004;38-50.
- 6. Guo J. Pharmaceutical applications of naturally occurring water-soluble polymers. 1998;PSTT 1:254-261.
- 7. Beneke CE. Polymeric Plant-derived Excipients in Drug Delivery. Molecules. 2009;14:2602-2620.
- 8. Pandey R and Khuller GK. Polymer based drug delivery systems for mycobacterial infections. Current Drug Delivery. 2004;1:195-201.
- 9. Chamarthy Sp and Pinal R. Plasticizer concentration and the performance of a diffusion-controlled polymeric drug delivery system. Colloids Surf A Physiochem Eng Asp. 2008;331:25-30.

- 10. Malafaya PB et al. Natural-origin polymers as carriers and scaffolds for biomolecules and cell delivery in tissue engineering applications. Adv Drug Deliv Rev. 2007;59:207-233.
- 11. Perepelkin KE. Polymeric materials of the future based on renewable plant resources and biotechnologies. Fibre Chemistry. 2005;37:417-430.
- 12. Malviya R et al. Applications of Mucilages in Drug Delivery A Review. Advances in Biological Research. 2011;5:1-7.
- 13. Jayasinghe PS et al. Effect of Extraction Methods on the Yield and Physiochemical Properties of Polysaccharides Extracted from Seaweed Available in Sri Lanka. Poult Fish Wildl Sci. 2016;4:150.
- 14. Sanalibaba P and Çakmak GA. Exopolysaccharides Production by Lactic Acid Bacteria. Appli Micro Open Access. 2016;2:115.
- 15. Pillai TG and Mini M. Bacterial Polysaccharides Potential Candidate for Vaccine Development. J Med Microb Diagn. 2016;5:224.
- 16. Sharma SK. Optimized Extraction and Antioxidant Activities of Polysaccharides from Two Entomogenous Fungi. J Bioanal Biomed. 2015;7:180-187.
- 17. Plazanet I et al. Direct Immunological Detection of Wood Cell Wall Polysaccharides after Microwave-Assisted Ionic Liquid Disruption. J Glycobiol. 2015;3:115.
- 18. Marudhupandi T and Inbakandan D. Polysaccharides in Aquatic Disease Management. Fish Aquac J. 2015;6:135.
- 19. Mohamed HA et al. Physicochemical Properties of Tamarind (Tamarindus indica) Seed Polysaccharides. J Food Process Technol. 2015;6:452.
- 20. Park JK. Algal Polysaccharides: Properties and Applications. Biochem Anal Biochem. 2015;4:176.
- 21. Ogutu FO et al. Ultrasonic Modification of Selected Polysaccharides-Review. J Food Process Technol. 2015;6:446.
- 22. Pawar HA et al. An Overview of Natural Polysaccharides as Biological Macromolecules: Their Chemical Modifications and Pharmaceutical Applications. Biol Med. 2015;7:224.
- 23. Ahmed OM and Ahmed RR. Anti-Proliferative and Apoptotic Efficacies of Ulvan Polysaccharides against Different Types of Carcinoma Cells In Vitro and In Vivo. J Cancer Sci Ther. 2014;6:202-208.
- 24. Abd El Baky H et al. Induction of Sulfated Polysaccharides in Spirulina platensis as Response to Nitrogen Concentration and its Biological Evaluation. J Aquac Res Development. 2013;5:206.
- 25. William BJ. Ask the Historian: The origin of the polymer concept. J Chem Edu. 2008;88:624-625.
- 26. Shukla RK and Tiwari A. Carbohydrate polymers: Applications and recent advances in delivering drugs to the colon. Carbohydr Polym. 2012;88:399–416.
- 27. Liu Z et al. Polysaccharides-based nanoparticles as drug delivery systems. Adv Drug Deliv Rev. 2008;60:1650–1662.
- 28. Saravanakumar G et al. Polysaccharide based nanoparticles: A versatile Platform for Drug Deliveryand Biomedical Imaging. Curr Med Chem. 2012;19:3212–3219
- 29. Satturwar PM et al. Biodegradation and in vivo biocompatibility of rosin: A natural film-forming polymer. AAPS Pharm Sci Tech. 2003;4:1-6.
- 30. Leakey BRR. Potential for novel food products from agroforestry trees. Areview. J Food Chemistry. 1999;66:1-14.
- 31. Marathee RM. Gelling behavior of polyose from tamarind kernelpolysaccharides. Food Hydrocolloids. 2002;16:423-426.
- 32. Khanna M et al. Polyose from seeds of Tamarindusindica of unique property and immense pharmaceutical use in trends in carbohydrate chemistry. 1997;4:97-81.
- 33. Graham HG and Uan. Extraction of soluble fiber. J Agric Food chem. 1988;36:494-497.
- 34. MarletteJA et al. Recovery of dietary fibre is dependent on the method of analysis. Am J Clin Nutr. 1989;50:479-485.
- 35. ChakravortilB et al. Detrmination of molecular weight of tamarindKernel polysaccharide. Indian J Technology. 1963;1:216-217.
- 36. Rodge AB et al. Effect of Hydrocolloid (guar gum) Incorporation on the Quality Characteristics of Bread. J Food Process Technol. 2012;3:136.
- 37. Krishnamoorthy RV. Potential of Perionyxexcavatusfor utilizing organicwastes. Pedobiologia. 1982;23:419-425.
- 38. Kar SS et al. Significance of RAP Content and Foamed Binder Content on Mechanistic Characteristics of Recycled Foamed Bituminous Mixes. J Civil Environ Eng. 2016;6:220.
- 39. Mishra S and Gomase VS. Prediction of Antigenic MHC Peptide Binders and TAP Binder of COX1 Protein through in silicoApproach. J Drug Metab Toxicol. 2016;7:201.
- 40. Mishra S and Gomase VS. "Cytochrome B"- Analysis of Hydrophobicity, Surface Accessibility, Antigenicity and Prediction of MHC I and MHC II Binders from Dracunculiasis. Med chem. 2016;6:041-046.
- 41. Begum S et al. Physical Characterization and Microstructure Evaluation of Titanium Dioxide Semiconductor Discs Processed with Binders. J Material Sci Eng. 2013;2:126.

- 42. Sangvanich T et al. Nanoporous Sorbent Material as an Oral Phosphate Binder and for Aqueous Phosphate, Chromate, and Arsenate Removal. J Nanomed Nanotechnol. 2014;5:222.
- 43. Grassmann A et al. ESRD patients in 2004: global overview of patient numbers, treatment modalities and associated trends. Nephrol Dial Transplant. 2005;20:2587-2593.
- 44. Hutchison AJ et al. Pharmacology, efficacy and safety of oral phosphate binders. Nat Rev Nephrol. 2011;7:578-589.
- 45. Hutchison AJ et al. Histological, radiological, and biochemical features of the adynamic bone lesion in continuous ambulatory peritoneal dialysis patients. Am J Nephrol. 1994;14:19-29.
- 46. Goodman WG et al. Vascular calcification in chronic kidney disease. Am J Kidney Dis. 2004;43:572-579.
- 47. 48. Tonelli M et al. Oral phosphate binders in patients with kidney failure. N Engl J Med. 2010;362:1312-1324.
- 48. Adeyanju JA et al. Optimisation of Deep-Fat Frying of Plantain Chips (Ipekere) using Response Surface Methodology. J Food Process Technol. 2016;7:584.
- 49. Sobowale SS et al. Effect of Extrusion Variables on the Extrudate Properties of Wheat-plantain Noodle. J Food Process Technol. 2016;7:547.
- 50. Aby N et al. Inoculated Traps, an Innovative and Sustainable Method to Control Banana Weevil Cosmopolites Sodidus in Banana and Plantain Fields. Adv Crop Sci Tech. 2015;3:194.
- 51. Okorie DO et al. Nutrient and Heavy Metal Composition of Plantain (Musa paradisiaca) and Banana (Musa paradisiaca) Peels. J Nutr Food Sci. 2015;5:370.
- 52. Kanazawa K and Sakakibara H. High content of dopamine, a strong antioxidant, in Cavendish banana. J Agric Food Chem. 2000;48:844-848.
- 53. Mohapatra D et al. Plantain and its byproduct utilization: An overview. J SciIndRes. 2010;69:323-329.
- 54. Ibrahim KS et al. Transcriptome Analysis in Banana. Transcriptomics. 2015;3:117.
- 55. Tortoe C et al. Multilinear Regression Approach in Predicting Osmo-Dehydration Processes of Apple, Banana and Potato. J Food Process Technol. 2011;2:122.
- 56. Farkas BE and Hubbard LJ. Analysis of heat transfer during immersion frying. Dry Technol. 2000;18:1269-1285.
- 57. Vitrac O et al. Characterization of heat and mass transfer during deep-fat frying and its effect on cassava chip quality. J Food Eng. 2002;53: 161-176.
- 58. Islam T et al. Studies on Swelling and Absorption Properties of the γ Irradiated Polyvinyl Alcohol (PVA)/Kappa-Carrageenan Blend Hydrogels. J Adv Chem Eng. 2016;6:153.
- 59. Gulenoor F et al. γ-Irradiated Polyvinyl Alcohol (PVA) and Citric Acid Blend Hydrogels: Swelling and Absorption Properties. Chem Sci J. 2016;7:125.
- 60. Sadhu SD et al. Thermal Studies of the Starch and Polyvinyl Alcohol based Film and its Nano Composites. J Nanomedic Nanotechnol. 2015;S7:002.
- 61. Chowdhury AMS et al. Studies on the γ-Irradiated Polyvinyl Alcohol (PVA) Blended Gelatin Films. J Adv Chem Eng. 2015;5:141.
- 62. Kawakami T et al. Analysis of 19 preservatives in Polyvinyl Alcohol Cooling Towels Used in Japan by High Performance Liquid Chromatography with Photo Diode Array Detector. J Environ Anal Chem. 2015;2:122.
- 63. Muppalaneni S and Omidian H. Polyvinyl Alcohol in Medicine and Pharmacy: A Perspective. J Develop Drugs. 2013;2:112.
- 64. Mabrouk M et al. Fabrication, Characterization and Drug Release of Ciprofloxacin Loaded Porous Polyvinyl Alcohol/Bioactive Glass Scaffold for Controlled Drug Delivery. Bioceram Dev Appl. 2013;S1:009.
- 65. Hassanien AM et al. Fabrication of Polyvinyl Alcohol/Cellulose Acetate (PVA/CA/PEG) Antibacterial Membrane for Potential Water Purification Application. Hydrol Current Res. 2013;4:146.
- 66. Muhammed A et al. Polyvinyl Alcohol-Cellulose Acetate Composite Reverses Osmosis Membranes: I. Synthesis and Characterization. Hydrol Curr Res. 2012;3:131.
- 67. Iwase H et al. Biological Effects of the Plasticizer Tris (2-Ethylhexyl) Trimellitate. Clin Pharmacol Biopharm. 2014;S2:004.
- 68. Shehata AS et al. Effects of Exposure to Plasticizers Di-(2-Ethylhexyl) Phthalate and Trioctyltrimellitate on the Histological Structure of Adult Male Albino Rats' Liver. J Clin Toxicol. 2013;3:169.
- 69. Syed HM et al. Studies on Preparation of Low Calorie Cake using Pearl Millet (Bajra) Maltodextrin. J Food Process Technol. 2011;2:125.
- 70. Fragoso LR et al. Pharmacokinetics of Maltodextrin Coated Cadmium Sulfide Quantum Dots in Rats. J Nanomedine Biotherapeutic Discov. 2016;6:139.
- 71. Satapathy P et al. Attenuation of Dopaminergic Neuronal Dysfunction in Caenorhabditis elegans by Hydrophilic Form of Curcumin. Neurochem Neuropharm Open Access. 2016;2:111.
- 72. Su K et al. Preparation of Polymeric Micelles of Curcumin with Pluronic P123 and Assessment of Efficacy against B16 Cells In vitro. Adv Pharmacoepidemiol Drug Saf. 2016;5:202.
- 73. Alwi I et al. The Effects of Curcumin against the Inflammatory Response in Patients with Acute Coronary Syndrome. Cardiovasc Pharm Open Access. 2016;5:185.

- 74. Tokumoto T. Effect of Curcumin on Sulfasalazine Pharmacokinetics in Healthy Volunteers. J Drug Metab Toxicol. 2016;7:206.
- 75. Luo S. Two Novel Curcumin Analogues Induced Reactive Oxygen Species Generation and Mitochondrial-Related Apoptosis in Human Breast Cancer MCF - 7 Cells. J App Pharm. 2016;8:215.
- 76. Naglaa AB et al. Protective Effect of Curcumin versus N-acetylcystein on Acetaminophen Induced Hepatotoxicity in Adult Albino Rats. J Cytol Histol. 2015;S3:018.
- 77. Prasad S and Tyagi AK. Curcumin and Cystic Fibrosis Defects: A Spicy Treatment. Genetics. 2015;S7:e001.
- 78. Jain A. Curcumin Inhibit PhIP-Induced Carcinogenicity by Regulating Expression of Nrf2 and FOXO Targets, and BRCA-1 and P16 Expression in Breast Epithelial Cells. J Carcinog Mutagen. 2015;6:236.
- 79. Figueroa AP et al. Evaluation of the Neuroprotective Effects of Curcuminoids on B35 and SH-SY5Y Neuroblastoma Cells. Med Aromat Plants. 2015;4:197.
- 80. Menaa F. Curcumin Nano-Sized Delivery Systems against Cancers: From Bench to Clinics. J Pharma Care Health Sys. 2015;2:e135.
- 81. Morningstar MW et al. Controlled Release Curcumin for the Treatment of Pain Related to Adult Degenerative Scoliosis: A Retrospective, Open-Label, Case-Controlled Series. J Pain Relief. 2015;4:192.
- 82. Karri VVSR et al. Multiple Biological Actions of Curcumin in the Management of Diabetic Foot Ulcer Complications: A Systematic Review. Trop Med Surg. 2015;3:179.
- 83. Azad GK and Tomar RS. Epigenetics of Curcumin: A Gifted Dietary Therapeutics Compound. J Carcinog Mutagen. 2015;6:206.
- 84. Jacob PS et al. Use of Curcumin in Periodontal Inflammation. Microinflammation. 2014;1:114.
- 85. Chaturvedi P et al. Media Optimization in Immobilized Culture to Enhance the Content of Curcumin in Curcuma longa (Zingiberaceae) and Protein Profile of Treated Samples in Static Culture. Nat Prod Chem Res. 2014;S1:002.
- 86. Shehzad A. Curcumin: Ancient Drug, Modern Challenges, Malignant Pancreatitis. Pancreat Disord Ther. 2013;3:e131.
- 87. 88. Sagi SSK et al. Prophylactic Administration of Curcumin Abates the Incidence of Hypobaric Hypoxia Induced Pulmonary Edema in Rats: A Molecular Approach. J Pulm Respir Med. 2014;4:164.
- 88. Muthumani M. Tetrahydrocurcumin Potentially Attenuates Arsenic Induced Oxidative Hepatic Dysfunction in Rats. J Clin Toxicol. 2013;3:168.
- 89. Hejazi J et al. A Pilot Clinical Trial of Radioprotective Effects of Curcumin Supplementation in Patients with Prostate Cancer. J Cancer Sci Ther. 2013;5:320-324.
- 90. Gaikwad KK et al. "Studies on the Development and Shelf Life of Low Calorie Herbal Aonla-Ginger RTS Beverage by Using Artificial Sweeteners". J Food Process Technol. 2013;4:200.
- 91. Hussein AMS et al. Utilization of Yoghurt and Sucralose to Produce Low-calorie Cakes. J Nutr Food Sci. 2016;6:447.
- 92. Tamimi L et al. Pioglitazone HCl Levels and Its Pharmacokinetic Application in Presence of Sucralose in Animals Serum by HPLC Method. Pharm Anal Acta. 2014;5:318.
- 93. Mnaa S et al. Effects of Heavy Metals and Monosodium L-Glutamate in Food Flavors on Albino Rats. J Biomol Res Ther. 2015;4:127.
- 94. Zimba PV and Grimm CC. Statistical Approaches to Optimize Detection of MIB Off-Flavor in Aquaculture Raised Channel Catfish. J Aquac Res Development. 2015;6:319.
- 95. Green BW and Schrader KK. Effect of Stocking Large Channel Catfish in a Biofloc Technology Production System on Production and Incidence of Common Microbial Off-Flavor Compounds. J Aquac Res Development. 2015;6:314.
- 96. Ruan ED et al. Sensitive Analysis of Off-flavor Compounds, Geosmin and 2-Methylisoborneol, in Water and Farmed Sturgeon by using Stir Bar Sorptive Extraction Coupled with Thermal Desorption and Gas Chromatography-Mass Spectrometry. J Chromatograph Separat Techniq. 2014;5:228.
- 97. Gowder SJ T. Safety Assessment of Food Flavor Cinnamaldehyde. Biosafety. 2014;3:e147.
- 98. Giardino PA. Child Abuse and Neglect: Are Cases Increasing or Decreasing After 50+ Years of Paediatric Attention?. Clinics Mother Child Health. 2016;13:235.
- 99. Newton LE et al. Stabilization of Nasoenteric Feeding Tubes Using Nasal Bridles In Paediatric Patients. Matern Pediatr Nutr. 2016;2:111.
- 100. Jeganathan VSE et al. Paediatric Aphakic Glaucoma: A Diagnostic and Management Challenge. Optom Open Access. 2016;1:115.
- 101. Srivastava S et al. Importance of Subtle Behavioural Changes in the Diagnosis of Paediatric Psychosis. J Neuropsychopharmacol Mental Health. 2016;1:108.