Silicon's Role in Improving Plant Salinity Tolerance

Yuchen Xia, Lecheng Liu, Junliang Yin* and Yongxing Zhu*

Hubei Key Laboratory of Waterlogging Disaster and Agricultural Use of Wetland /College of Agriculture/ College of Horticulture and Gardening Yangtze University, Jingzhou, Hubei 434025, China College of Agricultutr, Yangtze University, Jingzhou, Hubei 434025, China

Mini Review

ABSTRACT

Received: 28/03/2018 **Accepted:** 03/04/2018 **Published:** 07/04/2018

*For Correspondence

Yongxing Zhu, College of Horticulture and Gardening, Yangtze University, Jingzhou, China. Tel: 18710318976.

Junliang Yin, College of Agricultutr, Yangtze University, Jingzhou, Hubei 434025, China.

E-mail: yongxingzhu@yangtzeu.edu.cn yinjunliang@nwafu@edu.cn

Keywords: Silicon, Sat stress, Plant water status, Antioxidant defense

Salinization is a main abiotic stress that affects crop growth and productivity all over the world, thus how to improve the resistance levels of crops to salinity has attracted worldwide attention. Silicon (Si) is the second most prevalent element in the earth's crust. When the solution pH is below 9, silicon is absorbed by higher plants in the form of silicic acid $[Si(OH)_4]$. Although silicon has not been considered as essential element for higher plant, it is generally considered to be a 'beneficial element'. A number of studies have shown that silicon can increase plant resistance to multiple abiotic and biotic stresses, including drought, salt stress, heavy metal toxicity, freezing, and plant diseases.

INTRODUCTION

Salinization is a main abiotic stress that affects crop growth and productivity all over the world, thus how to improve the resistance levels of crops to salinity has attracted worldwide attention ^[1]. Silicon (Si) is the second most prevalent element in the earth's crust. When the solution pH is below 9, silicon is absorbed by higher plants in the form of silicic acid $[Si(OH)_4]^{[2]}$. Although silicon has not been considered as essential element for higher plant, it is generally considered to be a 'beneficial element' ^[3]. A number of studies have shown that silicon can increase plant resistance to multiple abiotic and biotic stresses, including drought, salt stress, heavy metal toxicity, freezing, and plant diseases ^[2-3]. In early years, studies suggested that silicon mainly exerts its protective action via the formation of a physical barrier by precipitating as SiO₂ and being incorporated into biological structures ^[2]. With the in-depth research, more and more studies suggest that silicon also active involve in physiological and biochemical processes in plants ^[4,5].

One important role of silicon is its alleviation effect of salt stress in plants, which has been reported in various species, including rice, barley, wheat, cucumber, tomato, and *Cicer arietinum* ^[6-11]. Till now, investigations about the exact modulating mechanism(s) of silicon to plant physiology have been reported at both physiological and molecular (e.g. genomic and proteomic) levels, which significantly advanced our knowledge about the silicon in plants.

Three different modes of silicon uptake have been proposed for plants, that is, active, passive, and rejective uptake. Silicon transporters play important roles in silicon uptake and have been identified in both monocotyledon (e.g. rice, barley, maize, wheat) and dicot (e.g. pumpkin, cucumber) ^[12-21]. The ability of plant to take up silicon may affect plant stress resistance ability ^[22]. Therefore, more work is needed to investigate the silicon uptake and distribution in different plant species.

Salt stresses usually hinder plants growth in two ways: (i) osmotic stress that limits the water availability for plants and affects water status; and (ii) ion toxicity that disturbs essential biochemical reactions through impairing enzyme activities and protein functions ^[23-24]. Most previous studies have focused on the mediation effect of silicon onion imbalance under salt stress. To be specific, silicon application can decrease Na⁺ uptake and its transport from roots to shoots and makes Na⁺ being more evenly distributed over the whole root section ^[25]. Besides, silicon affects the uptake of some other essential nutrients (e.g. Ca, K,

and N) to alleviate competition between salt ions and other essential nutrients uptake ^[2]. Most recently, studies suggested that polyamines may participate in the alleviation effect of silicon under salt stress through regulating Na⁺/K⁺ ratio ^[26].

DISCUSSION

Recently, more and more researchers showed that, under salt stress, silicon could improve plant water status through enhancing root water uptake and its transport to leaves, which mitigate ion toxicity by a dilution effect. In wheat, the alleviative effects of silicon have been found to be more pronounced in the osmotic stress phase than ion toxicity phase ^[23]. Besides, siliconmediated up-regulating of aquaporin gene expression and silicon-induced accumulation of compatible solutes like soluble sugars play important roles in increasing water uptake ^[5,26].

Salt stress leads to the formation of reactive oxygen species (ROS) that seriously disrupt normal metabolism ^[24]. Silicon application alleviates oxidative stress by regulating the antioxidant defense and decreasing the production of ROS ^[9]. Meanwhile, silicon could ameliorate the damage of photosynthetic apparatus and pigment induced by salt stress and contributes to the improvement of the photosynthetic performance ^[27]. Salt stress inhibits photosynthesis through causing the accumulation of photosynthetic assimilates in the leaves and decreasing assimilates export to the roots. In cucumber, silicon has been reported to decrease the soluble sugar levels in leaves and increased starch content in the roots, which could alleviate photosynthetic feedback repression and provides more energy storage in the roots under salt stress condition ^[4]. However, the mechanisms by which silicon alleviates salinity stress via regulating carbohydrate metabolism need a deeper investigation in different species.

The molecular mechanism for silicon-mediated salt tolerance is still not very clear. Omics-based technologies, including transcriptome and proteome, provide a powerful tool to understand the mechanisms by which silicon alleviates environmental stresses at the molecular level. Transcriptome and proteome studies reveal that silicon could regulate the response of plants against salt stress through modulating the expression of transcription factors and hormone-related genes and the translation of associated protein ^[28,29]. Besides, there may be a crosstalk between silicon and signaling molecules including ethylene (ET), salicylic acid (SA), and polyamines (PAs) ^[30,32]. These researches empowered us the insight understanding about the alleviation mechanisms of silicon to environmental salinity stress, both at the physiological and molecular level.

CONCLUSION

In the future, the regulation effect of silicon on polyamines metabolisms and aquaporin expression need to be further experimentally tested. Sugars can function as compatible solutes, immediate substrates for intermediary metabolism, as well as signaling molecules in controlling metabolism, stress resistance, growth, and development in plants ^[24,33]. Thus further experiments are needed to reveal the specific mechanisms that silicon regulating carbohydrate metabolism. Meanwhile, molecular mechanisms of the alleviation effect of silicon under environmental stresses still needs to be investigated in more detail at the genomic, transcriptomic, epigenetics, proteomic, and metabolomic levels.

ACKNOWLEDGEMENTS

This work was supported by the National Natural Science Foundation of China (No. 31701911), the Open Project Program of Engineering Research Center of Ecology and Agricultural Use of Wetland, Ministry of Education (No. KF201707).

REFERENCES

- 1. Guo Z, et al. Exogenously applied poly-γ-glutamic acid alleviates salt stress in wheat seedlings by modulating ion balance and the antioxidant system. Environ Sci Pollut R. 2017;24:6592-6598.
- 2. Zhu YX and Gong HJ. Beneficial effects of silicon on salt and drought tolerance in plants. Agron Sustain Dev. 2014;34:455-472.
- 3. Luyckx M, et al. Silicon and plants: current knowledge and technological perspectives. Front Plant Sci. 2017;8:411.
- 4. Zhu YX, et al. The regulatory role of silicon on carbohydrate metabolism in *Cucumis sativus* L. under salt stress. Plant Soil. 2016;406:231-249.
- 5. Zhu YX, et al. Silicon improves salt tolerance by increasing root water uptake in *Cucumis sativus* L. Plant Cell Rep. 2015;34:1629-1646.
- 6. Gong HJ, et al. Silicon deposition in the root reduces sodium uptake in rice (*Oryza sativa* L.) seedlings by reducing bypass flow .Plant Cell Environ. 2006;29:1970-1979.
- 7. Liang Y, et al. Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of saltstressed barley (*Hordeum vulgare* L.). J Plant Physiol. 2003;160:1157-1164.
- 8. Tuna AL, et al. Silicon improves salinity tolerance in wheat plants. Environ Exp Bot. 2008;62:10-16.
- 9. Zhu Z, et al. Silicon alleviates salt stress and increases antioxidant enzymes activity in leaves of salt-stressed cucumber (*Cucumis sativus* L.). Plant Sci. 2004;167:527-533.

RRJBS | Volume 7 | Issue 2 | April-June 2018

- 10. Li H, et al. Beneficial effects of silicon in alleviating salinity stress of tomato seedlings grown under sand culture. Acta Physiol Plant. 2015;37(4).
- 11. Garg N and Bhandari P. Silicon nutrition and mycorrhizal inoculations improve growth, nutrient status, K⁺/Na⁺ ratio and yield of *Cicer arietinum* L. genotypes under salinity stress. Plant Growth Regul. 2016;78(3):371-387.
- 12. Ma JF and Yamaji N. Functions and transport of silicon in plants. Cell Mol Life Sci. 2008;65:3049-3057.
- 13. Ma JF, et al. A silicon transporter in rice. Nature. 2006;440:688-691.
- 14. Ma JF, et al. Genotypic difference in silicon uptake and expression of silicon transporter genes in rice. Plant Physiol. 2007;145:919-924.
- 15. Mitani N, et al. Identification and characterization of maize and barley Lsi2-like silicon efflux transporters reveals a distinct silicon uptake system from that in rice. Plant Cell. 2009;21: 2133-2142.
- 16. Chiba Y, et al. HvLsi1 is a silicon influx transporter in barley. Plant J. 2009;57:810.
- 17. Yamaji N, et al. Functional characterization of a silicon transporter gene implicated in silicon distribution in barley. Plant Physiol. 2012;160:1491-1497.
- 18. Mitani N, et al. Identification of maize silicon influx transporters. Plant Cell Physiol. 2009;50:5-12.
- 19. Montpetit J, et al. Cloning, functional characterization and heterologous expression of *TaLsi1*, a wheat silicon transporter gene. Plant Mol Biol. 2012;79:35-46.
- 20. Mitani N, et al. Isolation and functional characterization of an influx silicon transporter in two pumpkin cultivars contrasting in silicon accumulation. Plant J. 2011;66:231-240.
- 21. Sun H, et al. Isolation and functional characterization of CsLsi1, a silicon transporter gene in Cucumis sativus. Physiol Plant. 2017;159: 201-214.
- 22. Rios JJ, et al. Silicon-mediated improvement in plant salinity tolerance: the role of aquaporins. Front Plant Sci. 2017;8:948.
- 23. Chen D, et al. Silicon increases salt tolerance by influencing the two-phase growth response to salinity in wheat (*Triticum aestivum* L.). Acta Physiol Plant. 2014;36:2531-2535.
- 24. Parida AK and Das AB. Salt tolerance and salinity effects on plants: a review. Ecotox Environ Saf. 2005;60:324-349.
- 25. Liang Y. Effects of silicon on enzyme activity and sodium, potassium and calcium concentration in barley under salt stress .Plant and Soil. 1999;209:217-224.
- 26. Yin L, et al. Silicon-mediated changes in polyamines participate in silicon-induced salt tolerance in Sorghum bicolor L. Plant Cell Environ. 2015;39:245-258.
- 27. Liang YC. Effects of Si on leaf ultrastructure, chlorophyll content and photosynthetic activity in barley under salt stress. Pedosphere. 1998;8:289-296.
- 28. Holz S, et al. Initial studies on cucumber transcriptome analysis under silicon treatment. Silicon. 2015;7:1-5.
- 29. Muneer S and Jeong BR. Proteomic analysis of salt-stress responsive proteins in roots of tomato (*Lycopersicon esculentum* L.) plants towards silicon efficiency. Plant Growth Regul. 2015;77:133-146.
- 30. Kim YH, et al. Silicon application to rice root zone influenced the phytohormonal and antioxidant responses under salinity stress. J Plant Growth Regul. 2014;33:137-149.
- 31. Liang XL, et al. Silicon does not mitigate cell death in cultured tobacco BY-2 cells subjected to salinity without ethylene emission. Plant Cell Rep. 2015;34:331-343.
- 32. Yin LN, et al. Silicon-mediated changes in polyamines participate in silicon-induced salt tolerance in Sorghum bicolor L.. Plant, Cell Environ. 2016;39:245-258.
- 33. Rolland F, et al. Sugar sensing and signaling in plants: conserved and novel mechanisms. Annu Rev Plant Biol. 2006;57:675-709.