Role of Xenobiotics and its Biodegradation

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Review Article

ABSTRACT

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A xenobiotic is a synthetic foreign substance present in a living being which is not ordinarily normally created or exhibit inside the life form. Chemicals that are foreign to the biosphere are known as xenobiotic compounds. In particular, medications, for example, antibiotics are xenobiotics in humans because the human body does not deliver them itself, nor they part of a normal food. Natural compounds can likewise get to be xenobiotics on the off chance that they are taken up by another life form. for example, the uptake of normal human hormones by fish discovered downstream of sewage treatment plant outfalls, chemical defenses produced by some organisms as protection against predators. The body evacuates xenobiotics by xenobiotic metabolism. Xenobiotic organs would be created in a manner that they would not be rejected by the resistant framework. Any xenobiotics produce an assortment of natural impacts, which is utilized when they are portrayed utilizing bioassay. The term xenobiotic is likewise used to organs transplanted starting with one animal type to another.

INTRODUCTION

Microbial depolymerisation procedures are sorted into two sorts, exogenous type forms and endogenous type forms^[1-3]. In an exogenous sort depolymerisation process, molecules reduce in size through freedom of monomers from their terminals. Biodegradability of PEG has been accounted for. PEG is depolymerized by freeing C2 mixes[4,5]. Systems produced for the exogenous sort depolymerisation procedures of PE were reached out to the exogenous depolymerisation procedures of PEG. Xenobiotic receptors including individuals from the atomic and solvent interpretation element superfamilies, can intercede the metabolic reaction of life form to the compound environment. The various studies are directed in xenobiotic receptors, the instrument about how these receptors play out their capacity because of xenobiotic test and their parts in the advancement of different diseases, for example, growth diabetes^[6-10].

The metabolic pathways related with xenobiotic receptors can be distinguished in light of the consequences of metabolomics examination. Metabolomics will assume more vital part in clarification of the capacity of xenobiotic receptors later on^[11,12]. Metabolomics, one high-throughput explanatory innovation, can efficiently profile the endogenous metabolites in biofluid, cell, and tissue. Right now, the ultra-execution fluid chromatography combined with electrospray ionization quadrupole time-of-flight mass spectrometry, gas chromatography mass spectrometry, and atomic attractive reverberation are the major systematic methods for metabolomics^[13-15].

Metabolomics, one high-throughput investigative innovation, can deliberately profile the endogenous metabolites in cell and tissue which has been generally used to distinguish the biomarkers for clinical infection^[16]. These metabolites can be at first resolved to be medication metabolites or endogenous metabolites taking into account their drifting plots^[17-23]. The further distinguishing proof of these metabolites will be performed through their MS/MS range and examination with valid compound. The examination of metabolomics, medication metabolites can be methodically decided and the potential poisonous metabolites can be recognized. All the more critically, the biomarkers instigated by medication introduction can be utilized to foresee drug activity or poisonous quality. What's more, between individual varieties uncovered by metabolomics can give the import data to customized drug in the centre^[25-10]. Wastewater slop rheology has made some amazing progress from being a traditional device to control wastewater treatment procedure to likewise permitting picking the best innovation for evacuation of developing contaminants furthermore the quality expansion courses of waste water^[31,32].

Biodegrability of xenobiotics

Xenobiotic compunds are chemicals which are outside to the biosphere. The physicochemical properties of the earth may influence and even control biodegradation execution. Sorption, immobilization and micropore entanglement are significant reasons for the ingenuity of numerous xenobiotics [33-36]. The structure of xenobiotic atoms is portrayed by "unphysiological" substituents and stable synthetic bonds, which block or even counteract biodegradation. The rates of xenobiotic biodegradation in nature may run from days and weeks to years and decades. In soil, for instance, oxygen accessibility is all the time the constraining variable of high-impact biodegradation forms. In addition, the nearness of contending microorganisms, or of predators brushing on the microbial consortium, additionally influences biodegradation^[37-45].

Ecological toxicants and xenobiotics trigger tissue and cell harm by bringing about necrotic cell passing which is a sudden and unregulated procedure^[45-52]. This idea has been tested in the previous decade because of the growing comprehension of the instruments of apoptosis-a customized cell passing. Ginger has been appeared to be a hepatoprotective specialist, and studies with different hepatotoxins like ethanol, paracetamol, lead, cadmium, antibiotic medication, organophosphorus mixes accept the property^[53-60].

Xenobiotic compounds, attributed to its refractory nature, are difficult to separate and corrupt. The multifaceted nature of its concoction piece adds to this [61-65]. For separating such intensifies the chemicals follow up on specific gatherings present in the compound. Regularly it is seen that the xenobiotics don't go about as a wellspring of vitality to organisms and subsequently they are not debased. The nearness of an appropriate substrate impels its breakdown. This substrate is known as co – metabolite and the procedure of corruption are known as co digestion system^[66,67]. In another procedure, the xenobiotics serve as substrates and are followed up on to discharge vitality. This is called needless digestion system. Detoxification is a procedure that declines the negative effect of xenobiotics or poisons, on the body. Practicing detoxification scrubs the body from inside and diminishes the reactions of xenobiotics^[68-73].

Xenobiotic compounds are man-made chemicals that are available in the climate at strangely high fixations. In any case, few xenobiotics are there that are impervious to microbial assault. Microorganisms can process the greater part of the actually happening xenobiotic mixes and this property is called as microbial trustworthiness^[74-79]. Those xenobiotic aggravates that oppose absorption from even organisms are called obstinate. Biodegradation execution in the regular living space is influenced and controlled by the physicochemical properties of the earth. Examination ought to concentrate on comprehension system of the association amongst microorganisms and xenobiotic compounds in the environment that must incorporate biochemical and in addition genetic engineering areas^[80-85].

Numerous microscopic organisms and parasites produce proteins that can follow up on an extensive variety of natural mixes. A non-development substrate is one that can't serve as the sole wellspring of carbon and vitality for an immaculate society of a bacterium and consequently can't bolster cell division [86-89]. Serendipitous digestion system and co-digestion system assume imperative parts in the expulsion of xenobiotic mixes from nature. Detoxification of toxins may include as solitary adjustment of structure to render a conceivably unsafe substance harmless. Ecologically satisfactory biodegradation, in which the base adjustment of the guardian compound important to expel properties happens [90]. Numerous lethal xenobiotics are dynamically more gathered in every connection of an evolved way of life, a procedure called bio-amplification^[91-94].

Sources of aromatic compounds in the environment include degradation of lignin in plants, use of detergents, pesticides, drugs and dyes etc. Several polycyclic aromatic hydrocarbons (PAH) released from industrial processes are carcinogenic [95,96]. Chloroaromatic compounds are the pollutants of major concern and are toxic, resist to biodegradation. Chlorobenzene, dichlorobenzene and trichlorobenzene are not easily biodegraded in biological treatment system. However, they are degraded by some soil microorganisms and also co-metabolically by strains of Pseudomonas putida. Chlorophenols and chlorocatechol are the intermediate compounds in the biodegradation of chlorobenzenes, various pesticides and other chloroaromatic compounds^[97-100].

REFERENCES

- 1. Watanabe M and Kawai F. Study on Role of Microorganisms in Depolymerization Processes of Xenobiotic Polymers. J Environ Anal Chem 2014;2:119.
- 2. Li F. Metabolomics: One Powerful Tool for the Understanding of Xenobiotic Receptors. J Chromat Separation Techniq. 2012;3:e107.

- 3. Li F. Xenobiotic Metabolomics: An Ideal Tool for Drug Metabolism. J Chromat Separation Techniq. 2012;3:e106.
- 4. Brar SK. Rheology Key to Process Control in Conventional and Advanced Wastewater Treatment Plants. Hydrol Current Res. 2012;3:e102.
- 5. Ding WX. Autophagy in Toxicology: Defense against Xenobiotics. J Drug Metab Toxicol. 2012;3:e108.
- 6. Loretta OO, et al. In Vitro Biodegradation of Palm Oil Mill Effluent (POME) by Bacillus subtilis, Pseudomonas aeruginosa and Aspergillus niger. J Bioremed Biodeg. 2016;7: 361.
- 7. Prabhavathi P, et al. Molecular Docking Studies on Potent Adsorbed Receptor of Protein: A New Target for Biodegradation of Indigo Dye. J Bioremed Biodeg. 2016;7:356.
- 8. Alariqi SAS, et al. Effect of Different Sterilization Methods on Biodegradation of Biomedical Polypropylene. J Environ Anal Toxicol. 2016;6:373.
- 9. Abd El-Ghany TM and Masmali IA. Fungal Biodegradation of Organophosphorus Insecticides and their Impact on Soil Microbial Population. J Plant Pathol Microbiol. 2016;7:349.
- 10. Jablonski MR, et al. Novel Photo-fenton Oxidation with Sand and Carbon Filtration of High Concentration Reactive Dyes both with and without Biodegradation. J Textile Sci Eng. 2016;6:251.
- 11. Prince RC, et al. Three Widely-Available Dispersants Substantially Increase the Biodegradation of Otherwise Undispersed Oil. J Marine Sci Res Dev. 2016;6:183.
- 12. Anike FN, et al. Co-Substrating of Peanut Shells with Cornstalks Enhances Biodegradation by Pleurotus ostreatus. J Bioremed Biodeg. 2016;7:327.
- 13. Isiodu GG, et al. Role of Plasmid-Borne Genes in the Biodegradation of Polycyclic Aromatic Hydrocarbons (PAHs) by Consortium of Aerobic Heterotrophic Bacteria. J Pet Environ Biotechnol. 2016;7:264.
- 14. Owhonka A and Gideon OA. The Role of Aerobic Microorganisms in the Biodegradation of Petroleum Hydrocarbons Laboratory Contaminated Groundwater. Fermentol Techno. 2015;4:122.
- 15. Lim SJ. Biodegradation: Enzymes Evolution. J Bioremed Biodeg. 2015;6:e168.
- 16. Shu CH, et al. Improving Biodegradation of Rice Straw Using Alkaline and Aspergillus niger Pretreatment for Methane Production by Anaerobic Co-Digestion. J Bioprocess Biotech. 2015;5:256
- 17. Elghonemy D. Role of Microbial Enzymes in the Biodegradation of Rice Straw via Biotechnological Techniques. J Rice Res. 2015;3:e120.
- Saborimanesh N and Mulligan CN. Effect of Sophorolipid Biosurfactant on Oil Biodegradation by the Natural Oil-Degrading Bacteria on the Weathered Biodiesel, Diesel and Light Crude Oil. J Bioremed Biodeg. 2015;6:314.
- 19. Chikere CB, et al. Molecular Assessment of Microbial Species Involved in the Biodegradation of Crude Oil in Saline Niger Delta Sediments Using Bioreactors. J Bioremed Biodeg. 2015;6:307.
- 20. Ipeaiyeda AR, et al. Biodegradation of Polycyclic Aromatic Hydrocarbons in Agricultural Soil Contaminated with Crude Oil from Nigeria Refinery using Pleurotus sajor-caju. J Bioremed Biodeg. 2015;6: 301.
- 21. Manoj B. Biodegradation of Coal Minerals by Gluconic Acid and its Effect on the Stacking Structure of Carbon: An Investigation. J Bioremed Biodeg. 2015;6:306.
- 22. El Mahdi AM, et al. Performance of Isolated Kocuria sp. SAR1 in Light Crude Oil Biodegradation. J Bioremed Biodeg. 2015;6:303.
- 23. Alvarenga N, et al. Biodegradation of Chlorpyrifos by Whole Cells of Marine-Derived Fungi Aspergillus sydowii and Trichoderma sp. J Microb Biochem Technol. 2015;7:133-139.
- 24. Oje Obinna A, et al. Variation in the Carbon (C), Phosphorus (P) and Nitrogen (N) Utilization during the Biodegradation of Crude Oil in Soil. J Pet Environ Biotechnol. 2015;6:206.
- 25. Mohapatra S and Pandey M. Biodegradation of Hexachlorocyclohexane (HCH) Isomers by White Rot Fungus, Pleurotus florida. J Bioremed Biodeg. 2015;6:280.
- 26. Paniagua-Michel J and Rosales A. Marine Bioremediation A Sustainable Biotechnology of Petroleum Hydrocarbons Biodegradation in Coastal and Marine Environments. J Bioremed Biodeg. 2015;6:273.
- 27. Godheja J, et al. Biodegradation of One Ring Hydrocarbons (Benzene and Toluene) and Two Ring Hydrocarbons (Acenapthene and Napthalene) by Bacterial Isolates of Hydrocarbon Contaminated Sites Located in Chhattisgarh: A Preliminary Study. J Pet Environ Biotechnol. 2015;6:202.
- 28. Kumar A, et al. Biodegradation of Feather by Microsporum fulvum Singly or in Combination with Other Fungi. J Bioremed Biodeg. 2014;5:265.
- 29. Mathews S. Biodegradation of Polychlorinated Biphenyls (PCBs), Aroclor 1260, in Wastewater by Isolate MD2 (Pseudomonas aeruginosa) from Wastewater from Notwane Sewage Treatment Plant in Gaborone, Botswana. J Bioremed Biodeg. 2014;5:266.
- 30. Adongbede EM and Sanni RO. Biodegradation of Engine Oil by Agaricus campestris (A White Rot Fungus). J Bioremed Biodeg. 2014;5:262.
- 31. Elmahdi AM, et al. Optimization of Libyan Crude Oil Biodegradation by Using Solid Waste Date as a Natural Low-Cost Material. J Bioremed Biodeg. 2014;5:252.

- 32. Abdeen Z, et al. Enhancement of Crude Oil Biodegradation by Immobilizing of different Bacterial Strains on Porous PVA Hydrogels or Combining of them with their produced Biosurfactants. J Pet Environ Biotechnol. 2014;5:192.
- 33. Ichor T, et al. Biodegradation of Total Petroleum Hydrocarbon by Aerobic Heterotrophic Bacteria Isolated from Crude Oil Contaminated Brackish Waters of Bodo Creek. J Bioremed Biodeg. 2014;5:236
- 34. Agarry SE and Oghenejoboh KM. Biodegradation of Bitumen in Soil and Its Enhancement by Inorganic Fertilizer and Oxygen Release Compound: Experimental Analysis and Kinetic Modelling. J Microbial Biochem Technol. 2014;S4:002.
- 35. Shah MP. Biodegradation of Azo Dyes by Three Isolated Bacterial Strains: An Environmental Bioremedial Approach. J Microbial Biochem Technol. 2014;S3:007.
- 36. Godheja J and Shekhar SK. Biodegradation of Keratin from Chicken Feathers by Fungal Species as a Means of Sustainable Development. J Bioremed Biodeg. 2014;5:232
- 37. Hu FC, et al. Identification and degradation capability of three pyrene-degrading Gordonia sp. strains. J Appl Ecol. 2011;22:1857-1862.
- 38. Noorjahan CM. Physicochemical Characteristics, Identification of Bacteria and Biodegradation of Industrial Effluent. J Bioremed Biodeg. 2014;5:219.
- 39. Sims GK. Bioavailability in Biodegradation and Function of Herbicides. J Bioremed Biodeg. 2014;5:e144.
- 40. Muthukumar T, et al. Biodegradation of Starch Blended High Density Polyethylene using Marine Bacteria Associated with Biofilm Formation and its Isolation Characterization. J Microb Biochem Technol. 2014;6:116-122.
- 41. Mekuto L, et al. Biodegradation of Free Cyanide Using Bacillus Sp. Consortium Dominated by Bacillus Safensis, Lichenformis and Tequilensis Strains: A Bioprocess Supported Solely with Whey. J Bioremed Biodeg. 2013;S18:004.
- 42. Obayori OS, et al. Biodegradation of Fresh and Used Engine Oils by Pseudomonas aeruginosa LP5. J Bioremed Biodeg. 2014;5:213.
- 43. Christiane A, et al. Biodegradation of Reactive Blue 4 and Orange G by Pycnoporus sanguineus Strain Isolated in Gabon. J Bioremed Biodeg. 2013;4:206.
- 44. Bihari Z. Current Trends in Bioremediation and Biodegradation: Next-Generation Sequencing. J Bioremed Biodeg. 2013;4:e138.
- 45. Kumar R, et al. Enhanced Biodegradation of Mobil Oil Hydrocarbons by Biosurfactant Producing Bacterial Consortium in Wheat and Mustard Rhizosphere. J Phylogenetics Evol Biol. 2013;4:158.
- 46. Afify AEMMR, et al. Stimulating of Biodegradation of Oxamyl Pesticide by Low Dose Gamma Irradiated Fungi. J Plant Pathol Microb. 2013;4:201.
- 47. Sukirtha TH and Usharani MV. Production and Qualitative Analysis of Biosurfactant and Biodegradation of the Organophosphate by Nocardia mediterranie. J Bioremed Biodeg 2013;4:198.
- 48. Patidar A, et al. Potential of Microbial Inoculated Water Hyacinth Amended Thermophilic Composting and Vermicomposting in Biodegradation of Agro-Industrial Waste. J Bioremed Biodeg; 2013;4:191.
- 49. Bhatia M, et al. Implicating Nanoparticles as Potential Biodegradation Enhancers: A Review. J Nanomed Nanotechol. 2013;4:175.
- 50. Sims GK. Current Trends in Bioremediation and Biodegradation: Stable Isotope Probing. J Bioremed Biodeg. 2013;4:e134.
- 51. Gerloff T. Impact of genetic polymorphisms in transmembrane carrier-systems on drug and xenobiotic distribution. T. Naunyn-Schmiedeberg's Arch Pharmacol. 2004;369:268.
- 52. Robertson JD and Orrenius S. Molecular mechanisms of apoptosis induced by cytotoxic chemicals. Crit Rev Toxicol. 2000;30:609-627.
- 53. He S, et al. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-alpha. Cell 2009;137:1100-1111.
- 54. Ding WX, et al. Bid-dependent generation of oxygen radicals promotes death receptor activation-induced apoptosis in murine hepatocytes. Hepatology. 2004;40:403-413.
- 55. Ding WX and Yin XM. Dissection of the multiple mechanisms of TNF-alpha-induced apoptosis in liver injury. J Cell Mol Med. 2004;8:445-454.
- 56. Wang Z, et al. The mitochondrial phosphatase PGAM5 functions at the convergence point of multiple necrotic death pathways. Cell 2012;148:228-243.
- 57. Sun L, et al. Mixed lineage kinase domain-like protein mediates necrosis signaling downstream of RIP3 kinase. Cell. 2012;148:213-227.
- 58. Cho YS, et al. Phosphorylation-driven assembly of the RIP1-RIP3 complex regulates programmed necrosis and virus-induced inflammation. Cell. 2009;137: 1112-1123.
- 59. Mizushima N, et al. Autophagy fights disease through cellular self-digestion. Nature 2008;451:1069-1075.
- 60. Ding WX. Role of autophagy in liver physiology and pathophysiology. World J Biol Chem. 2010;1: 3-12.
- 61. Pyo JO, et al. Essential roles of Atg5 and FADD in autophagic cell death: dissection of autophagic cell death into vacuole formation and cell death. J Biol Chem. 2005;280:20722-20729.

- 62. Cho JY, et al. Urinary metabolomics in Fxr-null mice reveals activated adaptive metabolic pathways upon bile acid challenge. J Lipid Res. 2010;51:1063-1074.
- 63. Li F, et al. Metabolomics reveals an essential role for peroxisome proliferator-activated receptor alpha in bile acid homeostasis. J Lipid Res. 2012;53:1625-1635.
- 64. Li F, et al. Comparative metabolism of cyclophosphamide and ifosfamide in the mouse using UPLC-ESI-QTOFMS-based metabolomics. Biochem Pharmacol. 2010;80:1063-1074.
- 65. Patterson AD, et al. Metabolomics reveals attenuation of the SLC6A20 kidney transporter in nonhuman primate and mouse models of type 2 diabetes mellitus. J Biol Chem. 2011;286:19511-19522.
- 66. Patterson AD, et al. Human urinary metabolomic profile of PPARalpha induced fatty acid beta-oxidation. J Proteome Res. 2009;8:4293-4300.
- 67. IdleJR and Gonzalez FJ. Metabolomics. Cell Metab. 2007;6:348-351.
- 68. Patterson AD, et al. Metabolomics reveals attenuation of the SLC6A20 kidney transporter in nonhuman primate and mouse models of type 2diabetes mellitus. J Biol Chem. 2011;286:19511-19522.
- 69. Li F, et al. A comprehensive understandingof thioTEPA metabolism in the mouse using UPLC-ESI-QTOFMSbased metabolomics.Biochem Pharmacol. 2011;81:1043-1053.
- 70. PattersonAD, et al. Human urinarymetabolomic profile of PPARalpha induced fatty acid beta-oxidation. J ProteomeRes. 2009;8:4293-4300.
- 71. Li F, et al. Metabolomics reveals the metabolicmap of procainamide in humans and mice. Biochem Pharmacol. 2012;83:1435-1444.
- 72. Khaled MAR, et al. Biodegradation of used lubricating and diesel oils by new yeast strains Candida viswanathu kc-2011. African Journal of Biotechnology. 2012;11:14166-14174.
- 73. Wu TY, et al. A holistic approach to managing Palm Oil Mill Effluent (POME): Biotechnology advances in the sustainable re use of POME. Biotechnology Advances. 2009;27:40-52.
- 74. Cheng J, et al. POME treatment using a two stage microbial fuel cell system Integrated with immobilized biological aerated filters. Bioresource Technology. 2010;101:2729-2734.
- 75. Lal B and Khanna S. Degradation of crude oil by Acinetobactercalcoaceticus and Alcaligenesodorans. J ApplBacteriol. 1996;81:355-362.
- 76. Saadoun I. Isolation and characterisation of bacteria from crude petroleum oil contaminated soil and their potential to degrade diesel fuel. J Basic Microbiol. 2002;42:420-428.
- 77. Urum K, et al. Optimum conditions for washing of crude oil-contaminated soil with biosurfactant solutions. Process Safety and Environm Protect. 2003;81:203-209.
- 78. Andrä J, et al. Endotoxin-like properties of a rhamnolipidexotoxin from Burkholderia (Pseudomonas) plantarii: immunecell stimulation and biophysical characterization. BiolChem. 2006;301:310-387.
- 79. PiddingtonCS, et al. Sequence and molecular characterization of aDNA region encoding the dibenzothiophene desulfurization operon of Rhodococcus sp. strain IGTS8.Appl. Environ. Microbiol. 1995;61:468-475.
- 80. Librando V and Pappalardo M. In silico bioremediation of polycyclic aromatic hydrocarbon: a frontier in environmental chemistry. J Mol Graph Model. 2013;44: 1-8.
- 81. Pointing SB. Feasibility of bioremediation by white-rot fungi. Appl Microbiol Biotechnol. 2001;57:20-33.
- 82. Pickard MA, et al. Polycyclic aromatic hydrocarbon metabolism by white rot fungi and oxidation by Coriolopsisgallica UAMH 8260 laccase. Appl Environ Microbiol. 1999;65:3805-3809.
- 83. Samanta SK, et al. Polycyclic aromatic hydrocarbons: environmental pollution and bioremediation. Trends Biotechnol 2002;20:243-248.
- 84. Wong JW, et al. Pig manure as a co-composting material for biodegradation of PAH-contaminated soil. Environ Technol. 2002;23: 15-26.
- 85. Agamuthu P, et al. Phytoremediation of soil contaminated with used lubricating oil using Jatrophacurcas. J Hazard Mater. 2010;179:891-894.
- 86. Gateuille D, et al. Mass balance and decontamination times of Polycyclic Aromatic Hydrocarbons in rural nested catchments of an early industrialized region (Seine River basin, France). Sci Total Environ. 2014;470:608-617.
- 87. Johnsen AR, et al. Principles of microbial PAH-degradation in soil. Environ Pollut. 2005;133:71-84.
- 88. Sanders M, et al. Origin and distribution of polycyclic aromatic hydrocarbons in surficial sediments from the savannah river. Arch Environ Contam Toxicol. 2002;43:438-448.
- 89. Tang L, et al. Contamination of polycyclic aromatic hydrocarbons (PAHs) in urban soils in Beijing, China. Environ Int. 2005;31:822-828.
- 90. Hedlund BP, et al. Polycyclic aromatic hydrocarbon degradation by a new marine bacterium, Neptunomonas naphthovorans gen. nov., sp. nov. Appl Environ Microbiol. 1999;65:251-259.
- 91. Yakimov MM, et al. Alcanivorax borkumemsisgen. nov., sp. nov., a new hydrocarbon-degrading and surfactant producing marine bacterium. Int J Syst Bacteriol. 1998;48:339-348.
- 92. Chikere CB, et al. Biorector-based bioremediation of hydrocarbon.-polluted Niger Delta marine sediment, Nigeria. 3 biotech. 2012;2:53-66.

- 93. Golyshin PN, et al. Oleiphilaceae fam nov., to include Oleiphilus messinensis gen. nov., sp. nov., a novel marine bacterium that obligatory utilizes hydrocarbons. Int J SystEvol Microbiol. 2002;52:901-911
- 94. Yakimov MM, et al. Oleispira antarctica gen. nov., sp. nov., a novel hydrocarbon- oclastic marine bacterium isolated from Antarctic coastal seawater. Int J Syst Evol Microbiol 2003;53:779-785.
- 95. Garcia MT, et al. Catabolic versatility of aromatic compound-degrading halophilic bacteria. FEMS microbiol ecol. 2005;54:97-109.
- 96. Dastgheib SM, et al. Biodegradation of polycyclic aromatic hydrocarbons by a halophilic microbial consortium. Appl Microbiol Biotechnol. 2012;95:789-798.
- 97. Said B, et al. Characterization of aerobic polycyclic aromatic hydrocarbon- degrading bacteria from Bizerte lagoon sediments Tunisia. J Appl Microbiol. 2007;107: 987-997.
- 98. Chikere CB and Azubuike CC. Catechol 2, 3-dioxygenase screening in putative hydrocarbon utilizing bacteria. Int Res J Microbiol. 2013;4:1-6.
- 99. Okore C, et al. Isolation and Characterization of Biosurfactants Producing Bacteria from Oil Polluted Soil. J Nat Sci Res. 2013;3:2225-0921.
- 100. McGenity TJ, et al. Marine crude-oil biodegradation: a central role for interspecies interactions. Aquat Biosyst. 2012;8:10.